

Evaluation of the chemical composition and in vitro antimicrobial activity of *Rosmarinus officinalis*, *Zataria multiflora*, *Anethum graveolens* and *Eucalyptus globulus* against *Streptococcus iniae*; the cause of zoonotic disease in farmed fish

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Abstract

There is a growing interest of industry to replace synthetic chemicals by natural products with bioactive properties from plant origin. The purpose of this study was to elucidate the factors affecting antimicrobial effectiveness of essential oils *Rosmarinus officinalis*, *Zataria multiflora*, *Anethum graveolens* and *Eucalyptus globulus* against food spoilage and pathogenic bacteria, *Streptococcus iniae*; the cause of zoonotic streptococcosis in fish. Food conservation is based on an intermittent search for foods with a high nutritional quality and microbial stability and it has been reached by the control of the growth / survival of spoiling and pathogen foodborne microorganisms. Based on several reports, fish streptococcosis is currently considered as one of the main limiting factors in the aquaculture industry, due to the significant economic losses (annually more than \$150 million) that these infections cause in different cultured fresh and seawater fish species worldwide. The sensitivity of *S. iniae* to antibacterial activity of the essential oils was determined using well diffusion assays and paper disc diffusion method. The ranges of minimum inhibitory concentration (MIC) of the oils and extracts were 3.9–250 and 7.8- 500 µg/ml and the ranges of minimum bactericidal concentration (MBC) values for the oils and extracts were found to be in the range of 7.8- 250 and 15.6-500 µg/ml, respectively. The essential oils exhibited antibacterial activity against *S. iniae*. The essential oil of rosemary showed the strongest antimicrobial activity.

Keywords: Essential oils (EO), *Rosmarinus officinalis*, *Zataria multiflora*, *Anethum graveolens*, *Eucalyptus globulus*, Antibacterial activity, Minimum inhibitory concentration (MIC)

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Introduction

There has been growing interest in the investigation of natural products for the discovery of active compounds with antimicrobial and antioxidant properties that can be applied to the food industry. As consumers are avoiding the consumption of products with synthetic additives or preservatives, natural active compounds could be an alternative to the employ of synthetic chemicals. Such compounds can be used to prolong the storage stability of food, by inhibiting the growth of foodborne spoilage or pathogenic microorganisms and protecting food from oxidative stress damage. Several authors have tested essential oils of aromatic plants to prolong the shelf-life of food (Mahmoud et al., 2006 ; Fernández *et al.*, 2009 ; Gómez-Estaca et al., 2010), while others have focused on the antioxidant and antimicrobial properties of plant extracts and essential oils (Atrea et al., 2009; Harzallah et al., 2011). Foodborne diseases are still a major concern in the world. According to the reports regarding foodborne microorganisms outbreak, *Salmonella enteric*, *Staphylococcus aureus*, *Bacillus cereus*, *Streptococcus iniae*, *Lactococcus* spp. and *Clostridium botulinum* were found in eggs, beef, chicken, fish and dairy products in the USA and the European Union, as well as in developing countries (Ho et al., 2010).

In the past few decades, the pathogen *Streptococcus iniae* has emerged as a major hindrance to aquaculture operations worldwide, causing economic losses measured in hundreds of millions of dollars annually. In addition, *S. iniae* has

established itself as a zoonotic risk, especially in areas of the world that preferentially prepare and consume raw fish. To date, at least 25 human causes of invasive streptococcal infection attributed to *S. iniae* have been confirmed in the USA, Canada, China and Taiwan, many of which were in immunocompromised patients; since there is currently no prospective epidemiological surveillance for human *S. iniae* infections, the true number may be much higher (Milani et al., 2010).

Rosemary, *Rosmarinus officinalis* L. (Labiatae) is an evergreen perennial shrub grown in many parts of the world. It has been reported to possess a number of therapeutic applications in folk medicines in curing or managing of a wide range of diseases. It is well known that the activity of rosemary extracts in medicine and food industry due to the presence of some important antioxidant oil and phenolic components, to prevent oxidative degradation of oil and lipid containing foods. Rosemary has long been recognized as having antioxidant molecules, such as rosmarinic acid, carnosol, rosmaridiphenol and these have found in ethanol-soluble fraction (Bakirel et al., 2008).

Zataria multiflora is a plant belonging to the Lamiaceae family that geographically grows only in Iran, Pakistan and Afghanistan. This plant has the local name of Avishan Shirazi (in Iran) and traditional uses such as antiseptic, anesthetic and antispasmodic. This plant is extensively used as a flavor ingredient in a wide variety of food in Iran. The main constituents of the essential oil of this plant are phenolic compounds such as

carvacrol and thymol (Misaghi and Akhondzadeh, 2007).

Anethum graveolens L. (Umbelliferae), known as dill, is an annual herb growing in the Mediterranean region, Europe, central and southern Asia and it is widely cultured in south eastern region of Iran. The plant is used both medicinally and as an aromatic herb and spice and cookery. Dill has been used traditionally for gastrointestinal ailments such as flatulence, indigestion, stomachache colic and to tract intestinal gas (Yazdanparast and Bahramikia, 2008).

Eucalyptus spp. (family Myrtaceae) originated in Australia, but these plants now grow in almost all tropical and subtropical areas, and are cultivated in many other climates. The genus *Eucalyptus* contains about 600 species. Of all the species, *Eucalyptus globulus* is the most widely cultivated in subtropical and Mediterranean regions. As *Eucalyptus* is a fast-growing tree, and is a suitable ingredient for paper manufacture, there has been extensive overseas forest plantation of *Eucalyptus* trees. Much research has been conducted on the medicinal properties of *Eucalyptus* spp. The leaf extract or essential oil from the leaves of *Eucalyptus* spp. has been reported to possess antifungal, antibacterial, mosquito repellent and antioxidant properties (Salari et al., 2006).

This report provides a basis for further exploitation and use of the four plant resources on human health and food safety. The aim of this study was to determine the chemical composition of *R. officinalis*, *Z. multiflora*, *A. graveolens* and *E. globulus* essential oil and to

characterize the in vitro antimicrobial activities of extracts and essential oil against *S. iniae*.

Materials and methods

Preparation of essential oil and extracts

R. officinalis, *E. globulus* and *A. graveolens* were collected from suburbs of Karaj city (Alborz Province, Iran) and *Z. multiflora* Boiss. was collected in Fars Province, south of Iran in June 2011 and identified by Institute of Medicinal Plants, Tehran, Iran. Air-dried aerial parts of the *R. officinalis*, *Z. multiflora* and *E. globulus* and seed of *A. graveolens* were subjected to steam distillation for 2 h using Clevenger-type apparatus. The essential oils were analyzed on an Agilent 6890 gas chromatograph interfaced to an Agilent 5973 N mass selective detector (Agilent Technologies, Palo Alto, USA). A vaporization injector operating in the split mode (1:50) at 250 °C was used, into which a fused silica capillary column (30m length × 0.32 mm internal diameter × 0.25µm film thickness; HP-5MS; 5% diphenyl, 95% dimethyl polydimethylsiloxane, Agilent Technologies) was installed. Briefly, the essential oils were collected from the airdried for 3 h using a Clevenger-type apparatus (Hanil Labtech Ltd., Incheon, Korea), and was dried over anhydrous sodium sulfate for 24 h, measured and stored in hermetically sealed dark-glass containers at -4°C until it was tested and analyzed by gas chromatography/mass spectrometry (GC/MS). The data were acquired under the following conditions: initial temperature 50 °C; program rate 2.5 °C; final temperature 300 °C and injector

temperature 290 °C. The carrier gas was helium and the split ratio was 0.8 ml/min. For GC–MS detection, an electron ionization system with ionization energy of 70 eV was used.

Thirty grams of the dried and powdered plant materials were extracted with methanol by using Soxhlet apparatus at 60 °C for 12 h. The extracts was filtered and concentrated under vacuum at 40 °C by using a rotary evaporator (Heidolph, Laborota 4000, Schwabach, Germany), yielding a waxy material (2.25 g, 7.5% w/w). This extract was suspended in water and extracted with chloroform (4 × 100 ml) to obtain 1.35 g (4.5%) polar and 0.84 (2.8%) non polar extracts. The extract was stored in darkness at 4 °C until used within a maximum period of one week (Jiang et al., 2011).

Bacterial strains and culture conditions

Lyophilized culture of *S.iniae* obtained from Department of Aquatic Animals Health, Faculty of Veterinary Medicine, University of Tehran, Iran, was used in this study. The lyophilized culture was grown in tube containing 10 ml of BHI broth (Merck KGaA, Darmstadt, Germany) twice consequently and incubated each time at 30 °C for 18 h. Then it was followed by streaking on BHI agar (Merck KGaA) slant and incubated at 30 °C for 18 h. The culture was stored at 4 °C as working culture and subcultured at monthly intervals. The bacterial cells of *S. iniae* grown in the broth were adjusted to an optical density (OD) of 0.02 at 600 nm using the Spectronic 20 spectrophotometer (Milton Roy Company, USA). This adjustment gave a cell concentration of 1×10^5 CFU / mL (Moosavy et al., 2008).

Paper disc diffusion method

The disc diffusion method was applied for the determination of antimicrobial activities of the extracts and essential oil (NCCLS, 2000a). Extracts and essential oils were dissolved in dimethyl sulfoxide (DMSO). *S.iniae* were cultured in a nutrient broth (Merck, Germany) for 24 h and diluted with sterilized peptone water. Then, 100 µl of each culture (10^5 CFU) was spread onto the surface of Mueller–Hinton agar (Oxoid, England) to create a bacterial lawn. Sterile blank filter paper discs of 6 mm in diameter (Oxoid, England) were wetted with 20 µl crude extract of *R. officinalis*, *Z. multiflora*, *E. globulus* and *A. graveolens* (100 mg/ml) and left to dry before being placed on the microbial lawn. The plates were incubated at 37 °C for 24 h. The antibacterial activity was compared with 10 µl of chloramphenicol (1 mg/ml), 20 µl of 5 % lactic acid and 20 µl of 10 % acetic acid. Antimicrobial activity was determined based on the diameter of the clear zone surrounding the paper discs (Celiktas et al., 2007). Three replicate discs were prepared for each extract and essential oil in this study.

Determination of minimal inhibitory concentration

The estimation of the minimal inhibitory concentration (MIC) and minimal bactericidal concentration (MBC) were measured by the broth microdilution method (NCCLS, 2002). The essential oils were individually dissolved in sterilized physiological saline solution (0.9% w/v) supplemented with Tween 80 (Sigma) at a final concentration of 0.5% (v/v). Serial

doubling dilutions of the oils were prepared in a 96-well microtiter plate in the range 1000 to 7.8 µg/mL. Each essential oil dilution (100 µL) was dispensed into the wells of a microtiter plate; each well was then inoculated with 100 µL of the suspension. The resulting suspensions were mixed with a micro-pipettor. The final concentration of each strain was adjusted to 10^5 – 10^6 CFU / mL. All microtiter plates against all microorganisms were incubated at 37 °C for 24 h. After incubation, the wells were examined for growth of microorganisms and the MIC was determined. The MIC is defined as the lowest concentration of the essential oil at which the microorganism does not demonstrate visible growth. The MBC were confirmed by reinoculating on agar plates with 10 µL of each culture medium from the microplates. The MBC is defined as the lowest concentration of the essential oil at which incubated microorganisms are completely killed. Each experiment was repeated three times (Fu et al., 2007).

Statistical analysis

Differences between extracts and essential oil were tested with analysis of variance (ANOVA). In order to satisfy ANOVA assumptions data were transformed, followed by multiple comparisons tests (Tukey's HSD) to identify differences between groups. All statistical analyses were tested at a 0.05 level of probability with the software STATISTICATM 6.1 (Statsoft, Inc., Tulsa, OK, USA).

Results

Chemical composition of essential oil

Regarding the chemical composition of the essential oils tested, they were shown to be complex mixtures of many components. The relative quantitative values of *R. officinalis* are presented in Table 1. The most important constituents of the rosemary are 1,8-cineole (78.6%) , alpha-pinene (5.87%), toluene (12.26%), Camphor (8.22%) and Berbonone (7.75%). Table 1 shows the identified compounds and percentage obtained by GC/MS, as well as the retention time listed in order of their elution from the DB-5 capillary column.

Table 1: Essential oil composition of *R. officinalis* identified by GC–MS

| Compound | Retention time (min) | Percentage |
|-----------------|----------------------|------------|
| 1-Hexanol | 8.32 | 0.35 |
| Alpha-pinene | 11.39 | 15.87 |
| Camphene | 12.01 | 4.20 |
| Verbenene | 12.26 | 0.57 |
| Beta-pinene | 13.36 | 0.48 |
| 3-Octanone | 13.98 | 2.76 |
| Beta-myrcene | 14.23 | 2.38 |
| Benzene | 15.85 | 1.15 |
| 1,8-cineole | 16.24 | 78.60 |
| Gamma-terpinene | 17.60 | 0.40 |
| 1-octanol | 18.33 | 0.39 |
| Linalool | 19.80 | 2.20 |
| Camphor | 21.92 | 8.22 |
| Propanoic acid | 22.19 | 0.82 |
| Berbonone | 22.64 | 7.75 |

| | | |
|---------------------------------|-------|-------|
| Bicycloheptane | 28.69 | 2.34 |
| Acetic acid | 25.27 | 0.97 |
| Toluene | 4.90 | 12.26 |
| Dodecane | 24.80 | 0.60 |
| Ethanol | 8.23 | 0.04 |
| Naphthalenone | 74.33 | 0.12 |
| Eicosadiene | 72.55 | 1.17 |
| 9-Octadecenoic acid | 60.24 | 1.89 |
| Octadecanoic acid | 60.76 | 0.65 |
| Phosphoric acid | 68.29 | 0.47 |
| 1,4,7,10,13,16-Hexaoxacycloocta | 73.87 | 2.53 |
| Hexadecanoic acid | 54.63 | 1.39 |
| Alpha- Fenchyl acetate | 40.75 | 1.65 |
| Heptadecene | 60.11 | 2.18 |
| Borneol | 23.07 | 5.45 |
| Butanoic acid | 24.36 | 5.70 |
| 15-Nonylphenyl | 74.80 | 0.40 |
| Benzaldehyde | 12.64 | 1.11 |
| 1,19-Eicosadiene | 72.55 | 1.17 |
| Gamma-sitosterol | 73.95 | 1.51 |
| Benzaldehyde | 12.64 | 1.11 |

Volatiles of *A.graveolens* essential oil accounting for 99.34 % of the essential oil revealed 21 different compounds composition that are identified in Table 2.

Table 2: Essential oil composition of *A. graveolens* identified by GC-MS

| Compound | Retention index ^a | Percentage |
|------------------------|------------------------------|-----------------|
| α - Pinene | 933 | 0.31 |
| Sabinene | 977 | tr ^b |
| β -Myrcene | 989 | 0.25 |
| α - Phellandren | 1004 | 0.75 |
| Limonene | 1030 | 19.89 |
| γ -Terpinene | 1059 | 0.34 |
| ρ -Cymene | 1090 | 0.41 |
| Linalool | 1096 | 1.68 |
| E-Limonene Oxid | 1139 | tr |
| Estragole | 1180 | 0.94 |
| α - Terpeneol | 1188 | 0.17 |
| Z-Dihydrocarvone | 1202 | 6.59 |
| E-Dihydrocarvone | 1211 | 7.36 |
| Cumin aldehyde | 1235 | 0.60 |
| D-Carvone | 1243 | 36.09 |
| Thymol | 1294 | 6.50 |
| Carvacrol | 1303 | 0.21 |
| β - Caryophyllen | 1429 | 0.32 |
| D-Germacrene | 1490 | 0.10 |
| Myristiscin | 1527 | tr |
| Dill apiole | 1634 | 16.83 |

^a Retention index on D B - 1 column , ^b tr = trace < 0.1 %

The essential oil of *A. graveolens* contained a complex mixture consisting mainly by D-Carvacrol (36.09%), such as

Limonene (19.89%), Dill apiole (16.83%), E-Dihydrocarvone (7.36%) and Z-Dihydrocarvone (6.59%).

Table 3: Essential oil composition of *Z. multiflora* Boiss. identified by GC-MS

| Compound | Retention index ^a | Percentage |
|------------------------|------------------------------|------------|
| Thujene | 930 | 0.19 |
| Alpha-pinene | 937 | 4.26 |
| Beta-pinene | 976 | 0.43 |
| Beta-myrcene | 985 | 0.85 |
| Eucalyptol | 1024 | 3.37 |
| Gamma-terpinene | 1055 | 7.34 |
| Linalool | 1090 | 0.68 |
| Thymyl methyl ether | 1236 | 0.47 |
| Carvacrol methyl ether | 1243 | 0.46 |
| Carvacrol | 1299 | 71.12 |
| Trans-caryophyllene | 1418 | 0.41 |
| Globulol | 1582 | 2.32 |
| Sum | | 91.90 |

^a Retention index on DB5 column .

The yield of the essential oil of air-dried aerial parts of the representative sample of *Z. multiflora* Boiss. was 1.66% (v/w). Percentages of components of the essential oil (as determined by GC-MS) are summarized in Table 3. GC-MS analysis resulted in the identification of 12 components representing 91.9% of the oil. The major compound was phenolic

monoterpene carvacrol (71.1%). Other important compounds were gamma-terpinene (7.34%), alphapinene (4.26%) and eucalyptol (3.37%). Table 4 shows, 1,8-eucalypol was the most abundant individual compound in *E. globulus* oil followed by α -pinene (9.22%), α -terpineol acetate (3.1%) and globulol (2.77%).

Table 4: Essential oil chemical constituents of *E. globulus* identified by GC-MS

| Compound | Retention time (min) | Percentage |
|------------------------------|----------------------|------------|
| 1,8-eucalypol | 20.67 | 72.71 |
| α - terpineol | 17.87 | 2.54 |
| Terpinen-4-ol | 19.44 | 0.34 |
| linalool | 24.56 | 0.24 |
| α - pinene | 14.34 | 9.22 |
| β -pinene | 15.89 | 0.40 |
| α - eudesmol | 26.54 | 0.39 |
| -globulol | 30.74 | 2.77 |
| epiglobulol | 32.89 | 0.44 |
| α - terpineol acetate | 23.21 | 3.10 |
| geranyl acetate | 32.23 | 0.71 |
| L-pinocarveol | 30.24 | 0.36 |
| β -sabinen | 19.44 | 0.25 |
| terpinolene | 22.19 | 0.19 |

Antibacterial activity

The in vitro antibacterial activity of essential oil and extracts of *R. officinalis*, *A. graveolens*, *Z. multiflora* and *E. globules* against *S. iniae* was qualitatively assessed by the presence or absence of inhibition zones. According to the results given in Table 5, extracts of *R. officinalis*, *A. graveolens*, *Z. multiflora* and *E. globules* showed antibacterial effect against *S. iniae* with their respective diameter zones of

inhibition of 30, 21, 18 and 16.8 mm whereas the essential oil showed interesting antibacterial effect with inhibition zones in the range of 45-18 mm, respectively. All the tested essential oils showed the antibacterial activity. The best antibacterial activity was observed with the essential oils of *R. officinalis* with larger diameter zones of inhibition, 45 mm (Table 5) and smallest MIC 3.9 µg/mL values against *S. iniae*.

Table 5: Inhibition zone diameters (mm)^a in well diffusion assays of the *R. officinalis*, *Z. multiflora*, *A. graveolens* and *E. globulus* against *S. iniae*

| Plant | <i>R. officinalis</i> | | <i>Z. multiflora</i> | | <i>E. globulus</i> | | <i>A. graveolens</i> | |
|--------------------|-----------------------|----------|----------------------|----------|--------------------|----------|----------------------|----------|
| | Essential oil | extracts | Essential oil | extracts | Essential oil | extracts | Essential oil | extracts |
| Zone of inhibition | 45 ±1.2 | 30±0.9 | 22±0.7 | 18±0.6 | 18±1.0 | 16.8±1.3 | 32±1.1 | 21±0.8 |

^a Each result is the mean ± SD of three replicates.

Minimum inhibitory concentrations (MIC) and minimum bactericidal concentrations (MBC) of the oils and extracts were determined using a broth microdilution method. The variability in the concentration of the main components present in the essential oils analysed led us to evaluate the antimicrobial activities of

the oils and extracts. As shown in Table 6, the MIC values for the oils and extracts of *R. officinalis*, *A. graveolens*, *Z. multiflora* and *E. globules* were found to be in the range of 3.9–250 and 7.8- 500 µg/ml and the MBC values for the oils and extracts were found to be in the range of 7.8-250 and 15.6-500 µg/ml, respectively.

Table 6: Minimum inhibitory concentration (MIC) and minimal bactericidal concentration (MBC) (µg / mL) of essential oil and extracts of *R. officinalis*, *A. graveolens*, *Z. multiflora* and *E. globulus*

| <i>S. iniae</i> | <i>E. globulus</i> | | | | <i>A. graveolens</i> | | | |
|-----------------|-----------------------|-------|---------------|-------|----------------------|-------|---------------|-------|
| | Extract | | Essential oil | | Extract | | Essential oil | |
| | MIC | MBC | MIC | MBC | MIC | MBC | MIC | MBC |
| | 500.0 | 500.0 | 250.0 | 250.0 | 31.2 | 31.2 | 7.8 | 15.6 |
| | <i>R. officinalis</i> | | | | <i>Z. multiflora</i> | | | |
| | Extract | | Essential oil | | Extract | | Essential oil | |
| | MIC | MBC | MIC | MBC | MIC | MBC | MIC | MBC |
| | 7.8 | 15.6 | 3.9 | 7.8 | 250.0 | 500.0 | 62.4 | 250.0 |

All tested essential oils are bactericidal and showed MBC values almost equal to MIC values.

Discussion

Although antibiotics have been used to control bacterial infections in fish for many years, several antibiotics have been banned from use in aquaculture due to their adverse effects. In addition, products from aquaculture facilities using antibiotics are not accepted by many countries (Park et al., 2009). Therefore, finding an alternative antimicrobial substance to replace antibiotics has become an issue of interest of many research groups. In cultured fishes, streptococcal infections often develop into lethal septicaemic conditions. Fish in both freshwater and marine environments may be affected, and farms in many parts of the world have consequently suffered serious economic losses. Over the past few years, the Gram-positive bacterium *Streptococcus iniae* has been associated with outbreaks of disease in several species of farmed freshwater and marine fishes (including rainbow trout, tilapia, hybrid striped bass and yellowtail) in the world. The organism has also been isolated from diseased humans. Vaccination was successfully used in rainbow trout farms, but outbreaks resumed thereafter. In particular, such information would be valuable for operation planning, and for the control and prevention of such fish pathogens entering into water column (AL-Bulushi et al., 2010).

Plant-derived essential oils due to their antimicrobial content possess potential significance as naturally occurring agents for food preservation.

Many volatile compounds naturally occurring in various essential oils possess strong antibacterial activities, thereby considered as natural antibacterial agents to inhibit the growth of food-borne pathogens. The renewal of interest in food science and technology, and increasing consumer demand for effective natural products means that quantitative data on plant based essential oils is required. The use of essential oils may improve food safety and overall microbial quality. If essential oils were to be more widely applied as antibacterials in foods, the organoleptic impact would be important. On the other hand, volatile oils, which often contain the principal aromatic and flavoring components of herbs and spices, even added into small quantity to foodstuffs, without affecting organoleptic properties, would retard bacterial contamination and therefore reduce the onset of spoilage. For practical addition of essential oils in food, it is important to know if oils at very low concentrations, have efficient antimicrobial effects without affecting sensory qualities (Holley and Patel, 2005).

Essential oils of *R. officinalis*, *A. graveolens*, *Z. multiflora* and *E. globulus* have been reported to inhibit a broad spectrum of microorganisms (Ghalem and Mohamed, 2008; Kaur and Arora, 2009; Sarikurkcu et al., 2010; Zaouali et al., 2010). The EOs evaluated in this study displayed a variable degree of antimicrobial activity against *S. iniae*. Differences in their chemical profile were observed as well. Regarding this concern, a number of studies on *R. officinalis*, *A. graveolens*, *Z. multiflora* and *E. globulus*

EO composition, which reveal a great variability in their chemical profile, have already been published. According to these reports, some components are currently encountered in all species, however in variable amounts (Abed, 2007; Moosavy et al., 2008; Wang et al., 2008; Khan et al., 2009).

As the data of the in vitro antimicrobial activity show, using the filter paper disc agar diffusion technique, the different essential oils inhibited growth to variable extents, depending on the essential oil and the bacterium assayed. Nevertheless, more precise data on the antimicrobial properties of the four studied essential oils were obtained through the determination of the minimum inhibitory concentration and minimum bactericidal concentration. As can be seen in Table 6, the essential oil with the highest antibacterial properties was *R. officinalis* followed by *A. graveolens* while *E. globulus* was the least active essential oil.

The MIC and MBC of essential oils in this study were similar to the known literature with a little difference, which could be for several reasons such as a different growing environment, different extraction methods of essential oils, and so on. Different essential oils have different antimicrobial activity because of their components (Celiktas et al., 2007; Szumny et al., 2010; Ozcan and Arslan, 2011).

The essential oils exhibited antibacterial activity against *S. iniae*. The essential oil of rosemary showed the strongest antimicrobial activity. Also, this study provided comparable quantitative antimicrobial data for four essential oils. The diameter of the zone of inhibition

adjacent to essential oils was estimated 45, 32, 22 and 18 mm, respectively (Table 5). The extracts of different plant resulted in variable zone of inhibition (30.0–16.8 mm) for *S. iniae*. The results showed that these essential oils can strongly prevent the growth of *S. iniae* GQ 850377.

However, MIC of the *R. officinalis* essential oils and diameter of zone of inhibition in comparison to other essential oils showed potent antimicrobial activity and inhibitory effect. This suggested *R. officinalis* as an effective antimicrobial agent among all kinds of tested herbs.

1,8-cineole acts as a main substance of the rosemary oil. The obtained results are in line with other studies (; Nieto et al., 2010; Zaouali et al., 2010; Jalili-Heravi et al., 2011; Visentín et al., 2011). Also, this activity could be attributed to the presence of oxygenated mono- and sesquiterpenehydrocarbons, and these finding are in agreement with the previous reports (Frutos and Herná ndez-Herrero, 2005; AL-Reza et al., 2009; Weerakkody et al., 2010). Among all the essential oils evaluated, *E. globulus* exhibited comparatively weaker antibacterial activity. Such differences can be due to distinct plant origins and essential oil chemical composition. It is generally accepted that essential oils having higher contents of oxygenated terpenes exhibit potent antibacterial potential (Hussain et al., 2011).

The results obtained showed that the essential oils had greater activity, whereas the extracts were found to be less efficient in radical scavenging and lipid peroxidation inhibition (Table 6). In general, extracts exhibited low

antimicrobial activities compared to the essential oils. The oil and the extract, at all concentrations, exhibited higher activities than the control; the differences were significant ($p < 0.01$). But additional investigations need to be performed in order to confirm the safety of these concentrations (MIC) for human consumption. Furthermore, the MBC/MIC ratio is clearly higher than 1, indicating a bacteriostatic effect of the essential oil.

In addition, this study showed that *S. iniae* was more sensitive to essential oil of *R. officinalis* than other essential oils. The MBC of *R. officinalis* was generally lower than the others three oils.

When comparing the data obtained in different studies, most publications provide generalization about whether or not a plant oil or extract possesses activity against Gram-positive and Gram-negative bacteria.

Among all oils analyzed in this work, the essential oil of rosemary was the most effective as an antibacterial agent. The antibacterial activity has been attributed to the presence of some active constituents in the oils. Sensitivity of Gram-positive bacteria to essential oil of rosemary is in line with an earlier study (Oussalah et al., 2006; Kaur and Arora, 2009; Panahi et al., 2011; Teixeira et al., 2012).

Zaouali et al. (2010) showed that *R. officinalis* have more inhibitory effects on Gram - positive bacteria than Gram - negative bacteria, which is probably because of the different cell wall membrane structures between Gram-positive and Gram-negative bacteria. Takahashi et al. (2004) showed of the 26

tested bacterial species, the extracts of *E. globulus*, *E. maculate* and *E. viminalis* significantly inhibited the growth of Gram-positive bacteria but these extracts did not show strong antibacterial activities against Gram-negative bacteria.

The essential oil content of different plants varies (together with the biologically active compounds contained) depending on which part of the plant it is obtained from (flower, stem, leaves, whole plant, etc.), the variety of the plant, its harvest season, and the method of cultivation.

These differences might have been derived from local, climatic and seasonal factor. The method of extracting the plant material and its form (whether crushed or finely powdered) affected their antibacterial activity. Essential oils rich in phenolic compounds are widely reported to possess high level of antimicrobial activity which has been confirmed and extended in the present studies. It is believed that the phenolic components of essential oils show strongest antimicrobial activity, followed by aldehyde, ketones and alcohols (Jarrar et al., 2010). However, if plant oils are used for food preservation, issues of safety and toxicity will need to be addressed. Also, it should be undertaken in fish and processed fish to confirm the antimicrobial efficiency level of these essential oils. These essential oils could serve as potential antimicrobial agents to inhibit spoilage growth in food.

Further studies should evaluate the safety and toxicity of *R. officinalis*, *A. graveolens*, *Z. multiflora* and *E. globulus* extracts and essential oils to human consumption before considering their use

for food preservation or medicinal purposes.

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