DOR: 20.1001.1.15622916.2018.17.3.15.7]

The effect of gokshura (*Tribulus terrestris*) and nettle root (*Urtica dioica*) extracts on growth rate and sex reversal in convict cichlid (*Cichlasoma nigrofasciatum*)

Babahajiani P.1; Shokrollahi B.1*; Gharibkhani M.2

Received: May 2016 Accepted: December 2017

Keywords: Gokshura, Nettle root, Sex reversal, Growth rate, Convict cichlid

- 1-Department of Animal and Aquatic Sciences, Agriculture School, Sanandaj Branch, Islamic Azad University, Sanandaj, Kurdistan, Iran.
- 2-Department of Fisheries, Astara Branch, Islamic Azad University, Astara, Iran.
- *Corresponding author's Email: Borhansh@gmail.com

Introduction

There are many differences between sex of fishes in many traits before and after puberty. Differences in growth rate, size, behavior patterns, and breeding time between males and females have great economic importance (Turan and Cek, 2007). Therefore, many breeders prefer to delay or prevent puberty or to produce unisex populations. Exogenous steroids can be effective in sex reversal of fishes (Al-ablani and Phelps, 2002). Two hormones, estradiol-17β (femalespecific hormone) and 17α methyltestosterone (male-specific hormone) are the most commonly used hormones for sex reversal (Stepherd and Bromage, 1995). Feminization (Liu et al., 1996) and musculinization (Gale et al., 1999) of catfish and Nile tilapia can be performed by direct synthetic hormonal treatment. Previous studies suggested that steroid hormones had disadvantages such as being costly and suppressing growth in teleosts (Hunter and Donaldson, 1983), producing sterile populations and occasional paradoxical feminization with prolonged duration at early stages of gonadal development in African catfish (Van Den Hurk et al., 1989), channel catfish (Goudie et al., 1983), rainbow trout (Solar et al., 1984) and cichlid (Varadaraj et al., 1994). On other hand, steroid hormones are more expensive than plant extracts, and their application in fish is time-consuming labour-intensive and requires professional skills (Cek et al., 2007a). Medical herb extracts are used as an alternative to steroid hormones for sex reversal in different species of fishes.

Gokshura (*Tribulus terrestris*) significantly caused masculinization in

African catfish (Turan and Cek, 2007), convict cichlid (Cek et al., 2007a) and guppy (Cek et al., 2007b). Gokshura could increase testosterone levels and improve athletic performance humans (Adimoelja and Adaikan, 1997, Adimoelia. 2000: Bucci. 2000: Gauthaman et al., 2002) and promote the libido and spermatogenesis in animals and humans (Gauthaman et al., 2003). Gokshura contains a number of substances known as steroidal saponins. Protodioscin. the most dominant saponin in gokshura, is thought to be the main substance responsible for the enhancement of testosterone production (Ganzera et al., 2001). Protodioscin has been reported to increase the levels of dehydroepiandrosterone (Adimoelja and Adaikan, 1997), dihydrotestosterone and dehydroepiandrosterone sulfate (Gauthaman et al., 2000).

Nettle root (Urtica dioica) has a long history of use in folkloric and science based herbal medicine. Nettle root has substantial pharmacological effects such as an immune stimulator, anti-carcinogenic, anti-inflammatory, antioxidant and antiallergenic activities (Glusker and Rossi, 1986; Akbay et al., 2003). The importance of nettle root lignans (such 3.4divanillyltetrahydrofuran) in benign prostatic hyperplasia (BPH) and other estrogen-sensitive androgenand conditions may be due to interference with binding of sex hormone binding globulin (SHBG) to testosterone, the testosterone receptor, and/or the SHBG receptor (Hryb et al., 1995; Schottner et al., 1997).

Convict cichlid (*C. nigrofasciatum*) is a medium-sized cichlid from Central America (Axelrod *et al.*, 1971). Convict cichlid was used as a model fish in the current research because of its broad range of resistance to water hardness, pH and temperature. Convict cichlid is also efficiently attainable and can be proliferated cheaply (Wisenden, 1994; Fraser, 1996). It produces large numbers of viable eggs (100–800 per batch) under optimal conditions. The development period from fertilization to hatching is about 3 days at 27°C (Cek *et al.*, 2007a).

The objectives of this study were to investigate the possible effects of gokshura (*T. terrestris*) and nettle root on sex reversal and growth rate in convict cichlid as an alternative technique.

Materials and methods

Laboratory conditions and experimental design

Six pairs of convict cichlid with a mean weight (±standard deviation) of 8.3 (± 0.4) g and mean length of $(\pm \text{ standard})$ deviation) 7.3 (± 0.3) cm were procured from an ornamental fish dealer (Langrood City, Guilan). Parent pairs were kept in an aquarium containing recirculation water $(26\pm1^{\circ}C)$ exposed 12:12 (light:dark) photoperiod. Once a female spawned in a mussel shell, the eggs hatched within approximately 3 days. The newly hatched larvae were immediately taken out of the hatching tank, counted, measured and transferred to 15 small glass aquaria, each containing 10L water that were regularly aerated with a 4-cm air stone. A total of 525 fry were randomly assigned to five different treatments with three replicates each of which received one of five immersion doses of goskhura and nettle root tested: 0 (T1), 200 mgL⁻¹ nettle root (T2), 300 mgL⁻¹ nettle root (T3), 10 mgL⁻¹ gokshura (T4) and 200 mgL⁻¹ nettle root plus 10 mgL⁻¹ gokshura. The larvae were fed three times a day with commercial flake food supplemented with freshly hatched artemia and tubifex throughout the experimental period of 2 months. Hatchlings were subjected to immersion in water containing gokshura and nettle root extracts from the age of day one to 8 weeks. The aquarium system was immobile, and bathing medium was changed 2 times in a week with the same concentration of gokshura and nettle root in each treatment.

Plants extraction

250 grams of whole washed and shade dried gokshura whole plant and nettle root (3 times) were powdered and extracted with 500 mL 70% ethanol (Hussain *et al.*, 2009).

Measurement of parameters

At the end of the experiment, all fish (60 days old) were anesthetized with 2-phenoxethanol (0.04%) and stored at -20°C for assessing whole body weights (BW) and lengths (BL). Then growth parameters including specific growth rate (SGR) (Clark *et al.*, 1990), daily growth rate (DGR), percent of body weight gain (BWG%) (Ergun *et al.*, 2003), condition factor (CF) ((Weight/(length)^3)*100) and survival

rate (SR) were calculated. Secondary sexual characteristics were used to distinguish males from females. Also, the structure of the testis and ovaries were observed by naked eye and examined under a light microscope (Olympus BX 50) at the end of the experiment.

Statistical analysis

The Chi Square test was used to test whether the observed sex ratios were significantly different from the expected sex ratios. Differences among groups were evaluated by one-way analysis of variance (ANOVA) using the SAS software (SAS 9.2). Duncan testing was performed for intergroup comparisons; *p*<0.05 was considered statistically significant.

Results and discussion

The aim of the present study was to evaluate the effect of gokshura and nettle root on the sex reversal and growth rate in convict cichlid. The effects of different immersion concentrations of plant extracts on sex reversal of convict cichlid for 60 days are presented in Table 1. The observed sex ratio in the control treatment (T1) was not significantly different. Nettle root treated fish (T2 and T3) yielded 100 % female fish, while in the gokshura treatment (T4), there were no significant statistically differences between male and female counts (p>0.05). We achieved significantly higher (68.76%) masculinization by immersing 0-day-old fry for 60 days in water containing gokshura and nettle root (T5) (*p*<0.05).

Table 1: Effect of gokshura and nettle root treatment on sex reversal in convict cichlid.

Treatments	Sex ratio (%)		Sex distributions		χ^2 (p-value)
	Female	Male	Female	Male	
Control (T1)	46.65	53.85	30	35	NS
T2	100	0	87	0	***
T3	100	0	76	0	***
T4	39.73	60.27	29	44	NS
T5	37.14	62.86	26	44	*

NS: not significant (p>0.05), *: significant (p<0.05), ***: significant (p<0.001)

The results indicated that gushura (10 mgL⁻¹) plus nettle root (200 mgL⁻¹) significantly effective were masculinization of convict cichlid. Masculinization of fish which received $mg L^{-1}$ gokshura was nonsignificantly effective. Previous studies showed the positive effects of gokshura 200 and 300 mgL^{-1}) on (100,masculinization of convict cichlid (Cek et al., 2007a) and African catfish (Turan and Cek, 2007). In contrast, 50 and 100 mgL⁻¹ of gokshura had no effect on sex reversal in Poeciliata reticulata (Cek et al., 2007b). Fish studies suggested that higher doses may have generated better results masculinization; we cannot support this conclusion from our results. In the current study, first we used the same amount of aqueous or/and alcoholic gokshura extract (200, and 300 mgL⁻¹). However after a few turns around (about 3 minutes), the treated larvae died out. Therefore, we examined another amount (90, 80 to 10 mgL⁻¹) of gokshura extract and all levels, except 10 mgL⁻¹ led to the death of treated larvae. Finally, our results showed that gokshura is toxic to convict cichlid in doses higher than 10 mgL⁻¹. Studies in human (Adimoelja, 2000; Bucci, 2000)

and rat (Gauthaman et al., 2002; Gauthaman et al., 2003) demonstrated gokshura treatments strongly affected androgen metabolism, and significantly increased testosterone or testosterone precursor levels. Kavitha and Subramanian (2011) showed that the activities of testicular functional enzyme levels were increased in gokshura treated groups compared with that of the control in Poecilia latipinna fishes. A high rate of masculinization was observed in our study (in T4 and T5) but whether this potency is caused increases in androgens by testosterone cannot be figured out, because we did not measure the plasma level during testosterone the experiment.

The effects of different immersion concentrations of gokshura and nettle root on growth indices of convict cichlid for 60 days are presented in Table 2. Mean of BW, BL, SGR, DGR, and CF for the 60-day BWG% experimental period differed significantly among treatments (p<0.05). BW and BL decreased in immersion treatments compared to control fish. In T5 treatment, the lowest BW, BL, SGR, DGR and BWG% were seen. CF index was substantially higher in T3 and T5 treatments than in other treatments. Higher doses of nettle root (T3) led to higher BW, BL, SGR, DGR, BWG%, CF parameters compared to lower doses (T2). SR in the extract-

treated groups did not significantly differ from the control indicating that Gokshura and nettle root had no negative effect on survival rate.

Table 2: Effect of gokshura and nettle root treatment on growth parameters in convict cichlid.

	T1	T2	Т3	T4	T5
BW (g)	2.19 ± 0.05^{a}	0.83±0.021°	1.42 ± 0.02^{b}	1.73 ± 0.02^{ab}	0.66±0.01°
BL (cm)	4.61 ± 0.05^{a}	2.84 ± 0.01^{c}	2.87 ± 0.03^{c}	3.97 ± 0.02^{b}	1.99 ± 0.01^{d}
DGR	0.017 ± 0.00^{a}	0.006 ± 0.00^{c}	0.011 ± 0.00^{b}	0.014 ± 0.00^{b}	0.005 ± 0.00^{c}
BWG (%)	98.60 ± 0.05^{a}	96.22 ± 0.08^{b}	97.83 ± 0.04^{a}	98.23±0.02a	95.31±0.091°
SGR	3.55 ± 0.02^{a}	2.75±0.01°	3.20 ± 0.01^{b}	3.37 ± 0.01^{ab}	2.56 ± 0.01^{c}
CF	2.18±0.103°	3.69 ± 0.10^{c}	6.22 ± 0.21^{b}	2.79 ± 0.05^{c}	8.64 ± 0.29^{a}
SR (%)	81. 9±2.51	82.85 ± 9.89	82.85 ± 5.94	89.52±7.43	87.62±10.73

The values with different letters are significantly different (p<0.05).

Significant decreases in body weight parameters (BW, BL, SGR, DGR and BWG) were seen in all extract treated fish compared to control fish in the current study. Contrarily, Cek et al. 2007b), Kavitha (2007a, and Subramanian (2011) and Yılmaz et al. (2013) found that the growth rate of fish treated with gokshura was faster than that of the controls in Cichlasoma nigrofasciatum. Р. reticulata. latipinna and Oncorhynchus mykiss. On the other hand, Turan and Cek (2007) gokshura had suggested that substantial effect on growth rates in Clarias gariepinus. Similar to our results, survival ratios in the gokshuratreated fish were similar to that in controls (Cek et al., 2000a, 2007b; Turan and Cek, 2007; Yılmaz et al., 2013).

Disagreement of the results of the current study with previous mentioned studies can be attributed to extract type and related compounds. In these studies, the industrial gokshura extract was used, whereas in our study, whole plants were purchased from local

markets and then gokshura extract was prepared.

The current study was the first attempt yielding complete feminization of convict cichlid immersed in Nettle root (200 and 300 mgL⁻¹) compared to other experimental treatments. Growth rate parameters (BW, BL, SGR, DGR and BWG) had decreased in fish receiving nettle root in comparison with the control group, but survival rates were not significantly different among treatments as nettle root had no effect on survival rates. There are no published data for evaluating the effect of nettle root on sex reversal or growth rate in fish species. However the results of a study in rats showed that the body weight was reduced intraperitoneally in rats that received 300 mgkg⁻¹day⁻¹ nettle root compared to the control group and the mean number of spermatogonia per tubular tubules, leydig cells mm⁻² in diameter tubules (µm) were decreased in the experiment group treated with 300 mgkg⁻¹day⁻¹ Nettle root (Pourahmadi et al., 2014). The effect of nettle root on the body weight is due to ingredients such as flavonoids that induce lipid hydrolysis competitive by blockage of phosphodiesterase, and also inhibition of 3-hydroxy-glutaril CoA key to cholesterol biosynthesis in the liver that can reduce body weight or flavonoids that regulate the energy metabolism and body weight by binding to the site where ATP binds to enzymes and receptors (Kandaswmi, 1999).

In another report, Pourahmadi et al. (2013) showed that the nettle root extract led to a significant increase in LH, whereas the effect on FSH was variable and non-significant. We did not asses the effect of nettle root on gonadotropines or sex hormones in convict cichlid. Nettle extract contains linoleic acid, which can decrease cholesterol. It may reduce testosterone by lowering cholesterol. Diminishing testosterone levels, increases LH and FSH. LH affects the levdig cells and leads to an increase in their number and activity (Morovvati et al., 2013). Nettle root contains compounds such as sterols, flavonoids and polysaccharides that have antiandrogen effects and deal with the testosterone function (Akbay et al., 2003; Ganzera et al., 2005). Nettle root extract has at least 18 types of sterols and 8 types of lignan (Kraus and 1990). It prevents Spiteller, formation of the active form of testosterone, dihydrotestosterone, by inhibition of the enzyme 5-alpha reductase (Koch, 2001; Nahata and Dixit, 2012). Nettle root extract inhibits aromatase, therefore, it prevents the conversion of testosterone to estrogen, and also prevents androgen binding to androgen receptors (Chrubasik *et al.*, 2007). These descriptions may be possible causes of the feminization of convict cichlid in our study.

In conclusion, the results of the current study do not support the enhancement effect of gokshura on masculinization and the growth rate of convict cichlid. On the other hand, our results add a support to the effect of nettle root on the feminization property. Future studies subjected at measuring the amount of sexual hormone levels after nettle root treatment in convict cichlid supply more deterministic evidence about the effects of nettle root on the sex ratio and whether it can be successfully used as an agent in fish culture. Further investigations necessary to determine the effects of nettle root on other cultivable fish species.

Acknowledgments

The authors express their gratitude to Islamic Azad University, Sanandaj Branch for its financial support.

References

Adimoelja, A. and Adaikan, P.G., 1997. Protodioscin from herbal plant *Tribulus terrestris* L.improves male sexual functions possibly via DHEA. *International Journal of Impotence Research*, 9, 64.

Adimoelja, A., 2000. Phytochemicals and the breakthrough of traditional herbs in the management of sexual dysfunctions. *International Journal of Andrology*, 23(2), 82-84.

- Akbay, P., Basaran, A.A., Undeger, U. and Basaran, N., 2003. In vitro immunomodulatory activity of flavonoid glycosides from *Urtica dioica* L. *Phytotherapy Research*, 17(1), 34-37.
- **Al-ablani, S.A. and Phelps, R.P., 2002.** Paradoxes in exogenous androgen treatments of bluegill. *Journal of Applied Ichthyology*, 18, 61-66.
- Axelrod, H.R., Emmens, C.V., Sculthorpe, D., Wingler W.V. and Pronek, N., 1971. Exotic tropical fishes. TFH Publications, Inc. Jersey City, NJ.
- Bucci, L.R., 2000. Selected herbals and human exercise performance.American Journal of Clinical Nutrition, 72(2), 624-636.
- Cek, S., Turan, F. and Atik, E., 2007a. Masculinization of convict cichlid (*Cichlasoma nigrofasciatum*) by immersion in *Tribulus terrestris* extract. *Aquaculture International*, 15, 109–119.
- Cek, S., Turan, F. and Atik, E., 2007b. The effects of Gokshura, *Tribulus terrestris* on sex reversal of guppy, *Poecilia reticulata*. *Pakistan Journal of Biological Sciences*, 10(5), 718-725.
- Chrubasik, J.E., Roufogalis, B.D., Wagner, H. and Chrubasik, S., 2007. A comprehensive review on the stinging nettle effect and efficacy profiles. Part II: urticae radix. *Phytomedicine*, 14(8), 568-579.
- Clark, A.E., Watanabe, W.O., Olla, B.L. and Wicklund, R.I., 1990. Growth, feed conversion and protein utilization of Florida red tilapia feed

- isocaloric diets with different protein levels in seawater pools. *Aquaculture*, 88, 75-85.
- Ergun, S., Yigit, M. and Turker, A., 2003. Growth and feed consumption of young rainbow trout (*Oncorhynchus mykiss*) exposed to different photoperiods. *Journal of Aquaculture Bamidged*, 55, 132-138.
- **Fraser, S.A., 1996.** The influence of predators on adaptation behaviour in adult convict cichlid *Cichlasoma nigrofasciatum*. *Canadian Journal of Zoology*, 74, 1165-1173.
- Gale. W.L.. Fitzpatrick, M.S.. Lucero, M., Contreras-Sanchez, W.M. and Schreck, C.B., 1999. Masculinization of Nile tilapia (Oreochromis niloticus) by immersion in androgens. Aquaculture, 178, 349-357.
- Ganzera, M., Bedir, E. and Khan, I. A. 2001. Determination of steroidal saponins in Tribulus terrestris by reversed-phase high-performance liquid chromatography and evaporative light scattering detection. *Journal of Pharmaceutical Sciences*, 90(11), 1752-1758.
- Ganzera, M., Piereder, D., Sturm, S., Erdelmeier, C. and Stuppner, H., 2005. Urtica dioica agglutinin: identification. separation, and quantitation of individual isolectins by capillary electrophoresis capillary electrophoresis-mass Electrophoresis, spectrometry. 26(9), 1724-1731.
- Gauthaman, K., Adaikan, P.G., Prasad, R.N.V., Goh, V.H.H. and Ng, S.C., 2000. Changes in hormonal paramdeters secondary to

- intravenous administration of *Tribulus terrestris* extract in primates. *International Journal of Impotence Research*, 12(2), 6.
- Gauthaman, K., Adaikan, P.G. and Prasad, R.N. 2002. Aphrodisiac properties of *Tribulus terrestris* extract (Protodioscin) in normal and castrated rats. *Life Science*, 71(12), 1385-1396.
- Gauthaman, K., Ganesan, A.P. and Prasad, R.N., 2003. Sexual effects of puncturevine (*Tribulus terrestris*) extract (protodioscin): an evaluation using a rat model. *Journal of Alternative and Complementary Medicine*, 9(2), 257-265.
- Glusker, J.P. and Rossi, M., 1986.

 Molecular aspects of chemical carcinogens and bioflavonoids.

 Progress in Clinical and Biological Research, 213, 395-410.
- Goudie, C.A., Redner, B.D., Simco, B.A. and Davis, K.B., 1983. Feminization of channel catfish by oral administration of steroid sex hormones. *Transactions of the American Fisheries Society*, 112, 670-672.
- Hryb, D.J., Khan, M.S., Romas, N.A. and Rosner, W., 1995. The effect of extracts of the roots of the stinging (Urtica dioica) nettle on the interaction **SHBG** with its receptor human prostatic on membranes. Planta Medecine, 61(1), 31 - 32.
- **Hunter G.A. and Donaldson E.M., 1983.** Hormonal sex control and its application to fish culture. *Fish Physiology*, 9B, 223–303.

- Hussain, A.A., Mohammed, A.A., Ibrahim, H.H. and Abbas, A.H., 2009. Study the biological activities of *Tribulus terrestris* extracts. *World Academy of Science, Engineering and Technology*, 57, 433-435.
- **Kandaswmi, C., 1999.** Effect of plant the flavonods on mammalian cells. *Pharmacology Review*, 54(2), 673-751.
- **Kavitha, P. and Subramanian, P., 2011.** Influence of *Tribulus terrestris* on testicular enzyme in fresh water ornamental fish *Poecilia latipinna*. *Fish Physiology and Biochemistry*, 37(4), 801-807.
- **Koch, E., 2001.** Extracts from fruits of saw palmetto (*Sabal serrulata*) and roots of stinging nettle (*Urtica dioica*): viable alternatives in the medical treatment of benign prostatic hyperplasia and associated lower urinary tracts symptoms. *Planta Medecine*, 67(6), 489-500.
- Kraus, R. and Spiteller, G., 1990. Phenolic compounds from roots of *Urtica dioica*. *Phytochemistry*, 29(5), 1653-1659.
- Liu, S., Yao, Z. and Wang, Y., 1996.

 Sex hormone induction of sex reversal in the teleost *Clarias lazera* and evidence for female homogamety and male heterogamety. *Journal of Experimental Zoology*, 276, 432-438.
- Morovvati, H., Najafzadehvarzi, H. and Rashidi, K., 2013. Effect of *Urtica dioica* extract on histological and histometrical changes of testis of hamster after testosteron administration. *Zahedan Journal of*

- Research in Medical Sciences, 15(11), 4-8.
- Nahata, A. and Dixit, V.K., 2012.

 Ameliorative effects of stinging nettle (*Urtica dioica*) on testosterone-induced prostatic hyperplasia in rats. *Andrologia*, 44(1), 396-409.
- **Pan, L.Q., Luan, Z.H. and Jin, C.X., 2006.** Effects of Na⁺/K⁺ and Mg²⁺/Ca²⁺ rations in saline groundwaters on Na⁺- K⁺- ATPase activity, survival and growth of *Marsupenaeus japonicus* post larvae. *Aquaculture*, 261, 1396-1402.
- Pourahmadi, M., Bagheri, M., Karimi-Jashni, H., Kargar-Jahromi, H. and Zarei, S., 2013. The effect of hydroalkoholic extract *urtica dioica* on concentrations of sex hormones in adult male rats. *Journal Jahrom University of Medical Sciences*, 10(4), 25-29.
- Pourahmadi, M., Karimi-Jashni, H., Bagheri, M. and Sotoodeh-Jahromi, A., 2014. The effect of hydro-alcoholic extract of *Urtica dioica* root on testes in adult rats. *Life Science Journal*, 11(5), 420-424.
- Schottner, M., Gansser, D. and Spiteller, G., 1997. Lignans from the roots of *Urtica dioica* and their metabolites bind to human sex hormone binding globulin (SHBG). *Planta Medicine*, 63(6), 529-532.
- Shepherd, J. and Bromage, N., 1995. Intensive fish farming. First Publishing. Billing and Sons Ltd. Worcester, 404 P.
- Solar, I.I., Donaldson, E.M. and Hunter, G.A., 1984. Optimization

- of treatment regimes for controlled sex differentiation and sterilization in wild rainbow trout (*Salmo gairdneri*) by oral administration of 17α -methyltestosterone. *Aquaculture*, 42,129-139.
- Turan, F. and Cek, S., 2007.

 Masculinization of African catfish (*Clarias gariepinus*) treated with Gokshura (*Tribulus terrestris*). The Journal of Aquaculture, 59(4), 224-229.
- Van Den Hurk, R., Richter, C.J.J. and Janssen, J., 1989. Effects of 17 alpha-methyltestosterone and 11 beta-hydroxyandrostenedione on gonad differentiation in the African catfish. *Aquaculture*, 83, 179-191.
- Varadaraj, K., Kumari, S.S. and Pandian, T.J., 1994. Comparison of conditions for hormonal sex reversal of Mozambique tilapias. *Progress in Fish Culture*, 56, 81-90.
- Wisenden, B.D., 1994. Factors affecting reproductive success in free ranging convict cichlids. *Canadian Journal of Zoology*, 72, 2177–2185.
- Yılmaz, E., Çek, S. and Mazlum, Y., 2013. Effects of synthetic and natural steroids on the growth, sex reversal and gonadal development of rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Ege Journal of Fishery and Aquatic Sciences*, 30(3), 123-131.