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The effects of lucantin red, yellow and astaxanthin on growth, hematological, immunological parameters and coloration in the Tiger Oscar (*Astronotus ocellatus* Agassiz,1831)

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Abstract

The present study aimed to determine the effects of lucantin red and yellow and also astaxanthin on the growth, hematological and immunological parameters, and coloration of the tiger oscar (Astronotus ocellatus). To this end, 120 fingerlings of tiger oscar were equally and randomly divided into 12 aquariums. The fish were fed with experimental diets for 8 weeks. Treatments included diets containing no pigment (control), 100 mg kg⁻¹ lucantin red, 100 mg kg⁻¹ lucantin yellow, and 100 mg kg⁻¹ astaxanthin, with three replicates apiece. At the end of the trial, growth factors, hematological and immunological parameters, and coloration were measured and the obtained data were statistically analyzed. The results showed that there is a significant difference between the control group and other treatments fed with pigments in terms of growth factors (p<0.05). However, no significant difference was found between treatments fed with pigments in this regard (p>0.05). The results indicated no significant difference between treatments in hematological parameters (p>0.05). The results demonstrated that there is a significant difference between the control group and other treatments in terms of lysozyme activity. In addition, a significant difference was observed between treatments 1 and 3 in this regard (p<0.05). However, there was no significant difference between treatments in other immunological parameters (total immunoglobulin, IgM). The statistical analyses suggested no significant difference between experimental treatments in terms of total carotenoids in the blood (p>0.05). Statistical analysis of color intensity variation revealed a significant difference between the control group and other treatments (p<0.05). The study findings showed that lucantin red and yellow and astaxanthin can improve growth factors and coloration and increase lysozyme activity in Tiger Oscar.

Keywords: Astaxanthin, Astronotus ocellatus, Lucantin, Coloration, Blood

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Introduction

The fish color is an important indicator in the aquaculture industry (Li et al., 2014). Along with the growth of aquaculture in the world, the industry of ornamental fishes propagation and rearing expanding (Erdogan et al., 2012; Guroy et al., 2012). The color of ornamental fishes is one of the most important qualitative criteria in determining their market value. Accordingly, a group of carotenoid pigments may grow in the skin of the fish influenced by using of special diet (Sornsupharp et al., 2015). A wide range of artificial and natural carotenoids are used as nutritional supplements improve skin aquaculture to color (Storebakken and No. 1992: Paripatananont et al., 1999; Gouveia et al., 2003; Hancz et al., 2003; Chien and Shiau, 2005; Kalinowski et al., 2005). As natural antioxidants, carotenoids play a major role in the health of living organisms by neutralizing the free radicals caused by the normal activity of cells and environmental stress (Chew, 1995). Various factors affect the coloration of the fish such as fish size and age, fecundity course. composition, the source and concentration of carotenoid, sexual maturity, and genetic factors. In addition, water temperature, salinity, and light intensity environmental parameters that influence fish coloration (Torrissen et al., 1989; Storebakken and No, 1992). Astaxanthin is the most important carotenoid pigment found in aquatic organisms (Christiansen et al., 1996; Guerin et al., 2003) and controls some biological functions including the prevention of the oxidation essential unsaturated fatty protection against negative effects of ultraviolet light, vitamin A generation, immunity reactions, bio-coloring, improvement of growth and reproductive behaviors (Torrissen et al., 1989; Lorenz and Cysewski., 2000). Mammals and most fishes are not able to synthesize the carotenoids they need and crustaceans also have a limited ability to convert other pigments to astaxanthin. Therefore, it is advisable that this pigment is directly added to the diet (Meyers, 1977; Guerin et al., 2003). The Tiger Oscar (Astronotus ocellatus) is a large, boldly colored South American cichlid characterized by its unique personality and striking intelligence. It is considered a popular aquarium fish in Europe and the U.S.A (Staeck and Linke, 1985; Loiselle, 1995; Keith et al., 2000). Lucantin red and yellow are other types of carotenoid pigments. The present study aims to determine the effect of three pigments on hematological factors. immunological parameters, and coloration of the tiger oscar.

Materials and methods

In this study, 120 fingerlings of tiger oscar, with a mean weight of 7.5±0.5 grams, were provided from an ornamental fishes propagation and rearing center in Guilan Province. The subjects were complete the same age and were the offspring of a single pair of brooders. After adaptation to the experimental environment, the subjects were randomly and equally assigned to 12 aquariums sized 50×40×33 (10 fish in each aquarium). Every 3 aquariums represent a single treatment with three replicates. Experimental treatments were as follows: -Control: Commercial feed without food additives

-Treatment 1: Commercial feed containing 100 mg per kg of lucantin red

-Treatment 2: Commercial feed containing 100 mg per kg of lucantin yellow

-Treatment 3: Commercial feed containing 100 mg per kg of astaxanthin

The base diet was Biomar feed which contained 54% protein, 18% raw fat, 10% cellulose, 10% ash, and 1.6% total phosphorus. The subjects were fed twice a day (morning and night) by 3% of their body weight for 8 weeks (Ghiasvand and Shapoori, 2006).

The pigments used in this research were purchased from **BASF** Company, Germany. Astaxanthin, 3,3'-dihydroxy-ßcarotene-4,4'-dione, was used as a purple powder with a particle size less than 0.6 mm and completely soluble in 40°C water. The chemical formula of Lucantine red and Lucantin yellow was 4,4'-dioxo-βcarotene and β-apo-8'-carotenoic acid ethyl ester, respectively. The desired amount of pigments was dissolved in 150 ml distilled water by an electromagnetic device (hot plate magnetic) and a magnet with a temperature of 50°C. Then, the resulting solution was sprayed on the food. The feed containing pigments was dried in a gentle drying machine, kept in a dark and cool place for a while, and then placed in a freezer until the end of the trial.

The mean water temperature, dissolved oxygen, pH, water hardness (DH), total ammonia before feeding, and ammonia after feeding during the trial were 27.15±1.32, 7.63±3.24, 8.0±3.33, 231.0±99.20, 0.1 mg L⁻¹, and 0.2 mg L⁻¹, respectively.

Measurement of growth factors and survival rate

The length of subjects was measured by using a caliper with a precision of one millimeter and their weight was measured by a digital scale with a precision of 0.001 gram. To determine the growth status of subjects and performance of different treatments, survival rate, body weight increase, feed conversion ratio, special growth rate, weight gain, condition factor and length were calculated using the following equations:

Survival rate (SVR)=(S-D)/S×100 (Mazurkiewicz *et al.*, 2008)

S=the number of subjects

D=the number mortalities

Body weight increase (BWI)= $100 \times (BW_1 - BW_0)/BW_0$ (Hung *et al.*, 1989)

 BW_0 = initial weight (g)

 BW_1 = final weight (g)

Feed conversion rate (FCR)= F/W_1-W_0) (Ronyai *et al.*, 1990; Abdelghany and Ahmad, 2002)

F=the amount of food consumed by the fish

 W_0 = the mean primary biomass (g)

 W_1 = the mean final biomass (g)

Special growth rate (SGR)= =[(LnW_1 - LnW_0) / t] ×100 (Ronyai *et al.*, 1990)

 W_0 =the mean primary biomass (g)

W₁=the mean final biomass (g)

t=time period (day)

Weight gain (WG)=final weight (g)—initial weight (g) (Ricker, 1979; Tacon, 1990)

Condition factor (K)= $W/L^3 \times 100$ (Ojolick *et al.*, 1995)

W= fish weight (g)

L= fish length (cm)

Length=final length(cm)-initial length(cm)

Hematological and Immunological parameters

hematological order measure parameters at the end of the trial, 3-6 fish from each treatment were randomly selected and anesthetized using the extract of clove powder (0.5 ml in one liter of water). Then, after drying the fish, blood sampling from the caudal peduncle were done using heparinized insulin syringes, and blood samples were poured into microtubes containing 20 µl heparin. The microtubes were numbered and prepared for hematological tests in the next steps (Thrall, 2004). For serological studies, blood samples in Eppendorf tubes without heparin were centrifuged at 300 rounds per minute for 10 minutes (Labofuge, Heraeus Sepatch, Germany). Then, the serum was separated, poured into new Eppendorf tubes using a sampler, and stored at -80°C. Red blood cells count (RBC), hematocrit (HCT), hemoglobin (HB), white blood cells count (WBC), differential white blood cells count (lymphocytes, eosinophils, neutrophils, and monocytes), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC) were the hematological parameters measured in this study (Klontz, 1994). To measure lysozyme, the method proposed by Clerton et al. (2001) was used, and immunoglobulin (IgM) and total immunoglobulin were measured by the method of Siwicki and Anderson (1993).

Measurement of total carotenoid and coloration

The method of Weber (1988) was used to measure total carotenoid. In addition, the procedure proposed by Yam and **Papadakis** (2004)used for was determining color variation in subjects. This method is based on the processing of images taken by a digital camera at the same light intensity and conditions. To this end, 6 fish were randomly selected from each replicate and anesthetized in the extract of clove powder. Then, some images of their left side were taken by a Cannon digital camera (12.1 pixels) from a distance of 30 cm. The intensity of colors was measured at caudal peduncle or R.O. area, which is a common point in all strains of oscar. Imaging was done in 4 feeding before with stages: diets containing pigments, 10 days after feeding with diets containing pigments, 20 days after the second imaging, and 27 days after feeding with diets containing pigments.

Data analysis

The Kolmogorov-Smirnov test was used to test the normal distribution of data and the homogeneity of variances was examined using Levene's test. In addition, one-way analysis of variance (ANOVA) was used for statistical analysis of data. The mean values were compared using ANOVA and Duncan's multiple range test in SPSS, at the 95% interval of confidence. Excel 2010 was used to draw charts.

Results

According Table 1, significant to difference was observed between the control and other treatments fed with pigments in growth factors (survival rate, weight gain, FCR, SGR, final weight, condition factor, and length increase) (p<0.05). The results of hematological and immunological parameters assessment have been presented in Tables 2 and 3.

Table 1: The mean values of growth factors (mean \pm SD) in different treatments. Different letters (a-d) indicate a significant difference (p<0.05).

Diets	SVR(%)	BWI(%)	FCR	SGR(%)	WG(g)	K	Length(cm)
Control	85 ± 0.7^{a}	101.77±1.51 ^a	1.85 ± 0.27^{b}	1.25±0.13 ^a	7.63 ± 1.13^{a}	2.96±0.27°	0.90±0.14 ^a
Treatment 1	$100\pm0^{\mathrm{b}}$	280 ± 1.32^{bc}	0.92 ± 0^{abc}	2.38±0.21 ^{bc}	21.02 ± 0.25^{bc}	2.53 ± 0.14^{ab}	3.30±0.21 ^{bc}
Treatment 2	100 ± 0^{b}	254 ± 1.90^{b}	1.1 ± 0.08^{c}	2.26 ± 0.09^{b}	19.12±1.42 ^b	2.51 ± 0.17^{ab}	3.08 ± 0.04^{b}
Treatment 3	100 ± 0^{b}	$271{\pm}1.34^{bc}$	0.90 ± 0.04^{abc}	2.34 ± 0.06^{bc}	20.36 ± 1.01^{bc}	2.4 ± 0^{ab}	3.41 ± 0.12^{bcd}

Table 2: The mean values of hematological parameters (mean \pm SD) in different treatments.

Diets	WBC (N×10 ³)	RBC (N×10 ⁶)	HB (g dl ⁻¹)	HCT (%)	MCV (fl)	MCH (pg)	MCHC (g dl ⁻¹)	Neutrophilis (%)	Lymphocytes (%)	Monocytes (%)	Eosinophils (%)
Control	5050±1.7	1.88 ± 0.53	7.55±0.07	7.55±0.07	173±0	40±0	23±0	26.5±0.7	70.5±1.36	6±1.41	0
Treatment 1	4550±2.37	1.81±0.40	7.4±0.28	7.4±0.28	168±2.82	41±0	24±0	26±1.41	69.5±2.12	3 ± 2.82	0
Treatment 2	4550±2.23	1.62±0.66	6.65±1.2	6.65 ± 1.2	172.5±2.77	40.5±0.7	23.5±0.7	22.5±3.53	72±2.82	5±0	0.5 ± 0.7
Treatment 3	5200±2.45	1.86±0.99	7.25±0.35	7.25±0.35	163.5±2.53	39±0	23.5±0.7	24.5±2.95	70.5±1.77	4.5±1.48	0.5 ± 0.7

In addition, there was no significant difference between treatments during the

trial (56 days) in terms of hematological parameters (p>0.05) (Table 2).

Table 3: The mean values of immunological parameters (mean \pm SD) in different treatments. Different letters (a-d) indicate a significant difference (p<0.05).

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Diets	IgM (ml dl ⁻¹)	Total Immunoglobulin (mg ml ⁻¹)	Lysozyme (µg ml ⁻¹)				
Control	12.75±2.47	19.75±0.35	19.5±3.53 ^a				
Treatment 1	14.6±0.56	20.5±1.12	$35{\pm}1.41^{abc}$				
Treatment 2	14.95±1.48	19±1.82	$45.5{\pm}1.67^{abcd}$				
Treatment 3	14.1±1.55	17.75±1.76	75 ± 1.72^{d}				

The highest lysozyme activity was related to the treatment fed with astaxanthin, and there was a significant difference between the control other treatments fed with astaxanthin (p<0.05) (Table 3).

Table 4 shows the results on the measurement total carotenoid and intensity variation of the main and primary skin colors in 4 steps.

Table 4: The mean values of total carotenoid and intensity variation of the main and primary skin colors (mean \pm SD) in different treatments. Different letters (a-d) indicate a significant difference (p<0.05).

Diets	Total Carotenoid (μg ml ⁻¹)	Stages	R	G	В	С	M	Y	K
Control	2.9±0.14	Stage 1	78.77±1.02ª	109.78±2.5ª	101±3.9ª	37.33±3.1ª	46.5±2.29a	48.66±1.2ª	28.66±2.1ª
		Stage2	85.66±5.56ª	83.88±3.02ª	60.99±1.4d	48.55±1.57 ^d	43.66±4.66ª	63.88±2.1ª	43.33±2.21ª
		Stage3	173±3.09ª	127±3.64 ^{cd}	32.66±1.71b	20.66±3.46 ^{cd}	56.44±1.40a	93.55±3.14ª	28.55±1.3°
		Stage4	120±1.73ª	80.33±1.33bc	35.22±3.51b	38.66±2.51d	50.77±1.89a	87.33±2.6ª	38.22±1.37b
Treatment 1	6.05±0.49	Stage 1	78.49±0.27ª	109.71±0.24a	101±0.91ª	37.54±0.72ª	46.5±0.7ª	48.57±0.45ª	28.43±0.18a
		Stage2	143±5.50bc	86.55±2.36abc	42.88±2.2abcd	28.11±1.68c	58.66±1.61bc	86.99±1.69bcd	24.77±2.5d
		Stage3	193.22±1.54b	79.88±2.11b	15.88±1.07ª	16.88±2.83¢	78.88±1.76°	100±0b	7.44±2.3b
		Stage4	176±1.14b	80.10±2.87¢	20.66±1.65ª	19.77±2.02abc	68.22±1.01¢	95.88±1.50b	12.55±1.89a
Treatment 2	6.75±1.62	Stage 1	78.52±0.33ª	109.08±0.24ª	101±0.84ª	37.60±0.92ª	46.48±0.04ª	48.28±0.55ª	28.79±0.32ª
		Stage2	154.66±6.19bcd	111.55±2.19d	27.21±2.71bcd	25.99±1.65bcd	48.22±2.66ab	90.22±2.67 ^{cd}	26.33±2.60d
		Stage3	211.88±1.34¢	120±1.44°	14.77±1.24ª	10.77±1.26ab	62.22±2.78b	99.66±0b	6.55±1.5ª
		Stage4	177±2.89b	112±2.69d	19.1±1.66ª	20.66±1.85bc	60.88±1.57a	97.44±1.27b	12.99±2.51a
Treatment 3	5.6±1.54	Stage 1	78.22±0.12ª	109.31±0.54ª	101.77±0.17ª	36.14±0.44ª	46.17±0.82ª	48.17±0.39ª	28.21±0.49a
		Stage2	174.55±1.85d	101.11±2.54bcd	49.66±2.65d	21.22±1.19ab	65.66±1.20 ^{cd}	85.99±1.33bc	13.44±1.96bc
		Stage3	190±2.98b	62.77±1.38a	15.44±1.84ª	17.77±1.46°	83.44±1.19d	99.88±1.19b	7.99±1.15ab
		Stage4	176±2.79b	68.66±1.6ab	29.99±2.3ª	20.55±1.07bc	82.21±1.5d	99.33±1.33b	11.44±1.57a

There was no significant difference between the control other treatments fed with pigments in total carotenoid of blood (p>0.05) (Table 4).

Variation in the intensity of primary colors (RGB) and secondary colors (CMYK) in the first stage of imaging (before beginning of feeding with pigmented food) In this stage of imaging, no pigmented feed was given to the fish. As a result, no significant difference was observed between treatments in the intensity of primary and secondary colors (p>0.05) (Table 4).

Variation in the intensity of primary colors (RGB) and secondary colors (CMYK) in the second and third stages of imaging (10 and 20 days after the beginning of feeding with pigmented food)

Since the fish were fed with pigmented food in the second and third stages of imaging, a significant difference was found between the control and treatments fed with pigmented food in the intensity of primary and secondary colors (p<0.05) (Table 4). In terms of color intensity, because of feeding the fish with pigmented food, the intensity of red (R) showed a significant difference from other primary colors (G and B) and primary colors (K, M, C). In other words, with the increase of red color in the skin, the intensity of blue, green, turquoise, and black somewhat reduces. This suggests that the use of pigmented food increased the intensity of red in caudal area of fish. In all stages of imaging, R and Y coefficients led to the highest increase in color intensity in this area.

Variation in the intensity of primary colors (RGB) and secondary colors (CMYK) in the fourth stage of imaging (27 days after the beginning of feeding with pigmented food)

There was a significant difference between the control and treatments fed with pigmented food (p<0.05) (Table 4). Given that feeding was stopped in this stage of imaging, the red color in the caudal area caused no significant difference between treatments, except for the treatment fed with lucantin yellow. This indicates that pigments are stored the tissue, muscle, and liver and their effect does not decline after the end of feeding with pigmented food.

Discussion

The survival rate in all treatments, except the control, was 100%. There was a significant difference between the control group and other treatments in this regard (p<0.05). The highest weight gain was related to treatment 1 and the lowest value was observed in the control group. There was a significant difference between the control and other treatments in terms of weight gain, while such a difference was not found between treatments 1, 2, and 3 (p>0.05). The lowest and the highest FCR belonged to the treatment fed with astaxanthin-containing diet and control, respectively. In this regard, a significant difference was found between the control and the treatment fed with the diet containing lucantin yellow. The highest and the lowest SGR were related to treatment 1 (2.38±0.21) and the control (1.25 ± 0.13) . There was a significant difference between the control and other treatments (p < 0.05). The highest final weight was found in the treatment fed with

the diet containing lucantin red and the lowest value was related to the control, which suggests a significant difference between the control and other treatments. In terms of condition factor, there was a significant difference between the control and other treatments. In addition, a significant difference was observed between the control and other treatments in terms increased length. Verakunpiriya et al. (1997) stated that pigment sources can improve growth factors. Prescription of 5% aloe vera causes a significant increase in weight, SGR, and condition factor and reduction of FCR in oscar (Saeedi et al., 2014). Beiranvand et al. (2015) showed that addition of spirulina powder to the diet of the Zebra Danio (Danio Hamilton, 1822) increased body weight and SGR. Amar et al. (2001) reported that carotenoids are able to improve the growth, the immune system, and survival rate of fish. Moreover, they can cause protection against harmful effects of fat oxidation (Liebler, 1993; Waagbo et al., 2003). There are different views about the effect of carotenoid pigments on growth factors of different fish species. While some results indicate that these pigments cannot improve growth factors in rainbow trout, Atlantic salmon, cod, Arctic char, and red porgy (Storebakken and Goswami, 1996; Barbosa et al., 1999; Mansour et al., 2006; Tejera et al., 2007; Sawanboonchun et al., 2008; Sheikhzadeh et al., 2012), the results of some other researchers suggest the positive effect of these pigments on growth improvement. This difference may be attributed to fish species, development stages, and type of carotenoid. Li et al. (2014) reported that diets containing

astaxanthin and Haematococcus pluvialis efficiently improved growth factors in large yellow croaker. Specifically, weight gain was significantly higher in fish fed with H. pluvialis than those fed with the diet containing astaxanthin. The accumulation of fat influenced by the total body fat and the condition factor can justify this finding. Christiansen and Torrissen (1995) stated that small amounts of astaxanthin (1.5 mg kg⁻¹) can lead to the highest growth and survival in fries of Atlantic salmon (Salmo salar Linnaeus, 1758). Christiansen et al. (1995) showed that feeding of Atlantic salmon and Salmon salar pars with semi-purified diets contain 60 mg kg⁻¹ astaxanthin for 315 days significantly improved their growth compared to the control group. In the present study, the subjects fed with diets containing pigments showed the highest BWI, SGR, WG, and increased length and the lowest FCR compared to the control. There was a significant difference between all treatments in terms of growth factors. It seems that the addition of these pigments to the diet of tiger oscars has been effective in the improvement of their growth.

The results of hematological parameters measurement at the end of the trial presented significant difference treatments in any of between parameters, although some of them underwent a reduction or increase during the trial. Regarding hematological parameters and blood cell indices, the results of this study were both consistent and inconsistent with the findings of previous studies. Rezaei et al. (2013) studied the effect of adding the extract of *Zhumeria majdae* (150,300,600 mg kg⁻¹)

the diet on growth factors and hematological and immunological parameters in catfish (Pangasianodon hypophthalmus) and concluded that the extract of this plant has no significant impact on hematological parameters. Ansarifard et al. (2018) studied the Effects of Arthrospira platensis on growth, skin color and digestive enzymes of Koi, Cyprinus carpio Their results indicated inclusion of 5-10% A. platensis in diets has a significant positive effect on growth rate, pigmentation, and improvement of digestive and liver enzymes activities in koi fish. Tukmechi et al. (2011) stated that some factors such as environmental factors, especially due to the coldness of the fish, seasons, salinity, photoperiod, temperature, density, physiological parameters, species, reproductive cycle, puberty status, age, gender, nutritional conditions, sampling time and method, and accuracy and sensitivity measurement methods can affect growth factors and survival and make a difference in the interpretation of researchers.

Some studies show that feeding of fish and poultry with Spirulina can increase body resistance to diseases, improve survival rate and the growth, and enhance the function of the immune system (Hayashi et al., 1998; Belay, 2002). Li et al. (2014) reported that increasing the levels of astaxanthin and H. pluvialis in diet of large yellow croaker (Pseudosciaena crocea) increases the activity of lysozyme. level of The lysozyme is one of the important indicators of innate immunity in fish (Saurabh and Sahoo, 2008). Previous studies on mammals show that lysozyme can be activated by diets containing

astaxanthin (Chew et al., 2011; Park et al., 2011).

The main objective of stimulants is to improve non-specific immunity. Due to the higher efficiency of the nonspecific immune system than the specific immunity (compared to warm-blooded animals), the effect of stimulants is more prominent in the immune system of fishes. Lysozyme is released into the blood by neutrophils and macrophages (Sakai, 1999). Chien et al. (2003) showed that astaxanthin can stimulate the immune system of juveniles the giant tiger prawn (Penaeus monodon). Generally, increased lysozyme activity in the serum and mucus of fish can be indicative of the immune system stimulation and improvement of the immune response. By enhancing the complement system and lysozyme, carotenoid pigments increase the total number of leukocytes and phagocytes and thereby cause the stimulation of the immune system, increased immunity, and resistance to pathogens. Wang et al. (2015) reported that dietary astaxanthin in **Pacific** white shrimp (Litopenaeus vannamei) significantly affects the hemolymph immunological index, including total haemocyte counts, phagocytic activity of haemocyte, serum anti-superoxide radical activity, serum phenoloxidase activity, serum antibacterial activity and serum bacteriolytic In the present study, subjects of activity. Treatment 3 presented the highest lysozyme activity compared to the control and treatments 1 and 2. Li et al. (2014) stated that the effect of astaxanthin and H. pluvialis on total immunoglobin of long yellow croaker is negligible. Previous studies on warm-blooded animals have

indicated that beta-carotene and canthaxanthin can improve the proliferation of T and B cells (Bendich and Shapiro, 1986). However, there is no evidence of the effect of carotenoids on the improvement of immunoglobin products.

Despite the increase of total carotenoid in treatments 1, 2, and 3 compared to the control, it was not statistically significant. Variation of the intensity of the primary and secondary colors on the skin was noticeable in all treatments during the four stages of imaging, and statistical analyses show a significant difference between the control and other treatments in this regard. Mashalchi et al. (2010) showed that diets containing astaxanthin and danualila changed the skin color into red in white oscars. Hassaninia et al. (2016) stated that the addition of lucantin pink to the diet of oscar can change their skin color and significantly increase the intensity of red in this fish. Choubert and Heinrich (1993) and Sommer et al. (1991) reported that high levels of sterilized astaxanthin and healthy spores with low absorbency in non-homogenized healthy algae produce lower total carotenoid in the skin and muscles of rainbow trout compared to astaxanthin homogenized and Bjerkeng (2000) stated that coloration of fish, in addition to biological factors, is influenced by pigments content of the diet, the concentration of pigments, duration of feeding, and diet composition. Pham et al. (2014) studied the effects of carotenoid sources of diet on the olive flounder (Paralichthys olivaceus) and concluded that the red color of this fish increases by feeding on diets containing carotenoids, but the brightness, clarity, and yellow color of the skin are not affected by carotenoid sources. Safari and Mehraban

Sang Atash (2015) showed that the extract of achiote (Bixa orellana) increases the level of carotenoids in the blood of rainbow trout. Moreover, they have been stated that there is a positive correlation between the carotenoid content of blood and final weight and SGR and a negative relationship between the carotenoid content of blood and FCR. This is consistent with the findings of the present study. Shapoori et al. (2012) believe that the difference in the color intensity caused by natural and artificial pigments are due to the quality, concentration, and the absorption period of these materials. Carotenoids are a major ingredient of diets but their distribution and intensity are probably influenced by genetic factors. Generally. chromatophores in fishes include melanophores, xanthophores, erythrophores, and iridophores that are responsible for the revealing of colors on the skin of fish. However, the pigment patterns in fish are affected by the different position of chromatophores. In addition, the pattern of pigment cells may be caused by patterning mechanisms away from local environmental guidance or the interaction between neighboring chromatophores (Kelsh, 2004). Sornsupharp et al. (2015) showed that diets containing carotenoids can increase the total carotenoid content of color in the skin and muscles flowerhorn cichlids. This is due to the concentration of carotenoids in the diet, which plays a major role in the skin coloring in this species. It seems that in the present study, treatment 1 has a better performance than other treatments.

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