

Introduction

Fucoxanthin, a major carotenoid present in the chloroplasts of brown seaweeds, contributes to more than 10% of the estimated total production of carotenoids in nature (Miyashita *et al.*, 2011; Peng *et al.*, 2011). This pigment, along with chlorophylls and β -carotene, widely distributed in brown algae like *Eisenia bicyclis*, *Laminaria japonica*, and *Undaria pinnatifida*, and diatoms such as *Phaeodactylum tricorutum* and *Odontella aurita* (Peng *et al.*, 2011; Takaichi, 2011). Diatoms are unicellular planktonic microalgae and exhibit a characteristic golden-brown color due to a high amount of fucoxanthin that plays a major role in their light-harvesting complex of photosystems (Saudi-Helis *et al.*, 1994; Tezovsnis *et al.*, 1997; Bertrand, 2010).

Fucoxanthin has a unique structure with an unusual allenic bond and a 5, 6-monoepoxide in its molecular structure, and is will metabolized into fucoxanthinol, amarouciaxanthin A, and halocynthiaxanthin after absorption into the human body (Sangeetha *et al.*, 2010).

It provides protective effects on liver, blood vessels of the brain, bones, skin and eyes. It has anti-obesity, anti-diabetic properties, and anti-inflammatory, anti-malarian and anti-angiogenic activities. Moreover it is very effective in inhibiting cell growth and inducing apoptosis in human cancer cells. Particularly, fucoxanthin is distinctly more potent than β -carotene and astaxanthin for anti-obesity activity and the induction of apoptosis in human leukemia (D'Orazio *et al.*, 2012; Kim *et*

al., 2011; Peng *et al.*, 2011). However, although fucoxanthin is clearly a valuable pigment with various biological activities, its use has been limited due to the low extraction efficiency from marine sources and the difficulty of chemical synthesis (Yamano *et al.*, 1995; Kanazawa *et al.*, 2008; Kajikawa *et al.*, 2012). Several studies have been conducted to extract fucoxanthin from brown macroalgae (Kanazawa *et al.*, 2008; Kim *et al.*, 2010), but because these macroalgae are traditional foods in South-East Asia and some European countries, and they contain very low concentrations of fucoxanthin, the production of fucoxanthin from brown macroalgae is not commercially feasible (Kim *et al.*, 2012). Consequently, searching for alternative sources of fucoxanthin is necessary.

Isochrysis galbana is a small unicellular (~5–7 μ m) phytoplankton without a cell wall (Wikfors and Patterson, 1994), from which fucoxanthin could be extracted easily. This microalga contains high concentrations of carotenoid, the long chain polyunsaturated fatty acid, eicosapentaenoic acid (EPA, 20:5 ω 3) commonly grown for the commercial production of live food and lipids (Spolaore *et al.*, 2006, Domer *et al.*, 2014).

Therefore, in the present study; we assumed that *I. galbana* can accumulate high concentration of fucoxanthin in routine culture condition. Thereafter we investigated the effect of nitrogen concentration and salinity on fucoxanthin production. Also the

antioxidant activities of crude fucoxanthin produced were evaluated.

Materials and methods

Culture conditions

The marine microalga *I. galbana* was provided from the Iran Shrimp Research Center. Basal culture media was made with the f/2-Si formula at two different salinity, 20 and 35‰ (T₁ and T₂). Also five different nitrogen concentrations 2, 4, 8, 12mM (T₃, T₄, T₅, and T₆) were investigated (all cultures were run in 3 replicate). The microalgae were cultivated in 1.5-l plastic bottles at 20°C using f/2-Si medium (Guillard and Ryther, 1962), prepared from sterilized distilled water and sea salt, and air was continuously supplied at 5 l/min by an air-lift pump. Light was provided by 60 W fluorescent lamps at an intensity of 2,500 lx. The culture was continuously active and lasted for 10 days after onset of the process. The cells were filtered through a 0.45µm Millipore membrane filter on Büchner funnel (Schott brand) and were freeze-dried at -20°C.

Determination of fucoxanthin contents of I. galbana

Afterward, filter papers were solved in methanol at room temperature for 1 h to extract fucoxanthin from algal samples and then centrifuged at 12000 rpm for 10 minutes twice. Fucoxanthin was quantified using a HPLC system. The mobile phase of methanol and water was eluted with a 1 ml min⁻¹ flow rate by increasing the methanol from 90 to 100% over 30 min and holding for the following 10 min and the

chromatogram was recorded at 445 nm. Each concentration was analyzed by HPLC as described above.

The scavenging activity of DPPH radical was determined as describe by Sachindra *et al.*(2007) briefly, 2 ml methanolic fucoxanthin solution (0.05–0.3 mg ml⁻¹) was mixed with 2 ml 0.16 mM methanolic solution of DPPH. The mixture was shaken vigorously and incubated for 30 min at room temperature in the dark. The absorbance was measured at 517 nm. Ascorbic acid was taken as a positive control. The scavenging ability was calculated as:

DPPH radical scavenging activity (%) = $[1-(A_1-A_2)/A_0] \times 100$,

Where A₀ is the absorbance in the lack of fucoxanthin (using distilled water instead of fucoxanthin), A₁ is the absorbance in the presence of fucoxanthin, and A₂ is the absorbance of methanolic fucoxanthin solution (using methanol instead of DPPH).

Statistical analysis

All data were determined from three replicate of each experiment. Mean values and standard deviations were calculated with Microsoft Excel software. For additional exploration of the differences, the results of one-way ANOVA were compared with Fisher's Least Significant Difference (LSD) in SPSS 19.

Results

The maximum density of the cultures occurred on day 10, in all 6 treatments but a low growth rate occurred in the culture with the highest nitrogen concentration, 12 mM nitrogen supply,

and the 20‰ salinity, containing a low density even after 10 days. Therefore

the biomass didn't harvest from T₁ and T₇.

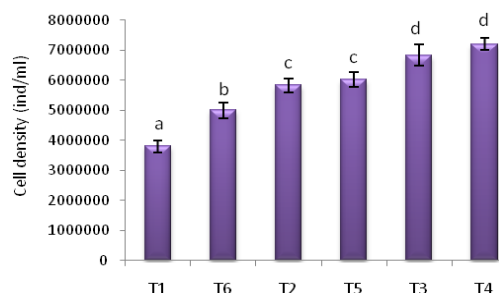


Figure 1: Effect of salinity and nitrogen concentrations on algal biomass (cell density).

Data are shown as Mean \pm STDEV ($n=3$).

The fucoxanthin contents and even color of *I. galbana* in 5 treatments

changed during 10-day cultivation in the high nitrogen cultures (Fig. 2).

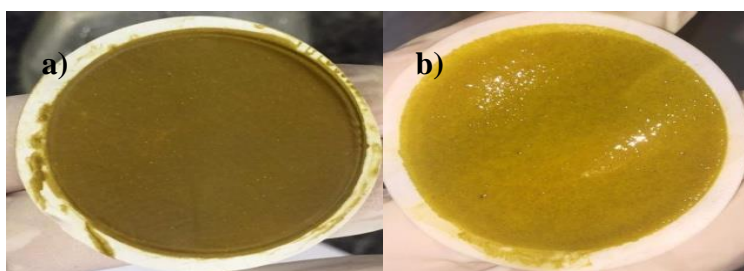


Figure 2: Effect of nitrogen concentration on color of *Isochrysis galbana*. Filter paper with algal sample harvested from a) basal culture medium and, b) culture medium containing four mM.

The fucoxanthin extracted from the 5 treatments increased from 12.1 mg g⁻¹

to 18.1 mg g⁻¹ during 10-day cultivation in the high nitrogen cultures.

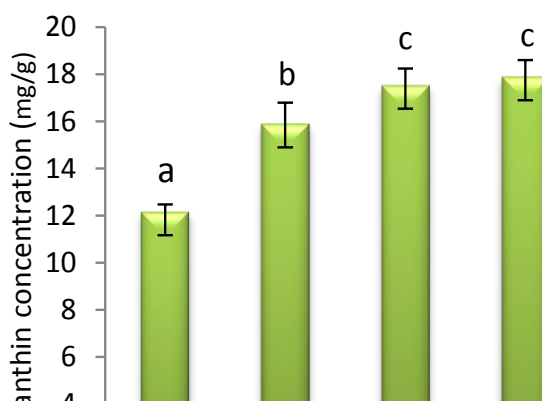


Figure 3: Fucoxanthin concentration of *Isochrysis galbana* cultivated in two different salinities (T₁ and T₂) and 3 different nitrogen supplies (T₃, T₄ and T₅). Each value is expressed as mean \pm STDEV ($n = 3$).

Fucoxanthin extracted from *I. galbana* exhibited strong antioxidant properties, with the effective concentration for 50% scavenging (EC₅₀) of 1,1-dihpenyl-2-picrylhydrazyl (DPPH) radical (Fig. 4). Also the DPPH radical

scavenging activity was linearly dependent on the fucoxanthin concentration; the effective concentration for 50% scavenging (EC₅₀) was 0.2 mg ml⁻¹(Fig. 4).

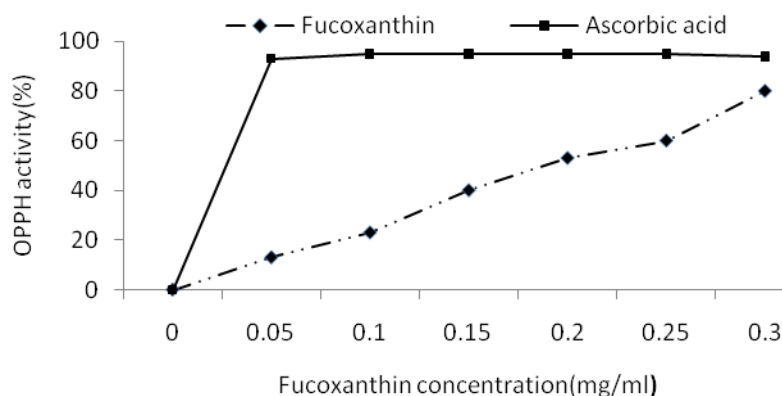


Figure 4: Antioxidant assay for the purified fucoxanthin from *Isochrysis galbana*, scavenging of DPPH radical.

Discussion

Nitrogen concentration and salinity are two major factors affecting growth and pigment biosynthesis of microalgae (Hu, 2004; De la Peña., 2007). A common trend of cellular response to stress conditions, such as high or low salinity and nitrogen depletion, appears to increase secondary carotenoids (e.g., β -carotene, astaxanthin, lutein), which serve as photoprotective agents (Hu, 2004).

In this research, we evaluated the potential of fucoxanthin production in *I. galbana* in different growth experiments. At first step *I. galbana*, were cultivated in basal culture media with 2 different salinities (T₁ and T₂) and different levels of nitrogen (T₃, T₄, T₅ and T₆), then all harvested samples were extracted for fucoxanthin and were analyzed by HPLC at 445 nm.

The maximum density of the cultures of *I. galbana* occurred in 35‰ salinity and media containing four mM nitrogen, also *Isochrysis* harvested from medium containing four mM nitrogen had the highest amount of fucoxanthin (18.1 mg g⁻¹) (Fig. 3). Fucoxanthin produced by *O. aurita* cultivated in basal culture media was measured 4.28 but maximum fucoxanthin concentration of 18.47 mg g⁻¹ was obtained in cultures grown in low light and high nitrogen supply (Xia *et al.*, 2013). Harker *et al.*(1996) reported that the astaxanthin content increased when *H. pluvialis* was cultivated in media deficient in nitrogen. Conversely fucoxanthin concentration decreased in *O. aurita* cultured in the low nitrogen media. Carreto and Catoggio (1976) found that fucoxanthin act as a primary carotenoid, whereby transferring light energy to the

photosynthetic reaction centers for photosynthesis (Hu, 2004). Under stress conditions, changes in the organization of the photosynthetic apparatus (e.g., chloroplast fragmentation, degradation of thylakoid membrane) occur, chlorophyll *a* and other pigments involved in photosynthesis decrease, while the secondary carotenoids increase. These variations in pigment content might be as a quotient between photosynthetically active pigments and other functional pigments.

I. galbana cultured in basal media containing 35‰ salt had the lowest amount of fucoxanthin (12.1 mg g^{-1}), increasing nitrogen (N-NO_3) to four mM, led to a considerable increase in cell density and the fucoxanthin concentration in the cells (18.1 mg g^{-1}) (Figs. 1 and 3). Herzig and Falkowski (1989) also proved that the carotenoid and chlorophyll content are in accordance with nitrogen deficiency. Moreover they demonstrated the nitrogen is a limiting factor for the carotenoid production. Thereafter De la Pena (2007) confirmed that under nutrient repletion chl *a* and photosynthetic pigments decrease, while secondary carotenoids such as fucoxanthin increase.

Whereas in other studies on macroalgae, the fucoxanthin concentration was too low, for example (0.01 mg g^{-1}), in *Sargassum fusiforme*, (1.01 mg g^{-1}) in *S. duplicatum*, (0.73 mg g^{-1}) in *Undaria pinnatifida* for fresh sample (Kim *et al.*, 2012), and 1.01 mg g^{-1} in dried sample of *S. duplicatum*. In contrast, the reported fucoxanthin concentration in microalgae ranges

from 2.24 in *Chaetoceros gracilis* to 18.47 mg g^{-1} in dry biomass of *O. aurita*, which is higher than that found in macroalgae, indicative of the great potential of diatoms as a promising source of fucoxanthin for various commercial applications.

Fucoxanthin extracted from *I. galbana* exhibited strong antioxidant properties. The DPPH radical scavenging activity was linearly dependent on the fucoxanthin concentration; the effective concentration for 50% scavenging (EC50) was 0.2 mg ml^{-1} . It was reported that the extracts of brown seaweed *Cystoseira hakodatensis* is exhibited a strong DPPH radical scavenging activity, due largely to the presence of fucoxanthin (Airanthi *et al.*, 2011). Sachindra *et al.* (2007) assessed the radical scavenging abilities of macroalgae-derived fucoxanthin and its two metabolites, fucoxanthinol and halocynthiaxanthin, against DPPH, and suggested that fucoxanthin and fucoxanthinol exhibited antioxidant activity higher than or similar activities to α -tocopherol.

The production of fucoxanthin from *I. galbana* was very attractive and promising, with the maximum yield of 18.1 mg g^{-1} achieved in the culture medium containing four order nitrogen. Also the purified fucoxanthin showed strong antioxidant properties. Finally, our results may aid the commercial development of this microalgae for large-scale fucoxanthin production as a natural bioresource for human health.

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