Research Article Assessing the diversity of macroinvertebrates communities and their relationship with environmental factors in the Persian Gulf and the Gulf of Oman

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Abstract

Marine benthic macrofauna plays an important role in ecosystem processes in the marine benthic zone. This study aims to investigate the biodiversity pattern and identifying benthic macroinvertebrates in the coastal zones of the Persian Gulf and the Gulf of Oman. Two sampling periods were performed in this study; the first: from November 2018 to January 2019 and the second: from December 2019 to January 2020. Sampling was carried out at 19 sites with three replicates (171 samples). A total of 164 species belonging to 85 families and 31 orders were identified. The most abundant families were Ampeliscidae and Apseudidae with mean values of 19.5 ± 42 and 11.1 ± 18 in m², respectively. The most abundant class were Malacostraca and Polychaeta. The family Nephtyidae with eight identified species was the most in all stations. Amphipholis squamata, Byblis sp., Capitella *capitate*, and *Amphioplus* sp. were the significant species that contributed to the differentiation between stations. Furthermore, according to CCA analysis, dissolved oxygen (DO) showed a strong direct relationship with Callianassidae, Anthuridae, Paraonidae, Maeridae, and Corophiidae. Also, a strong indirect relationship was found between Haminoeidae and DO. Turbidity showed a strong direct relationship with Flabelligeridae and as well as weak direct with Cirratulidae. Furthermore, chlorophyll-a showed a strong direct relationship with Nereididae, and other parameters showed a medium direct relationship with Maldanidae and Haminoeidae. The Composition of bed sediment was also showed a strong relationship with macroinvertebrates communities in the study area. The diverse health status was not totally in a good condition and needs to be improved. These findings could be applied in further studies for habitat management and monitoring programs.

Keywords: Macroinvertebrate assemblages, Diversity index, Persian Gulf, Gulf of Oman

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Introduction

Marine benthic macrofauna plays an important role in ecosystem processes such as dispersion and burial, pollutant metabolism, nutrient cycling. and production (Snelgrove, secondary 1998). Although biomass and diversity of benthic macroinvertebrates represent a temporal and seasonal pattern, they are employed as aquatic ecosystem health indicators (Margues et al., 2009). Benthic macroinvertebrates react to anthropogenic disturbances and natural changes such as temperature, pH, DO, and pollution in their habitats in different ways (Saghali et al., 2013; Gerami et al., 2016). Climate change affected the marine ecosystem and the complexity of the food webs (Schofield et al., 2010). Increased global average temperature, declining aquatic oxygen concentrations, and changes in wind strength are the main factors that impact the behavior of ocean currents behavior. leading to changes in the distribution and abundance of marine ecosystems (Hoegh-Guldberg and Bruno, 2010). Fluctuation in the diversity pattern of marine macroinvertebrates as marine food web secondary production can affect trophic networks and cause complex changes trophic to relationships (Nordström and Bonsdorff, 2017).

Environmental variables at multiple scales affected the diversity and density of macroinvertebrates communities differently (Townsend *et al.*, 1983). Therefore, it is essential to study their relationship spatially and temporally to develop our understanding of marine ecosystems. A better understanding of macroinvertebrates-environmental variables relation increases the accuracy and precision of macroinvertebratebased bioassessment (Li et al., 2012). Many studies have been reported the effect of environmental variables on marine benthic communities around the world (Teske and Wooldridge, 2004; Gamboa-García et al., 2018; Xu et al., 2019; Wu et al., 2019; Alsaffar et al., 2020; Patang and Soegianto, 2020). However, data about the status of marine macroinvertebrate communities of the Persian Gulf is limited especially species level at the and scant information is available to address how environmental variables influence macroinvertebrate communities in the Persian Gulf. Gerami et al. (2016) studied spatial and temporal variation of macroinvertebrates in the shrimp fishing grounds of the Hormozgan waters and reported the seasonal variation of 19 macroinvertebrate orders. Hajializadeh et al. (2020)investigated microbenthic species diversity and their biological trait mangrove-associated patterns in habitats of Hormozgan province waters and reported total of 122 macrofauna taxa in their study. They declared that deposit-feeding, crawlers, medium-size, and free-living were the dominant trait modalities in all habitats. Azizi et al. (2020) studied macrobenthic fauna in rocky habitats of Bushehr province coastal waters and reported total of 66 taxa in all studied stations. They indicated that mollusks were abundant in all three sites and Portunid crabs and Polychaets were the lowest abundant groups. Kosari *et al.* (2021) conducted a study on dentification of macrobenthos in the Yekshabe Estuary in the Persian Gulf and reported 80 species. They found that Polychaeta was the dominant group in all stations and there was no significant difference between species richness in all stations.

The objectives of this study were (i) to investigate the influence of the most important environmental variables on macroinvertebrates; (ii) to identify and evaluate the diversity of macroinvertebrates, and (iii) assess the relationship between the composition of bed sediment and macroinvertebrates communities. The results of this study could be used in the monitoring of marine species habitat health and assessing the productivity of the marine food web.

Materials and methods

Two sampling periods were performed in this study. The first period started from November 2018 to January 2019, and the second period from December 2019 to January 2020. Sampling started form station one in North-West coastal waters of the Hormozgan Province and continued to station 19 to the east up to coastal waters of Sistan and Balouchistan Province (Fig. 1).



Figure 1: Locations of sampling sites in the Persian Gulf and the Gulf of Oman, Iran (2018-2020).

The vessel R / V Ferdows-1 was used which was equipped with GPS. Sampling stations were selected randomly to cover most coastal waters of Hormozgan province coastal waters. Sampling depths were varied between 10 to 50 meters. Three replicate samples of macroinvertebrates per station (171 samples) were covered in the study period. Sampling was carried out by Van Veen grab with a 0.0256 m^2 cross section. Physicochemical properties of water were measured by CTD including Temperature (°C), Conductivity (ms/cm), Salinity (ppm), DO (g/L), pH, Chlorophyll-a (mg/L), and Turbidity. Samples were sieved through 0.5 mm mesh. The remained samples were fixed with 97% ethanol and transferred to the laboratory. Most illustrated keys such as Naderloo (2017); Al- Yamani *et al.* (2012);

Debruyne (2003); Fauchald (1997); Bosch *et al.* (1995) and Sterrer (1986) were used to identify macroinvertebrates up to species level. In order to compare biodiversity between sampling sites and the health status, the Shannon-Wiener Index (Shannon and Wiener, 1948) was used. The algorithm was:

$$H' = -\sum P_i log_2 P_i$$

Where: P_i is the proportion of individuals belonging to species i in the sample. The Shannon-Wiener index usually varies between 0 to 5 (Marques *et al.*, 2009). However, Molvær *et al.* (1997) introduced a classification table for assessing habitat health status based on the Shannon-Wiener index (Table 1).

0	es considered as a of Shannon-Wiener				
Index value					
Shannon-Wiener Shannon-Wiene					
index	value				
High status	>4 bits/individual				
Good status	3-4 bits/individual				
Moderate status	2-3 bits/ individual				
Poor status	1-2 bits/ individual				
Bad status	0-1 bit / individual				

Margalef Index (Margalef, 1969) was also employed to quantify the diversity of identified species in the study area. The formula was:

$$D = \frac{(S-1)}{Log_e N}$$

Where S = number of species and N = total number of individuals. Margalef Index values below four identified as typical of polluted areas (Ros and Cardell, 1991). The Composition of bed

sediment aggregation was measured through a method explained bv Buchannan (1984) and the percent of total organic matter was also calculated by method Holme and McIntyre (1984). In order to assess the similarity between and founding stations significant discriminant species SIMPER analysis Significant preformed. differences between stations were analyzed through the PERMANOVA method. Canonical correspondence analysis (CCA) was applied to extract major gradients among combinations of macrobenthic assemblages and environmental variables. Principal component analysis (PCA) was performed to find the main component before CCA analysis. The analysis of similarities (ANOSIM) was used to examine the spatial patterns of macroinvertebrate assemblages. For parametric analyses, the abundance data were square-root transformed to reduce heteroscedasticity before running the similarity matrix. All the analyses were carried out with R statistical packages and a vegan package was used to run analysis (Oksanen et al., 2015).

Results

The results showed that a total of 164 species were identified in all sites, belonging to 85 families and 31 orders. The most abundant families were Ampeliscidae and Apseudidae with mean values of 19.5 ± 42 (n/m²) and 11.1 ± 18 numbers in square meter in all stations, respectively. In addition, the most abundant class was Malacostraca and Polychaeta, respectively (Fig. 2).



Figure 2: Mean abundance of benthic macroinvertebrates of the Persian Gulf and the Gulf of Oman (2018-2020).

The most diverse family was all Nephtyidae with 8 identified species in

all stations (Table 2).

 Table 2: Identified macroinvertebrates in all sampling sites of the Persian Gulf and the Gulf of Oman (2018-2020)

Species	Family	Suborder	Order	Subclass	Class	Phylum
Magelona longicornis Johnson, 1901	Magelonidae		Canalipalpata	Polychaeta incertae sedis	Polychaeta	Annelida
Owenia fusiformis	0					
Delle Chiaje, 1844	Oweniidae					
Maldane cristata Treadwell, 1923	Maldanidae			Sedentaria		
Praxillella gracilis						
(M. Sars, 1861)						
Clymenella torquata (Leidy, 1855)						
Aricidea (Aricidea) wassi Pettibone, 1965	Paraonidae					
Aricidea longobranchiata Day, 1961						
Aricidea (Acmira) assimilis						
Tebble, 1959						
Leodamas chevalieri (Fauvel, 1902)	Orbiniidae					
<i>Orbiniella</i> sp.						
Day, 1954						
<i>Orbinia</i> sp. Quatrefages, 1866						
<i>Ophelia</i> sp.	Opheliidae					
Savigny, 1822	Opnemidae					
Ophelina acuminata						
Örsted, 1843						
Ophelina cylindricaudata						
(Hansen, 1879)						
Capitella capitata (Fabricius, 1780)	Capitellidae					
Capitella sp.1						
Blainville, 1828						

Species	Family	Suborder	Order	Subclass	Class	Phylum
Capitella sp.2						
Blainville, 1828						
Rashgua rubrocincta Wesenberg-Lund, 1949						
Cossura laeviseta Hartmann-Schröder, 1962	Cossuridae					
Cossura longocirrata Webster & Benedict, 1887						
Cossura pygodactylata Jones, 1956						
Cossura sp.						
Webster & Benedict, 1887						
Cirrophorus sp.	Paraonidae					
Ehlers, 1908	Taraomuae					
Poecilochaetus serpens Allen, 1904	Poecilochaetidae	Spioniformia	Spionida			
Paraprionospio pinnata (Ehlers, 1901)	Spionidae					
Prionospio rotalis Mohammad, 1970						
Scolelepis bonnieri (Mesnil, 1896)						
Genus sp.1						
Acrocirrus sp.	Acrocirridae	Cirratuliformia	Terebellida			
Grube, 1873	Teroenridue	Ciriaanonina	Terebellidu			
Amage auricula Malmgren, 1866	Ampharetidae					
Amphicteis sp.						
Grube, 1850						
Samythopsis grubei McIntosh, 1885						
Chaetozone corona Berkeley & Berkeley, 1941	Cirratulidae					
<i>Chaetozone</i> sp. Malmgren, 1867						
Cirriformia tentaculata (Montagu, 1808)						
Tharyx sp.						
Webster & Benedict, 1887						
Brada incrustata	Flabelligeridae					
Støp-Bowitz, 1948	- mooningeridue					
<i>Brada</i> sp. Stimpson, 1853						
Diplocirrus glaucus (Malmgren, 1867)						
Pherusa plumosa (Müller, 1776)						
<i>Sternaspis scutata</i> (Ranzani, 1817)	Sternaspidae					
Pista brevibranchiata Moore, 1923	Terebellidae	Terebelliformia				
Pista sp.						
Malmgren, 1866 <i>Terebella</i> sp.						
Linnaeus, 1767						

Species	Family	Suborder	Order	Subclass	Class	Phylum
Loimia sp.						
Malmgren, 1866						
Terebellides stroemii	Taicheberget					
Sars, 1835	Trichobranchidae					
Branchiomma sp. Kölliker, 1858	Trichobranchidae		Sabellida			
Parasabella aberrans (Augener, 1926)						
Sabella sp.						
Linnaeus, 1767						
Bhawania goodie	~		~ ~ ~ ~ ~			
Webster, 1884 Chrysopetalum sp.	Chrysopetalidae	Nereidiformia	Phyllodocida	Errantia		
Ehlers, 1864						
Syllis cornuta						
Rathke, 1843	Chrysopetalidae					
Hediste diversicolor						
(O.F. Müller, 1776)	Nereididae					
Leonnates indicus Kinberg, 1865						
Perinereis aibuhitensis						
(Grube, 1878)						
Perinereis						
nigropunctata (Horst ,1889)						
Perinereis sp.						
Kinberg, 1865						
Ancistrosyllis sp. McIntosh, 1878	Pilargidae					
Sigambra tentaculata (Treadwell, 1941)						
Sirsoe methanicola (Desbruyeres & Toulmond, 1998)	Hesionidae					
Glycera amboinensis McIntosh, 1885	Glyceridae	Glyceriformia				
Glycera capitata						
Örsted, 1843						
Glycera sp1.						
Lamarck, 1818						
Glycera sp2.						
Lamarck, 1818						
Glycera tridactyla Schmarda, 1861						
Glycera unicornis Lamarck, 1818						
Glycinde bonhourei Gravier, 1904	Goniadidae					
Glycinde sp.						
Müller, 1858						
Aglaophamus sp.		Phyllodocida				
Kinberg, 1866	Nephtyidae	incertae sedis				
Micronephtys sphaerocirrata (Wesenberg-Lund, 1949)						
Nephtys californiensis Hartman, 1938						
Nephtys cirrosa						
Ehlers, 1868						

Species	Family	Suborder	Order	Subclass	Class	Phylum
Micronephthys cornuta (Berkeley & Berkeley, 1945)						
Nephtys glabra						
Hartman, 1950						
Nephtys sp.						
Cuvier, 1817						
<i>Nephtys tulearensis</i> Fauvel, 1919						
Eteone sp.	Phyllodocidae	Phyllodociformia				
Savigny, 1822		,				
Harmothoe dictyophora (Grube, 1878)	Polynoidae	Aphroditiformia				
Harmothoe sp.						
Kinberg, 1856						
Euthalenessa sp. Darboux, 1899	Sigalionidae					
Leodice laticeps (Ehlers, 1868)	Eunicidae		Eunicida			
Scoletoma laurentiana (Grube, 1863)	Lumbrineridae					
Lumbrineris bifurcata McIntosh, 1885						
Ninoe pulchra Wesenberg-Lund, 1949						
Diopatra chiliensis	0					
Quatrefages, 1866	Onuphidae					
Diopatra cuprea						
(Bosc, 1802)						
Diopatra micrura						
Pires, Paxton, Quintino & Rodrigues, 2010						
Diopatra sp.						
Audouin & Milne Edwards, 1833						
Chloeia sp.	Amphinomidae		Amphinomida			
Lamarck, 1818			· · · · ·			
Linopherus hirsute (Wesenberg-Lund, 1949)						
Genus sp.1	Leptoplanidae	Acotylea	Polycladida			Platyhelminthe
Prosthiostomum sp. Quatrefages, 1845	Prosthiostomidae	Cotylea	-			-
Pseudoceros sp. Lang, 1884	Pseudocerotidae					
Ampithoe sp.						
Leach, 1814	Ampithoidae	Senticaudata	Amphipoda	Eumalacostraca	Malacostraca	Arthropoda
Cheiriphotis sp.						
Walker, 1904	Corophiidae					
Microphotis blachei Ruffo, 1952	Photidae					
Elasmopus pectenicrus (Spence Bate, 1862)	Maeridae					
Ceradocus sp.						
Costa, 1853						
Maera sp.						
Leach, 1814						

Species	Family	Suborder	Order	Subclass	Class	Phylum
Maera schieckei						
Karaman & Ruffo, 1971						
<i>Melita</i> sp. Leach, 1814	Melitidae					
<i>Platorchestia monodi</i> (Mateus, Mateus & Afonso, 1986)	Talitridae					
<i>Cyproidea</i> sp. Haswell, 1880	Cyproideidae	Amphilochidea				
Orchomene sp. Boeck, 1871	Tryphosidae					
Byblis sp.	Ampeliscidae					
Boeck, 1871 Eobrolgus spinosus	Phoxocephalidae					
(Holmes, 1905) <i>Penaeus</i> sp.	Penaeidae	Dendrobranchiata	Decapoda			
Fabricius, 1798						
Alpheus estuariensis Christoffersen, 1984	Alpheidae	Pleocyemata				
Alpheus lobidens						
De Haan, 1849 [in De Haan, 1833-1850]						
Athanas nitescens (Leach, 1814 [in Leach, 1813-1815])						
Athanas sp.						
Leach, 1814 [in Leach, 1813-1815]						
Callianassa sp.						
Leach, 1814 [in Leach, 1813-1815]	Callianassidae					
Myra pernix	Leucosiidae					
Galil, 2001	Leacosnade					
Philyra sagittifera (Alcock, 1896)						
<i>Heteropanope glabra</i> Stimpson, 1858	Pilumnidae					
Pilumnus longicornis Hilgendorf, 1879						
Pilumnus minutus						
De Haan, 1835 [in De Haan, 1833-1850]						
Charybdis (Goniohellenus) longicollis	Portunidae					
Leene, 1938						
Portunus segnis (Forskål, 1775)						
Apanthura sandalensis Stebbing, 1900	Anthuridae	Cymothoida	Isopoda			
Gnathia maxillaris (Montagu, 1804)	Gnathiidae					
Gnathia sp.						
Leach, 1814						
<i>Bodotria</i> sp.	Bodotriidae		Cumacea			
Goodsir, 1843	Bououndae		Cumacea			
Eocuma affine						

Species	Family	Suborder	Order	Subclass	Class	Phylum
Eocuma rosae						
Corbera & Galil, 2007						
Apseudes sp.	Apseudidae	Apseudomorpha	Tanaidacea			
Leach, 1814	ripsedardae	ripoeudomorphu	Tunnuureu			
Ctenapseudes chilkensis	Parapseudidae					
(Chilton, 1924) Squilla mantis						
(Linnaeus, 1758)	Squillidae	Unipeltata	Stomatopoda	Hoplocarida		
Genus sp.1	Acricoactinidae	Enthemonae	Actiniaria	Hexacorallia	Anthozoa	Cnidaria
Genus sp.2						
Amphioplus (Lymanella) hastatus (Ljungman, 1867)	Amphiuridae	Gnathophiurina	Amphilepidida	Myophiuroidea	Ophiuroidea	Echinodermta
Amphioplus sp.						
Verrill, 1899						
Amphipholis squamata (Delle Chiaje, 1828)						
Clypeaster humilis	Clypeasteridae	Clypeasterina	Clypeasteroida	Euechinoidea	Echinoidea	
(Leske, 1778)			,,			
Cardiomya sp.	Cuspidariidae					
A. Adams, 1864 Hiatula rosea						
(Gmelin, 1791)	Psammobiidae		Cardiida	Autobranchia	Bivalvia	Mollusca
<i>Pharaonella wallaceae</i> (A. E. Salisbury, 1934)	Tellinidae					
Serratina capsoides (Lamarck, 1818)						
Tellina sp.						
Linnaeus, 1758						
Hanleyanus vestalis (Hanley, 1844)						
Irus carditoides	Veneridae		Venerida			
(Lamarck, 1818) Protapes gallus						
(Gmelin, 1791)						
Timoclea sp.						
T. Brown, 1827						
Lamellolucina dentifera (Jonas, 1846)	Lucinidae		Lucinida			
Corbula taitensis Lamarck, 1818	Corbulidae		Myida			
Arcuatula senhousia (Benson, 1842)	Mytilidae		Mytilida	Autobranchia		
Anadara ehrenbergi	Arcidae		Arcida			
(Dunker, 1868)						
Scissileda tropica (Melvill, 1897)	Yoldiidae		Nuculanida	Protobranchia		
Cylichna collyra Melvill, 1906	Cylichnidae		Cephalaspidea	Heterobranchia	Gastropoda	
Bakawan rotundata (A. Adams, 1850)	Haminoeidae					
Ellobium sp.						
Röding, 1798	Ellobiidae		Ellobiida			
Omalogyra atomus (Philippi, 1841)	Omalogyridae					
Linatella caudate (Gmelin, 1791)	Cymatiidae		Littorinimorpha	Caenogastropoda		

Species	Family	Suborder	Order	Subclass	Class	Phylum
Cheilea cicatricosa (Reeve, 1858)	Haminoeidae					
Hypermastus epiphanes (Melvill, 1897)	Eulimidae					
Pseudonoba sp.	Iravadiidae					
O. Boettger, 1902	iravauluae					
Natica vitellus	Naticidae					
(Linnaeus, 1758)	inalicidae					
Neverita didyma (Röding, 1798)						
Zafra selasphora (Melvill & Standen, 1901)	Columbellidae		Neogastropoda			
<i>Crassispira</i> sp. Swainson, 1840	Pseudomelatomidae					
<i>Ethminolia degregorii</i> (Caramagna, 1888)	Trochidae		Trochida	Vetigastropoda		
Antalis longitrorsa (Reeve, 1842)	Dentaliidae		Dentaliida		Scaphopoda	
Lingula sp.	Lingulidae		Lingulida		Lingulata	Brachiopoda
Bruguière, 1791	Lingundae		Lingunua		Lingulata	Бтастворода

Results of diversity indices are represented in Table 3. According to the findings, maximum Shannon-Wiener and Margalef indices of some species were observed in station 3 while the minimum was in station 15. In addition, Maximum density found in station 15. According to Table 1, the health status of station 3 was high while other stations were good except for stations 15, 16, 17, and 19. The health status of station 15 was distinguished as bad while 16, 17, and 19 were marked as poor status.

 Table 3: Diversity indices of benthic macroinvertebrates of the Persian Gulf and the Gulf of Oman (2018-2020)

Station	Shanon	Margalof	Mean density	number of species
1	3.33	6.58	5.0	34
2	3.77	9.26	4.35	50
3	4.10	11.65	4.64	67
4	3.42	9.75	7.7	60
5	3.60	8.20	4.7	43
6	3.23	9.16	7.92	57
7	3.74	9.14	5.57	53
8	3.53	9.05	5.87	53
9	3.81	10.0	7.78	63
10	3.32	6.87	5.06	37
11	3.65	8.80	5.23	50
12	3.40	7.93	6.31	46
13	3.68	8.16	5.06	49
14	3.62	8.15	5.41	46
15	1.86	2.42	30.0	15
16	2.61	3.46	4.48	16
17	2.34	4.49	25.49	31
18	3.29	4.54	12.97	57
19	2.62	3.57	5.14	17

SIMPER analysis

Results of SIMPER analysis revealed that there were significant differences between stations in composition and abundance of macroinvertebrates (p<0.05). In addition, *Amphipholis squamata*, *Byblis* sp., *Capitella capitate*, and *Amphioplus* sp. were the most effective species that caused differentiation between stations. sediment composition analysis are represented in Table 4. According to the results, maximum TOM found in station 9 and minimum in 20. The maximum amounts of sand and silt were found in stations 15 while minimum amounts were found in 7 and 4, respectively. The minimum and maximum amounts of clay were found in stations 20 and 14, respectively.

Results of total organic matter and

Table 4: Total organic matter and sediment composition analysis of sediments of the Persian Gulf and the Gulf of Oman (2018-2020)

and	i the Gull of Oman (2	(018-2020)		
Station	TOM (%)	Sand (%)	Silt (%)	Clay (%)
1	4.75	32.9	63.0	4.1
2	4.15	46.6	49.3	4.1
3	7.12	22.6	72.6	4.8
4	7.29	30.3	67.6	2.1
5	8.58	24.3	72.6	3.1
6	6.84	26.3	71.6	2.1
7	9.71	22.6	72.3	5.1
8	7.02	46.6	52.1	1.3
9	7.29	70.6	26.3	3.1
10	14.97	59.8	34.9	5.3
11	17.52	65.4	31.3	3.3
12	12.91	38.3	57.4	4.3
13	9.98	54.3	43.1	2.6
14	14.99	61.7	33.0	5.3
15	4.57	98.0	2.3	4.3
16	14.47	73.7	21.0	5.3
17	12.68	61.7	33.0	5.3
18	4.15	93.7	5.0	1.3
19	2.68	83.7	13.0	3.3

Physico-chemical properties

Results of Physico -Chemical properties of water are represented in Table 5. The minimum and maximum amounts of temperature were found in stations 17 and 3, respectively, while these values for salinity were in stations 2 and 15. Conductivity and pH were minimum in station 4. Minimum amounts of DO were found in station 12 while maximum chlorophyll-a was measured in station 1. Maximum and minimum of turbidity found in stations 11 and 3, respectively.

PCA and CCA analysis

PCA analysis revealed that among 85 identified families, 33 family were the most effective component for dimension reduction. The identified

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families were Apseudidae,	ilies were Ampeliscidae, Melitidae, eudidae, Prosthiostomidae,		Corophiidae, Amphinomidae,	Terebellidae, Amphiuridae,
Maeridae,	Bodotriidae,	Photidae,	Nereididae, Talitridae	, Ampharetidae,
Callianassida	ie,	Cirratulidae,	Maldanidae,	Clypeasteridae,
Anthuridae,		Cossuridae,	Nephtyidae, Lumbrine	ridae, Opheliidae
Flabelligerid	ae, Sabellidae,	Glyceridae,	and Capitellidae and t	here upon, these
Haminoeidae	, Hesionidae,	Lingulidae,	families were selec	cted for CCA
Paraonidae,	Alpheidae,	Spionidae,	analysis.	

 Table 5: The mean values of physiochemical properties of water of the Persian Gulf and the Gulf of Oman (2018-2020).

Station	Temperature (°C)	Conductivity (ms/cm)	Salinity (ppm)	DO (g/L)	pН	Chlorophyll-a (mg/L)	Turbidity
1	25.95	56.37	36.69	91.72	8.15	2.02	15.32
2	24.09	53.90	36.33	59.75	7.89	0.30	14.16
3	26.77	57.44	36.80	74.53	8.13	1.22	7.48
4	22.82	52.59	36.36	38.48	7.83	0.20	8.68
5	25.77	56.16	36.66	88.48	8.07	0.56	14.08
6	25.71	56.09	36.66	72.56	8.08	0.44	14.18
7	25.94	56.40	36.71	101.63	8.14	0.86	19.68
8	25.96	56.40	36.70	102.65	8.15	0.60	17.32
9	24.53	55.60	37.26	57.16	8.27	0.60	12.04
10	24.55	55.94	37.51	3.86	8.25	2.00	33.64
11	24.82	56.39	37.63	8.71	8.24	1.36	60.38
12	24.83	56.73	37.87	2.62	8.26	1.38	18.22
13	24.25	57.39	38.87	49.34	8.25	0.88	13.04
14	23.48	56.29	38.70	65.72	8.18	1.36	11.68
15	24.28	57.68	39.06	87.45	8.24	0.88	11.82
16	23.05	54.74	37.86	79.35	8.18	1.24	13.46
17	22.78	54.41	37.85	90.51	8.25	1.48	11.68
18	24.58	57.73	38.83	87.76	8.19	0.42	16.26
19	23.69	55.91	38.22	114.71	8.11	0.84	10.58

According to CCA results (Fig. 3), DO showed a strong direct relationship with Callianassidae, Anthuridae, Paraonidae, Maeridae, and Corophiidae. In addition, the strong indirect relationship was found between Haminoeidae and DO. Turbidity showed a strong direct

relationship with Flabelligeridae and a weak direct with Cirratulidae as well. Furthermore, chlorophyll-a showed a strong direct relationship with Nereididae, and other parameters showed a medium direct relationship with Maldanidae and Haminoeidae.



Figure 3: Results of CCA analysis between physiochemical properties of water and main macroinvertebrates families of the Persian Gulf and the Gulf of Oman (2018-2020). Note: Temp: Temperature, Ras: Conductivity, Shor: Salinity, O: DO, Cl: Chlorophyll-a, COD: Turbidity

According to CCA analysis (Fig. 4) and sediment composition results, clay was located in the center of the chart and did not show a remarkable relationship with microbenthic families. Although a weak indirect relationship with all families could be considered for clay, sand showed a significant strong relationship with Bodotriidae, Maeridae. and Melitidae directly and with Amphiuridae and Spionidae, indirectly. On the other hand, silt showed a significant direct relationship with Spionidae and Amphiuridae and indirect with Maeridae and Melitidae. Finally, TOM showed a strong direct relationship with Nereididae and weak direct with Photidae.



Figure 4: Results of CCA analysis between sediment composition, TOM, and main macroinvertebrates families of the Persian Gulf and the Gulf of Oman (2018-2020).

Discussion

Marine macroinvertebrates as secondary producers play an important role in feeding benthic animals in the ecosystem (Dolbeth et al., 2003). The Persian Gulf and the Gulf of Oman are considered high productive as (Owfi al., 2016), ecosystems et therefore biodiversity in these two regions is high. Understanding the biodervisty abundance and of macroinvertebrates species is a fundamental step towards achieving

sustainable management of ecosystems in this region. Our results indicated that in 4 stations in the studied area, we faced harsh environmental challenges for macroinvertebrates (Table 3). In addition, only one station showed high biodervisty status while others were near the border of moderate and good status (Table 3). Several researchers reported decreasing a trend in macroinvertebrates marine in ecosystems in different regions of the world (Schofield et al., 2010; Bumbeer et al., 2016; Wu et al., 2019). In addition, Lahijanzadeh et al. (2019) declared that concentrations of toxic elements such as Al, Fe, Cu, and Zn increased gradually in tissues and shells of benthic macroinvertebrates in the Persian Gulf. Therefore, the risk of pollutants threatens biodervisty of macroinvertebrates in the region and affects the food web. During this study, the number of 164 species belonging to and 31 orders 85 families of macroinvertebrates were identified in the Persian Gulf and the Gulf of Oman (Table 2). Dehghan Madiseh et al. (2012) sampled benthic invertebrates from the North-West of the Persian Gulf and reported 29 identified species. They were also noted that most sampling sites showed slightly polluted status through slightly AZTI's Marine Biotic Index. Furthermore, Gerami et al. (2016)sampled from macroinvertebrates communities in the Persian Gulf and their sampling sites had overlap with sampling sites in this study. However, 15 orders of macroinvertebrates containing all identified orders in their study, and the biodiversity health status of the Shannon-Wiener index was the poor status for all sites.

Spatial variation in the composition of macroinvertebrates communities in all sites was observed and SIMPER analysis showed that *Amphipholis squamata*, *Byblis* sp., *Capitella capitate* and *Amphioplus* sp. were the most effective species in differences between stations. This might indicate that these species tend to have more temporal or spatial variation in the region. A. and Amphioplus squamata sp. belonging to class Ophiuroidea are the most distributed species in the Persian Gulf (Fatemi and Stöhr, 2019) and accustomed to a huge variety of habitats such as boulders, sand, or muds (Boissin et al., 2008). Byblis is also an abundant genus of order Amphipoda that is distributed in all parts of the Persian Gulf and the Gulf of Oman (Mohammadi Dehcheshme al.. et 2016).

The relationship between macroinvertebrates families and environmental factors was estimated based on CCA analysis (Fig. 3). DO, turbidity, and chlorophyll-a were the dominant environmental factors that directly or indirectly affected the distribution and community composition of macroinvertebrates in the study area. Gerami et al. (2016) were also reported that DO and chlorophyll-a were the best variables for explaining changes in abundance over time. Although they did not analyze turbidity in their analysis. It is well known that oxygen availability is a major factor influencing the composition of macrobenthic communities. Indeed, oxygen is the first factor in macrobenthic respiration (Steel, 1980). Furthermore, the amount of oxygen production in water bodies has connection with the concentration of chlorophyll. Kuhnt et al. (2013) suggested that there is a speciesspecific relationship between benthic foraminiferal species and bottom-water oxygen content. However, benthic habitats are complex, and a variety of variables affect environmental the diversity and densitv of macroinvertebrates. The composition of bed sediments and sediment's food availability are significant factors that composition affect of macroinvertebrates communities (Bumbeer et al., 2016; Jun et al., 2016; Mehdipour et al., 2018). According to the results, sand, silt, and TOM showed relationship strong with main а macroinvertebrates families and affected their distribution. Several researchers reported the response of macrobenthic communities to sediment treatment (Doeg and Milledge, 1991; Suren and Jowett, 2001; Connolly and Pearson, 2007). Aliakbarian et al. (2020) reported the relation between the type of sediment and macrobenthic species in the southeast of the Caspian Sea. Although their results were based on RDA analysis instead of CCA, they also reported that sand and slit were most related in comparison with clay. According to figure 4, some families of identified macroinvertebrates showed relation with sediment composition. Data for comparison is scarce in literature. However, the results of this study were in accordance with researchers around the world. Anderson et al. (2013) found that there is a relationship between seafloor structure and infaunal biodiversity and reported that Polychaeta especially the Maeridae family has a correlation with sand in the studied area. Furthermore, Toba and Sate (2013) stated that the Nereididae family is a Filter-feeder benthic

macroinvertebrates that has a positive correlation with TOM of the sediments. Their results were similar to the results of this study.

In fact, sediment as the habitat of benthic species plays a major role in their lifestyle and suspension-feeders are more abundant in sandy flats due to current better water situation (Aliakbarian *et al.*, 2020). Grazing process of fauna and infauna species in marine bed directly related to the sediment composition and amounts of sediments organic materials in (Mermillod-Blondin et al., 2005). In addition. wastewater inputs could penetrate the sediments and adversely affect macroinvertebrate diversity in aquatic ecosystems (Foomani et al., 2020). The same results were observed by Mancinelli et al. (1998) for the effect of TOM on microbenthic communities. They reported that the presence of organic matter significantly affects distribution of soft-bottom macrobenthos of the Northern Adriatic Sea.

This study reported the main benthic macroinvertebrates families and species in the Persian Gulf and the Gulf of Oman. Diversity of macroinvertebrates reported and identified species in this study showed a strong relationship with DO and chlorophyll-a concentration (Fig. 3). Other measured environmental factors were also showed a slight influence on the diversity of marine macroinvertebrates. The diverse health status was not totally in a good condition and needs to be improved. These findings could be useful in further studies for habitat management and monitoring programs.

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