In vitro antibacterial effect of ginger (Zingiber officinale) essential oil against fish pathogenic bacteria isolated from farmed olive flounder (Paralichthys olivaceus) in Korea

Hossain S.¹; De Silva B.C.J.¹; Wimalasena S.H.M.P.¹; Pathirana H.N.K.S.¹; Heo G.J.^{1*}

Received: November 2017 Accepted: July 2018

1-Veterinary Medical Center and College of Veterinary Medicine, Chungbuk National University, Cheongju, 28644, Korea

Keywords: *In vitro* antibacterial activity, Ginger essential oil, Fish pathogenic bacteria, Olive flounder

Introduction

World population growth and food demand have been two major elements leading to the expansion of animal and fish production. Recently, many efforts advocated the extension of intensive fish farming to optimize the need for animal protein (Hussein et al., 2013). In Korea, aquaculture has made rapid development in a short period of time, and many fish farms have been established in order to fulfill high consumer demand (Oh et al., 2006). Meanwhile. olive flounder (Paralichthys olivaceus) is one of the popular marine fishes cultured in Korea. which accounts approximately 50% of the annual fish production (Park et al., 2012).

Edwardsiellosis and streptococcosis are the most prevalent bacterial diseases in olive flounder which are responsible for high stock mortality. *Streptococcus*

parauberis, S. iniae and Edwardsiella tarda are the etiological agents of streptococcosis and edwardsiellosis, respectively (Park et al., 2014). Generally, antibacterials are applied for the treatment of bacterial diseases. However, the overuse and misuse of antibacterials cause different problems such as the risk of growing bacterial resistance, accumulation of hazardous residues in fish products and severe damage to aquatic ecosystems (Chenia, 2016).

Nowadays, there is a growing interest in the screening of plant extracts and essential oils for their antibacterial properties against fish pathogens, thereby exploiting natural antimicrobials to control bacterial diseases (Bulfon *et al.*, 2014). Natural substances are eco-friendly and cheap, so they could alleviate many of the side-effects that are often related to

^{*}Corresponding author's Email: gjheo@cbu.ac.kr

synthetic antimicrobials. Essential oils (EOs) have widely been used to control bacterial diseases in humans (Inovye *et al.*, 2001).

Ginger (Zingiber officinale) is a member of the family Zingiberaceae. Ginger essential oil (GEO) extracted from ginger rhizome has been studied for biochemical and pharmacological properties, and it was reported that the major constituents are zingiberene, βsesquiphellandrene and ar-curcumene (Nampoothiri etal., 2012). The fragrance and flavoring substance of GEO composing 1-3% of the weight of fresh ginger primarily consists of zingerone, shogaols and gingerols as the major pungent compounds. The biologically active compounds of GEO are mainly gingerols and shagols which have both antibacterial and antifungal properties (Supreetha et al., 2011; Shehata et al., 2013). Moreover, GEO has some other biological properties such as anti-oxidative, anti-cancer, antiparasitic, larvicidal, anti-diabetic, antiinflammatory and nephro/hepatoprotective activities (Kumar et al., 2011). GEO is effective against fishborne bacteria such as Aeromonas hvdrophila. Staphylococcus aureus. Listeria monocytogenes, Yersinia enterocolitica, Lactococcus garvieae and S. parauberis (Nya and Austin, Rattanachaikunsopon 2009: Phumkhachorn, 2009; Debbarma et al., 2012; Kim et al., 2016).

However, the antibacterial activity of GEO against fish pathogenic bacteria has not been studied extensively. Besides, it has not been assessed against any fish bacterial pathogens in

Korea. Therefore, this study aimed to investigate the in vitro antibacterial activity of GEO against fish pathogenic bacteria isolated from farmed olive flounder in Korea by determining the susceptibility with disk diffusion. inhibitory minimum concentration minimum (MIC) and bactericidal concentration (MBC) tests.

Materials and methods

Selection of bacterial strains

Five gram-negative and nine grampositive bacterial strains isolated from farmed Korean olive flounder (P. olivaceus) were used in this study. Gram-negative bacterial strains [E. tarda (FP5060, ED47, Yoshida and ED45) and Photobacterium damselae (FP4101)] and Gram-positive bacterial strains [L. garvieae (FP5245), S. iniae (FP5228, S186, S530 and S131) and S. parauberis (FP3287, S124, S527 and S1466)] collected from were Gyeongsang National University (Jinju, Korea) and National Institute of Fisheries Science (Busan, Korea). 100% pure ginger (Z. officinale) oil Ltd.. (Aromarant Co. Rottingen. Germany) which was purified by steam distillation from the rhizomes of ginger grown in India was purchased for this study.

Disk diffusion test with GEO

The antibacterial activity of GEO was determined by means of disk diffusion test. Every bacterial culture was adjusted to 0.5 McFarland standard (1.5 x 10⁸ CFU mL⁻¹) with sterile saline. The bacterial suspensions were spread over Mueller-Hinton agar (MHA) (MB

Cell, LA, CA) plates using a sterile cotton swab. Different concentrations of GEO (1:1= pure oil, 1:2, 1:5 and 1:10; 1 part of the GEO in respective parts of the solution) were prepared using dimethyl sulfoxide (DMSO) (OCI Co. Ltd, Seoul, Korea). Sterilized paper disks of 6 mm diameter (Advantec Toyo Kaisha, Ltd., Japan) were impregnated with 20 µL of different concentrations of GEO and placed on a MHA plate smeared with the bacterial strains. DMSO was used as the negative control and amoxicillin (10 µg) was used as the positive control. The plates were incubated for 48 h at 27°C and the diameter of the inhibition zone was measured in mm.

Multiple antibiotic resistance (MAR) index

The antimicrobial susceptibility was examined following the standards of Clinical and Laboratory Standards Institute (CLSI, 2014). A total of eleven antimicrobials including ampicillin (10 μg), cefotaxime (30 μg), ceftriaxone (30 tetracycline (15 chloramphenicol (30 µg), colistin (10 μg), gentamicin (10 μg), amikacin (30 μg), vancomycin (30 μg), nalidixic acid (30 µg) and ofloxacin (5 µg) (MB Cell, LA, CA) were used in this study for Gram-negative bacteria and nine antimicrobials including ampicillin (10 μg), cefotaxime (30 μg), ceftriaxone (30 tetracycline μg), (15)μg), chloramphenicol (30 µg), vancomycin (30 μg), ofloxacin (5 μg), erythromycin (15 µg) and clindamycin (5 µg) (MB Cell, LA, CA) were used for Grampositive bacteria. Multiple antibiotic resistance (MAR) index was calculated as the ratio of the number of antimicrobials to which the bacterial strain was resistant to the total number of antimicrobials to which the bacterial strain was exposed.

Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) tests

The MICs of GEO were determined by broth microdilution method in 96-well microtiter plates using different concentrations [8%, 4%, 2%, 1%, 0.50%, 0.25%, 0.125%, 0.063% and 0.31% (V/V)] (Fournomiti et al., 2015). The MICs were measured after incubation at 27°C for 48 h. For measurement of the MBC, 10 µL of medium from wells with no visible growth were transferred to tryptic soy agar (MB Cell, LA, CA) plates and incubated for 48 h at 27°C.

Results and discussion

Antibacterials have generally been used bacterial control diseases aquaculture for a long time. However, the frequent use of these drugs causes bacterial resistance. Recently, demand for using plant natural products replace antibacterial drugs has increased. The use of plant products especially essential oils for treating bacterial diseases in fish has become because of their popular antibacterial activities. Therefore, the present study was conducted determine the efficacy of GEO against pathogenic bacteria isolated from olive flounder.

According to the disk diffusion test results, GEO inhibited the growth of all bacteria except E. tarda. The inhibition zone diameters increased with the increasing concentrations of GEO. S. iniae strains showed high susceptibility and the highest inhibition zone (23 mm) was observed in 1:1 dilution of GEO (Table 1). However, all E. tarda strains were resistant both in disk diffusion and MIC tests. In the MIC test, each of the E. tarda strains was observed to have MIC >8% which was the highest MIC value. In a previous study, pathogenic E. tarda was resistant to GEO (Yoon et al., 2011). In contrast, GEO has been reported as an active agent against E. tarda infected Gourami fish (Osphronemus goramy) (Sarjito and Prayitno, 2015). The resistance of E. tarda in the present study could be due to the different composition of the chemical components Generally, the antibacterial activity of EOs is dependent on the constitution of EOs and the bacterial strains (Shehata et al., 2013). There are many variations in the chemical composition of GEO due to the environmental and genetic factors which make the antimicrobial action of GEO differ from that of bacteria (Snoussi et al., 2016). Similar to previous reports, only a few E. tarda strains employed in the present study deter the accurate conclusion about the antibacterial activity of GEOs against E. tarda. In addition, two E. tarda isolates were resistant to multiple antimicrobials and displayed high MAR index values. The reason for such resistance could be because E. tarda is intrinsically resistant to macrolides, lincosamides. streptogramins, glycopeptides and fusidic acid due to the presence of different proteins in the outer membrane including porin and efflux pump systems hindering the antimicrobials from entering the cells (Stock and Wiedemann, 2001; Peng et al., 2017).

Table 1: Inhibition zone with different GEO concentrations against fish pathogenic bacteria.

	Inhibition zone ^a (mm) with different GEO concentrations ^b					Amoxicillin (+ve
Bacterial strain					DMSO	
	1:1	1:2	1:5	1:10	(-ve control)	control)
Gram-negative bacteria						
Edwardsiella tarda (FP5060)	NA	NA	NA	NA	NA	27
E. tarda (ED45)	NA	NA	NA	NA	NA	24
E. tarda (ED47)	NA	NA	NA	NA	NA	24
E. tarda (Yoshida)	NA	NA	NA	NA	NA	27
Photobacterium damselae (FP4101)	8	6.5	NA	NA	NA	27
Gram-positive bacteria						
Lactococcus garvieae (FP5245)	13	9	8	NA	NA	25
Streptococcus iniae (S186)	19	15	13	10	NA	35
S. iniae (S530)	18	13	10	9	NA	38
S. iniae (S131)	23	20	15	12	NA	35
S. iniae (FP5228)	13	8	7	NA	NA	28
Streptococcus parauberis (S124)	9	6.5	NA	NA	NA	25
S. parauberis (S527)	9.5	6.5	NA	NA	NA	29
S. parauberis (S1466)	7	NA	NA	NA	NA	27
S. parauberis (FP3287)	13	8	7	NA	NA	28

^aInhibition zone; NA= No growth inhibition

MIC results of GEO tested for fish pathogenic bacteria ranged from 0.25 to

4% except for *E. tarda* (V/V) (Fig. 1). The highest MIC value of 4% (V/V)

^bConcentrations; 1:1= pure oil, 1:2, 1:5, 1:10= 1 part of GEO in respective parts of the dilution.

was observed in S. parauberis and the lowest MIC value of 0.25% (V/V) was observed in S. iniae strains (S131 and S186) among the gram-positive bacteria. In a recent study, GEO was active against fish pathogenic S. parauberis (Kim et al., 2016). A Gramnegative bacterial strain, P. damselae (FP4101) used in this study was sensitive to GEO in disk diffusion and MIC tests (MIC value 4%). According to the MIC results, GEO was effective against the majority of Gram-positive bacteria strains even lower concentrations compared to gramnegative bacteria. The weak antibacterial activity could be attributed to the presence of an outer membrane in gram-negative strains which possesses hydrophilic polysaccharides, thereby hampering the diffusion of hydrophobic compounds of EOs (Inovye et al., 2001).

The majority of the tested strains had MBC/MIC as <4. This implies the bactericidal mode of action of GEO pathogenic against fish bacteria. Considering the ratio of MBC/MIC, the EOs which showed bacterial killing effects with MBC/MIC <4 are designated as bactericidal for the tested while **EOs** bacteria, the demonstrated inhibitory effects with MBC/MIC>4 designated are as bacteriostatic (Bulfon et al., 2014).

In antimicrobial disk diffusion tests, *S. iniae* (S186, S530 and S131) strains were susceptible to all of the tested antimicrobials whereas other strains were resistant to 1 or more

antimicrobials (Table 2). The MAR index values have been calculated as ≥0.09. More importantly, two *E. tarda* (ED47 and ED45), one *L. garvieae* (FP5245), one *S. iniae* (FP5228) and two *S. parauberis* (S124, S1466) strains were denoted as high-risk strains as they had MAR index values of 0.45, 0.45, 0.44, 0.33, 0.44 and 0.22, respectively. An MAR index value of higher than 0.2 indicates that the bacteria were isolated from a source with a high risk of contamination where antimicrobials have often been used (Sandhu *et al.*, 2016).

Interestingly, exhibited **GEOs** antibacterial properties against the resisted tetracycline, strains that erythromycin and nalidixic acid which are common antimicrobials used in the Korean aquaculture industry (Park et al., 2014). It reveals the potentiality of GEO to be used as an alternative antibacterial agent against S. iniae, S. parauberis, L. garviae and P. damselae. Since E. tarda strains were resistant to GEO, further studies should be focused on determining the *in vitro* antibacterial activity of GEO against more strains of pathogenic E. tarda in fish. Besides, the number of species and the strains should be enhanced in order to acquire better understanding of the antibacterial property of GEOs. Also, the stability of GEO in aquatic environments, and the digestibility and the potential toxicity in fish should also be examined before the application of GEOs for control of fish bacterial infections in aquaculture systems.

Bacterial strain ^a	MAR index	No. of ineffective antimicrobials	Name of ineffective antimicrobials ^b	
Gram-negative bacteria				
Edwardsiella tarda (FP5060)	0.18	2	VA, CS	
E. tarda (ED45)	0.45	5	TC, CHL, VA, NAL, CS	
E. tarda (ED47)	0.45	5	TC, CHL, VA, NAL, CS	
E. tarda (Yoshida)	0.18	2	VA, CS	
Photobacterium damselae (FP4101)	0.09	1	VA	
Gram-positive bacteria				
Lactococcus garvieae (FP5245)	0.44	4	AMP, CTX, CRO, CHL	
Streptococcus iniae (FP5228)	0.33	3	CTX, CRO, DA	
Streptococcus parauberis (S124)	0.44	4	AMP, TC, E, DA	
S. parauberis (S527)	0.11	1	AMP	
S. parauberis (S1466)	0.22	2	AMP, E	
S. parauberis (FP3287)	0.11	1	AMP	
Bacterial strain ^a	MAR index	No. of ineffective antimicrobials	Name of ineffective antimicrobials ^b	

^a Bacterial strains are resistant to one or more antimicrobials

 $[^]b$ Antimicrobials abbreviation; AMP=ampicillin (10 μg), CTX=cefotaxime (30 μg), CRO=ceftriaxone (30 μg), TC=tetracycline (15 μg), CHL=chloramphenicol (30 μg), E=erythromycin (15 μg), DA=clindamycin (10 μg), VA=vancomycin (30 μg), NAL=nalidixic acid (30 μg) and CS=colistin (10 μg). Dsds

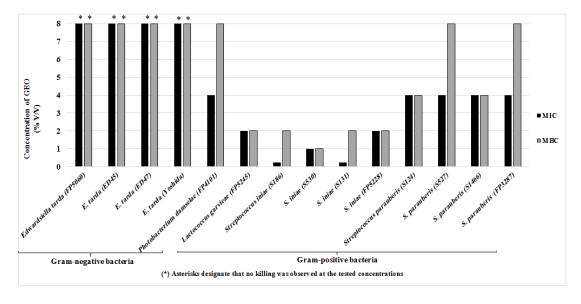


Figure 1: Comparison of MICs and MBCs of GEO against fourteen fish pathogenic bacterial strains.

Acknowledgements

The authors are thankful to Professor Tae-Sung Jung of the Institute of Animal Medicine, College of Veterinary Medicine, Gyeongsang National University (Jinju, Korea) and the National Institute of Fisheries Science (Busan, Korea) for providing the bacterial strains used in this study.

References

M., 2014. *In vitro* antibacterial activity of plant ethanolic extracts against fish pathogens. *Journal of the World Aquaculture Society*, 45, 545–557.

Chenia, H.Y., 2016. Prevalence and characterization of plasmid-mediated quinolone resistance genes in

- Aeromonas spp. isolated from South African freshwater fish. International Journal of Food Microbiology, 231, 26-32.
- CLSI, 2014. Performance standards for antimicrobial susceptibility testing of bacteria isolated from aquatic animals; second informational **CLSI** supplement. document VET03/VET04-S2. Wayne, PA: Clinical and Laboratory Standards Institute.
- Debbarma, J., Kishore, P., Nayak, B.B., Kannuchamy, N. and Gudipati, V., 2012. Antibacterial activity of ginger, eucalyptus and sweet orange peel essential oils on fish-borne bacteria. *Journal of Food Processing and Preservation*, 37, 1022–1030.
- Fournomiti, M., Kimbaris, A., Mantzourani, I., Plessas, S., Theodoridou, I., Papaemmanouil, V., Kapsiotis, I., Panopoulou, M., Stavropoulou, E., Bezirtzoglou, E.E. and Alexopoulos, 2015. A., Antimicrobial activity of essential oils of cultivated oregano (Origanum vulgare), sage (Salvia officinalis), and thyme (Thymus vulgaris) against clinical isolates of Escherichia coli, Klebsiella oxytoca and Klebsiella pneumoniae. Microbial Ecology in Health and Disease, 26, 23289.
- Hussein, M.M.A.H., Hassan, H.W. and Moussa, I.M.I., 2013. Potential use of allicin (garlic, *Allium sativum* Linn, essential oil) against fish pathogenic bacteria and its safety for

- monosex Nile tilapia (Oreochromis niloticus). Journal of Food, Agriculture and Environment, 11, 696-699.
- Inovve, S., Takizawa, J. Yamaguchi, H., 2001. Antibacterial activity of essential oils and their major constituents against respiratory tract pathogens by Journal gaseous contact. of Antimicrobial Chemotherapy, 47, 565-573.
- Kim, H.S., Kim, H.J., Choi, D.G., Jang, B., Cho, S.H., Kwon, M.G., Min, B.H. and Kim, D.S., 2016. Effect of various sources of dietary additives growth, on body composition, and challenge test juvenile survival of Rockfish Sebastes schlegeli. Turkish Journal of Fisheries and Aquatic Sciences, 16, 759-766.
- Kumar, G., Karthik, L. and Rao, K.V.B., 2011. A review on pharmacological and phytochemical properties of *Zingiber officinale* Roscoe (Zingiberaceae). *Journal of Pharmacy Research*, 4, 2963–2966.
- Nampoothiri, S.V., Venugopalam, V.V., Joy, B., Sreekuman, M.M. and Menon, A.N., 2012. Comparison of essential oil composition of three ginger cultivars from sub himalayan region. Asian **Pacific** Journal of**Tropical** Biomedicine, 2, 1347-1350.
- **Nya, E.J. and Austin, B., 2009.** Use of garlic, *Allium sativum*, to control *Aeromonas hydrophila* infection in rainbow trout, *Oncorhynchus mykiss*

- (Walbaum). *Journal of Fish Diseases*, 32, 963-970.
- Oh, M.J., Jung, S.J., Kitamura, S.I., Kim, H.J. and Kang, S.Y., 2006. Viral diseases of olive flounder in Korean hatcheries. *Journal of Ocean University of China*, 5, 45-48.
- Park, J.W., Lee, Y.M., Noh, J.K., Kim, H.C., Park, C.J., Hwang, I.J., Kim, S.Y. and Lee, J.H., 2012. The morphological study of wild and farmed olive flounder (*Paralichthys olivaceus*): The role of indirect selection within and between populations. *Development and Reproduction*, 16, 309-314.
- Park, S.B., Kwon, K., Cha, I.S., Jang, H.B., Nho, S.W., Fagutao, F.F., Kim, Y.K., Yu, J.E. and Jung, T.S., 2014. Development of a multiplex PCR assay to detect Edwardsiella tarda, Streptococcus parauberis, and Streptococcus iniae in olive flounder (Paralichthys olivaceus). Journal of Veterinary Science, 15, 163.
- Peng, B., Wang, C., Li, H., Su, Y.B., Ye, S.Z., Yang, M.J., Jiang, M. and Peng, X.X., 2017. Outer membrane proteins from specific patterns in antibiotic-resistant *Edwardsiella tara*. *Frontiers in Microbiology*, 8, 69.
- Rattanachaikunsopon, P. and P., Phumkhachorn, 2009. Protective effects of olive oilsupplemented fish diets experimental Lactococcus garvieae infection in tilapia. Bioscience,

- *Biotechnology and Biochemistry*, 73, 2085-2089.
- Sandhu, R., Dahiya, S. and Sayal, P., 2016. Evaluation of multiple antibiotic resistance (MAR) index and doxycycline susceptibility of *Acinetobacter* species among inpatients. *Indian Journal of Microbial Research*, 3, 299-304.
- Sarjito, L.A.P. and Prayitno, S.B., 2015. The short bathing effect of red ginger (*Zingiber officinale* var. rubrum) extract on the survival rate and growth rate of gourami (*Osphronemus gouramy*) infected by *Edwardsiella tarda*. *Journal of Aquaculture Management and Technology*, 4, 31-37.
- Shehata, S.A., Mohamed, M.S. and El-Shafi, S.A., 2013. Antibacterial activity of essential oils and their effects on Nile tilapia fingerlings performance. *Journal of Medical Sciences*, 13, 367-372.
- Snoussi, M., Trabelsi, N., Taleb, S.B., Dehmeni, A., Flamini, G. and Feo, V.D., 2016. Laurus nobilis, Zingiber officinale and Anethum graveolens essential oils: Composition, antioxidant and antibacterial activities against bacteria isolated from fish and shellfish. Molecules, 21, 1414.
- Stock, I. and Wiedemann, B., 2001.

 Natural antibiotic susceptibilities of Edwardsiella tarda, E. ictaluri, and E. hoshinae. Antimicrob.

 Antimicrobial Agents and Chemotherapy, 45, 2245–2255.

Supreetha, S., Sharadadevi, M., Sequeira, P., Simon, J., Shreyas, T. and Amit, M., 2011. Antifungal activity of ginger extract on *Candida Albicans*: An *in-vitro* study. *Journal of Dental Sciences and Research*, 2, 1-5.

Yoon, G.H., Jufaili, A.S., Ghabshi, A.A. and Mazrooei, N.A., 2011. The *in vitro* activity of a range of natural bioflavonoids against five species of pathogenic fish bacteria. *Agriculture and Marine Sciences*, 16, 41-50.