Antioxidant properties of sodium acetate, sodium citrate and sodium lactate on lipid oxidation in rainbow trout (*Onchorhynchus mykiss*) sticks during refrigerated storage (4°C)

Haghparast S.*; Kashiri H.; Shabanpour B. and Pahlavani M.H.

Received: May 2008 Accepted: June 2009

Abstract

This study was carried out to investigate the rate of lipid oxidation in fresh rainbow trout (*Onchorhynchus mykiss*) and in the sticks treated by (2.5% w/v) sodium acetate (NaA), sodium citrate (NaC) and sodium lactate (NaL). The pH value, free fatty acid (FFA), thiobarbituric acid (TBA) and sensory evaluation (odor, flavor and color) were determined on 0, 3, 6, 9 and 12 days of storage. The results showed that TBA and FFA in control were significantly higher than those in the other groups (P<0.05). Sticks immersed in NaA indicated a significant difference in formation of free fatty acids in comparison with other sodium salt-treated samples on 9 and 12 days after storage. Sticks dipped in NaL had a maximum level of pH at the end of the storage, whereas samples treated by NaA achieved significantly the lowest value of pH, 9 days after storage. Organoleptic assessments of the samples expressed more acceptability of sticks immersed in sodium salt solutions than the control after 3 days of storage. These indicated that sodium salts, particularly sodium acetate, have antioxidant properties.

Keywords: Rainbow trout, Antioxidant, Sodium salt, Lipid oxidation, Refrigerated storage

Agricultural Sciences and Natural Resources of Gorgan University, P.O. Box: 386 Gorgan, Iran

^{*}Corresponding author's email: sarah_haghparast@yahoo.com

Introduction

Fish products like fish sticks elaborated from fatty fish have lately attracted strong interest on fish processing industry and consumers due to their nutritional value, mainly associated to its high content of essential fatty acids, n-3 fatty acids (Puwastien et al., 1999). These polyunsaturated fatty acids (PUFA) were reported to have beneficial effects on human health (Steffens, 1997), including lower blood triglycerides, reduce abnormal heart rhythms, reduce blood pressure and improve blood clotting regulation (Nettlon, 1995). They are also considered to be susceptible to peroxidetion damage (Richards & Hultin, 2002). Fatty fish such as rainbow trout (Onchorhynchus mykiss) are particularly sensitive to oxidative changes during storage which cause quality deterioration, lipid hydrolysis and formation of unpleasant odors (Gram et al., 1990; Venugopal & Shahidi, 1996). To minimize the undesirable effects of lipid proxidation, various strategies are being applied, including the use of refrigeration and addition of antioxidants to fish flesh (Pirini et al., 2000; Scaife et al., 2000). Antioxidative effects of sodium organic salts derived from citric, lactic and acetic acids have already been studied on color and lipid oxidation in n-3 oil fortified ground beef patties (Lee et al., 2005), quality changes of packaged shrimp and catfish fillets (Zhuang et al., 1996), and microbial damage of Salmon slices during tray-packaged storage at 1°C (Sallam, 2007). The main evaluation criteria of fresh quality of fish, sensory evaluation, were not assessed in these studies. The aim of the present study is the commercialization of refrigerated sticks of

rainbow trout and as well as the assessment of antioxidative effects of sodium salts on the stability of lipids in rainbow trout sticks during refrigerated storage.

Materials and methods

Rainbow trout was purchased from a local aquaculture farm located near Mahiran Village (Golestan province) with an average length and weight of 30±8cm and 400±50g, respectively. Specimens were washed several times in tap water and transported to the laboratory within 2 hours after catch, in boxes containing ice. Fish sticks were prepared using common household practices. First head, skin, tail, fins, and backbone were removed, then gutted and the resulting fillets were cut into 4 sticks (30±5g each). Sticks were randomly divided into 3 batches (3kg each). The first and second one were dipped for 10 min in pre-chilled (4°C) aqueous solutions (2.5% w/v) of NaA, NaC and NaL (E Merck, Darmstadt, Germany), respectively, and the last one was immersed in pre-chilled distilled water as a control sample. The ratio of dipping solution to fish was 2.5 to 1. After draining for 5 min on a plastic sieve, the treatments were packaged separately and stored under refrigeration (4±1°C). When required, the covering polyvinylidene film and Styrofoam trays were replaced. At each sampling occasion (0, 3, 6, 9 and 12 days) samples of each treatment were removed for subsequent chemical and sensory analysis.

The thiobarbituric acid index (TBA-i) was determined according to Natseba *et al.*

(2005) and expressed as mg malonaldehyde per kg fish flesh. 200mg of fish sample was homogenized in a 25ml balloon with 1-bothanol.5ml of the homogenate was mixed well with 5 ml of TBA agent in a cabbedtest tube. The mixture was heated in water bath (~95°C) for 2 hours and then cooled at room temperature. The color development was measured at 530nm as following where "As" is the absorbance of the sample against a blank sample of distilled water (Ab).

(mg malonaldehyde per kg fish flesh) TBA = 50(As-Ab)/200

Free fatty acid (FFA) content was measured as described by Egan *et al* (1997). 150g of comminuted fish sticks were mixed thoroughly with 250ml of chloroform and for 10 min and filtered with Wattman filtration paper.10ml of the filtered liquid was used for oil sample weight after drying at 105°C. Free fatty acid content was expressed as oleic acid percent by the titration of filtered liquid (2ml) containing 3 droplets of phenol phetalein with 0.1 N NaOH as follows:

(% oleic acid) FFA= (ml of 0.1 N NaOH used in the titration $\times 28.2 \times 100$) / (oil sample weight $\times 1000$).

The pH value was recorded using a pH meter (713 pH meter, Metrohm Herisau Switzerland) and based on the method of Lanier (1992). Fish sample (5g) was homogenized thoroughly with 45ml of distilled water and the homogenate used for pH determination.

Sensory assessments included the evaluation of three parameters (flavor, color and odor) were conducted by five trained

panelists. According to the method of Karmer *et al.* (1966), 5 categories of acceptability were ranked: excellent quality (5), good quality (4), acceptable (3), fair quality (2) and reject (1). Samples of each treatment were taken at regular intervals and the scores given to the judgments were analyzed.

All data were presented as means± standard error. The study was conducted as a 4×5 factorial experiment (factor A; NaA, NaC, NaL and control), five levels of storage time (factor B; t₁, t₂, t₃, t₄ and t₅) in three replications. ANOVA was used to search for significant differences among levels of each factor and also for interactions between two treated factors (storage x time). Data from pH, chemical and sensory measurements were subjected to ANOVA followed by least significant difference test (LSD) using Statistical Analysis System (SAS Institute, Inc., 1995). Also, comparisons between means for levels of each factor within other factor were performed.

Results

The initial pH in rainbow trout sticks ranged from 5.44 to 5.60 (Table 1). A significant increase (P<0.05) was found in all samples except those treated with NaA up to the sixth day of storage (Fig. 1). On day 9, sticks dipped in NaA solution (2.5%) had significantly lower pH value than the control and other treated samples (P<0.05).

The level of thiobarbituric reactive substances in the refrigerated sticks was found to increase significantly (P<0.05)

with storage period (Table 2). By the sixth day, TBARS were above 0.8 for the control, whereas those sticks immersed in NaA aqueous solution did not reached the same level by the 12 days of storage. As shown in Fig. 2, it is deduced that the use of

sodium acetate solution (2.5% w/v) slowly increased the formation of thiobarbituric acid reactive substances in refrigerated rainbow trout sticks, particularly at the end of storage time when compared with NaL-treated samples.

Table 1: Changes in pH value during refrigerated storage under sodium acetate (NaA), sodium citrate (NaC), sodium lactate (NaL) and control (distilled water)

	Storage Time (days)					
Factors	t ₁ =0	t ₂ =3	t ₃ =6	t ₄ =9	t ₅ =12	
NaA	5.60±0.006 ^a _D	5.62±0.040 ^{ab} _{CD}	5.71±0.040 ° _C	5.91±0.018 ^b _B	6.21±0.034 ^b _A	
NaC	$5.60\pm0.023~^{a}_{E}$	$5.70\pm0.043~^{a}_{D}$	$5.90\pm0.010^{\ a}_{\ C}$	$6.05\pm0.030^{\ a}_{\ B}$	6.25±0.021 ^b _A	
NaL	$5.45\pm0.020~^{a}_{\ D}$	$5.50\pm0.060^{\ b}_{\ D}$	$5.84\pm0.060^{ab}_{C}$	$6.03\pm0.030^{\ a}_{\ B}$	$6.40\pm0.012^{\ a}_{\ A}$	
Control	$5.44{\pm}0.040~^{\rm a}{_{\rm E}}$	5.60 ± 0.040 ab $_{ m D}$	5.75 ± 0.006 bc $_{\rm C}$	6.02±0.051 ^a _B	6.19±0.000 ^b _A	

Different script letters characterize significant differences in each column (a-c) and in each raw (A-E) for P<0.05.

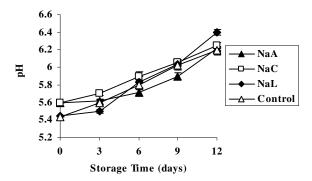


Figure 1: Comparison of pH value in rainbow trout sticks treated with NaA (sodium acetate), NaC (sodium citrate), NaL (sodium lactate) and under control. Bars denote standard error of the mean (n=3).

Table 2: Changes in TBARS value during refrigerated storage under sodium acetate (NaA),
sodium citrate (NaC), sodium lactate (NaL) and control (distilled water)

	Storage Time (days)					
Factors	t ₁ =0	t ₂ =3	t ₃ =6	t ₄ =9	t ₅ =12	
NaA	0.09±.005 ^a _C	0.18±0.030 ^a _C	$0.20\pm0.015^{\ c}_{\ C}$	$0.47\pm0.050^{\ c}_{\ B}$	0.81±0.110 ° _A	
NaC	0.09 \pm 0.015 $^{\mathrm{a}}$ _D	0.31 ± 0.050 a $_{C}$	0.25 ± 0.030 bc $_{\rm C}$	$0.54{\pm}0.040^{\text{ cb}}_{\text{ B}}$	$1.02\pm0.051^{\ bc}_{\ A}$	
NaL	0.11±0.011 a $_C$	$0.23\pm0.035~^{a}_{\ C}$	$0.33\pm0.060^{\ b}_{\ C}$	$0.75{\pm}0.050^{\ \mathrm{b}}_{\ \mathrm{B}}$	1.32±0.152 ^b _A	
Control	0.11±0.007 $^{\rm a}_{\rm \ D}$	$0.32\pm0.080~^{\mathrm{a}}_{\mathrm{C}}$	0.86±0.031 $^{\mathrm{a}}$ $_{\mathrm{C}}$	1.21±0.202 $^{\mathrm{a}}$ $_{\mathrm{B}}$	1.91±0.007 $^{\rm a}_{~\rm A}$	

Different script letters characterize significant differences in each column (a-c) and in each raw (A-D) for P<0.05.

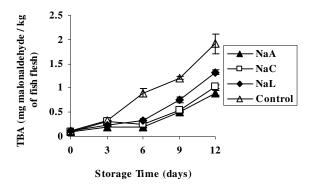


Figure 2: Comparative evolution of thiobarbituric acid (TBA-i) content in rainbow trout sticks treated with NaA (sodium acetate), NaC (sodium citrate), NaL (sodium lactate) and under control. Bars denote standard error of the mean (n=3).

Lipid hydrolysis in rainbow trout sticks occurred during refrigerated storage (Table 3). In both treated sticks with sodium salts and the control, free fatty acids increased over storage time. There were also significant differences (P<0.05) among sodium salt treated samples and the control after 3 days of storage. By the end of storage time (day 12), however, NaAtreated sticks achieved significant (P<0.05) lower FFA value of 3% in comparison with the control and Nal- or NaC-treated sticks which attained a higher levels of 8.48%,6.01% and 4.9%, respectively (Table 3 & Fig. 3).

Taste rating was conducted on treated rainbow trout sticks throughout 12 day-storage at 4°C (Table 4). As refrigerated storage time lengthened, flavor scores in treated rainbow trout sticks and in the control were significantly decreased (P<0.05). No significant differences (P>0.05) have been observed throughout the first 3 days of storage among different samples. Stored control sticks in refrigeration were considered organo-leptically unacceptable by 6th day of storage (Fig. 4).

Table 3: Changes in FFA content during refrigerated storage under sodium acetate (NaA), sodium citrate (NaC), sodium lactate (NaL) and control (distilled water)

	Storage Time (days)					
Factors	t ₁ =0	t ₂ =3	t ₃ =6	t ₄ =9	t ₅ =12	
NaA	0.37±0.027 ^a _C	0.34±0.070 ^b _C	0.43±0.082 ° _C	1.22±0.300 ° _B	3.00±0.190 ^c _A	
NaC	$0.36\pm0.030~^{a}_{\ C}$	0.37 ± 0.034 $^{\mathrm{b}}$ $_{\mathrm{C}}$	0.44 ± 0.031 $^{\rm c}_{ m C}$	$3.51\pm0.300^{\ b}_{\ B}$	$4.90\pm0.331^{\ b}_{\ A}$	
NaL	0.33 ± 0.056 a $_{C}$	0.41 \pm 0.016 $^{\mathrm{b}}$ $_{\mathrm{C}}$	0.86 ± 0.060 $^{\mathrm{b}}$ $_{\mathrm{C}}$	$3.90\pm0.370^{\ ab}_{\ B}$	6.01±0.200 ^b _A	
Control	$0.34\pm0.042~^{a}_{\ D}$	0.80±0.064 $^{\rm a}_{\ \rm D}$	2.12±0.145 $^{\mathrm{a}}$ $_{\mathrm{C}}$	$4.85{\pm}0.510~^{a}_{\rm \ B}$	8.48±0.100 ^a _A	

Different script letters characterize significant differences in each column (a-c) and in each raw (A-D) for P<0.05.

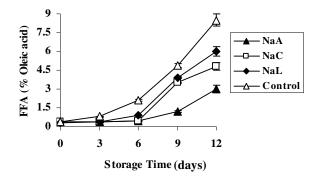


Figure 3: Comparative evolution of free fatty acid content in rainbow trout sticks treated with NaA (sodium acetate), NaC (sodium citrate), NaL (sodium lactate) and under control. Bars denote standard error of the mean (n=3).

Table 4: Changes in taste score during refrigerated storage under sodium acetate (NaA), sodium citrate (NaC), sodium lactate (NaL) and control (distilled water)

	Storage Time (days)					
Factors	t ₁ =0	t ₂ =3	t ₃ =6	t ₄ =9	t ₅ =12	
NaA	5.00±0.000 ^a _A	4.80±0.200 ^a _A	3.80±0.374 ^a _B	3.20±0.200 ^a _{BC}	2.60±0.245 ^a _C	
NaC	$5.00\pm0.000~^{a}_{A}$	4.60 ± 0.245 a $_{AB}$	$4.00{\pm}0.320~^a_{\rm \ B}$	$2.80{\pm}0.245~^{\rm a}_{\rm \ C}$	2.40±0.245 $^{\mathrm{a}}$ $_{\mathrm{C}}$	
NaL	$5.00\pm0.000~^{a}_{A}$	$4.40{\pm}0.245$ $^{a}_{\ A}$	$3.00\pm0.320~^{a}_{~B}$	$2.20\pm0.200^{\ b}_{\ C}$	1.40 ± 0.245 $^{\mathrm{b}}$ $_{\mathrm{D}}$	
Control	$5.00\pm0.000^{\ a}_{\ A}$	$2.60\pm0.245^{\ b}_{\ B}$	$2.00\pm0.320^{\ b}_{\ C}$	$1.20\pm0.200^{\ c}_{\ D}$	$1.00\pm0.000^{\ b}_{\ D}$	

Different script letters characterize significant differences in each column (a-c) and in each raw (A-D) for P<0.05.

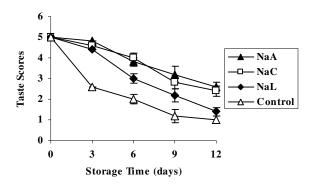


Figure 4: Comparison of taste scores given to rainbow trout sticks treated with NaA (sodium cetate), NaC (sodium citrate), NaL (sodium lactate) and under control. Bars denote standard error of the mean (n=3).

Results indicate a decreasing trend in scores given to odor in all samples during storage (Fig. 5). Sticks immersed in NaC, NaL and distilled water (as the control) solutions, were rated below 3 after 6 and 9 days at 4°C except for those treated with NaA which were rated above 3 even by the 12th day (Table 5).

Changes in the color of refrigerated rainbow trout sticks were also examined (Table 6). Storage time had a significant effect on the color scores for each of the control and treated samples. The surface of treated sticks, especially those treated with NaA, was not severely discolored and remained acceptable after 12 days storage. Analysis of color scores showed panel preference for sticks dipped in 2.5% NaA and NaC aqueous solutions over refrigerated control and samples treated with NaL.

Table 5: Changes in odor score during refrigerated storage under sodium acetate (NaA), sodium citrate (NaC), sodium lactate (NaL) and control (distilled water)

	Storage Time (days)					
Factors	t ₁ =0	t ₂ =3	t ₃ =6	t ₄ =9	t ₅ =12	
NaA	5.00±0.000 ^a _A	4.60±0.245 ^a _A	4.40±0.245 ^a _A	3.60±0.245 ^a _B	3.40±0.245 ^a _B	
NaC	5.00±0.000 ^a _A	$4.40{\pm}0.245~^{\rm ab}_{A}$	$4.20\pm0.200~^{a}_{~A}$	$3.00\pm0.320^{\ ab}_{\ B}$	$2.20\pm0.374^{\ b}_{\ C}$	
NaL	5.00±0.000 ^a _A	$4.20\pm0.374^{ab}_{\ A}$	$2.80\pm0.374^{\ b}_{\ B}$	$2.40\pm0.000^{\ b}_{\ BC}$	1.60 ± 0.245 bc $_{\rm C}$	
Control	5.00±0.000 ^a _A	$3.60\pm0.245^{\ b}_{\ B}$	$2.40\pm0.400^{\ b}_{\ C}$	$1.600{\pm}0.245^{\rm \ c}_{\rm \ D}$	$1.00\pm0.000^{\ c}_{\ D}$	

Different script letters characterize significant differences in each column (a-c) and in each raw (A-D) for P<0.05.

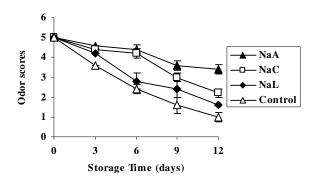


Figure 5: Comparison of odor scores given to rainbow trout sticks treated with NaA (sodium acetate), NaC (sodium citrate), NaL (sodium lactate) and under control. Bars denote standard error of the mean (n=3).

Table 6: Changes in color score during refrigerated storage under sodium acetate (NaA), sodium citrate (NaC), sodium lactate (NaL) and control (distilled water)

	Storage Time (days)					
Factors	t ₁ =0	t ₂ =3	t ₃ =6	t ₄ =9	t ₅ =12	
NaA	5.00±0.000 ^a _A	4.80±0.200 ^a _A	3.80±0.200 ^a _B	3.40±0.245 ^a _{BC}	3.00 ±0.000 ^a _C	
NaC	$5.00\pm0.000~^{a}_{A}$	$4.60\pm0.245^{-a}{}_{A}$	$3.60\pm0.245^{a}_{B}$	2.40 ± 0.245 $^{\mathrm{b}}$ $_{\mathrm{C}}$	$2.60\pm0.245~^{a}_{\ C}$	
NaL	5.00±0.000 ^a _A	$4.40\pm0.245^{a}_{B}$	$3.20\pm0.200^{-ab}_{C}$	2.00 ± 0.245 $^{\rm c}$ $_{\rm D}$	$1.80\pm0.200^{\ b}_{\ D}$	
Control	5.00±0.000 ^a _A	$3.40\pm0.245^{\ b}_{\ B}$	$2.60\pm0.245^{\ b}_{\ C}$	1.60 ± 0.245 $^{\rm c}$ $_{\rm D}$	$1.00\pm0.000^{\ c}$ E	

Different script letters characterize significant differences in each column (a-c) and in each raw (A-E) for P<0.05.

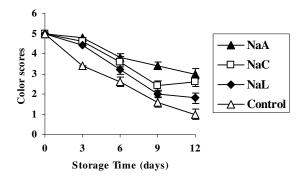


Figure 6: Comparison of color scores given to rainbow trout sticks treated with NaA (sodium acetate), NaC (sodium citrate), NaL (sodium lactate) and under control. Bars denote standard error of the mean (n=3).

Discussion

The effects of aqueous solutions of sodium salts (2.5% w/v) on the shelf-life of refrigerated rainbow trout sticks were studied on the basis of the generation of FFA, TBA, changes in pH and sensory values in this study. During storage the quality of fish degrades due to a complex process in which a number of physical, chemical and microforms deterioration biological of implicated (González et al., 2005). Oxidative rancidity is an important organoleptic characteristic for rejection or approval of fish after prolonged storage (Amanatidou et al., 2000).

As showed, by the end of storage, pH values of either sodium salt-treated sticks or the control significantly (P<0.05) increased as shown in Fig 1. Increases in pH may be attributed to the production of volatile base compounds by bacterial activity (Cann *et al.*, 1983). Similar observations were made by Al-Sheddy (1999) who reported that pH of camel meat was not significantly (P<0.05) affected under sodium acetate treatment (10% w/w) in comparison to sodium lactate (5% v/v of 60% solution) and trisodium citrate (1.5% w/w)-treated samples after 12 days storage at 4°C.

TBARs reactive materials that is generally produced in substantial amounts only from fatty acids containing three or more double bonds (Nawar, 1985), found in relatively high concentrations in Mediterranean fatty fish such as Sardines, Mackerel and Rainbow trout (Simeonidou *et al.*, 1998; Auborge, 2001; Chytiri *et al.*, 2004). In the current study, all sticks treated with sodium salt solutions of organic acids had significantly lower TBA

values after 3-days of storage than the control (P<0.05). In contrast, Sallam (2007) demonstrated that dipping sliced salmon in NaL did not produce any significant reduction in the TBA content (P>0.05) in the comparison with the control during 15 days storage at 1°C. However, results from previous study revealed a significant lower TBA value for catfish fillets treated with 2% sodium lactate after 2 days and through 8 days at 1°C as compared with the control (Williams et al., 1995). The approximately more increases in TBA content for both of the control and treated sticks in this study indicated that in rainbow trout, which is a fatty fish, lipids are highly susceptible to the autoxidation during refrigerated storage as previous reports proved that (Simeonidou et al., 1998). Although the fish may be acceptable for consumption with lower than 8 mg malonaldehyde/kg (Schormüller, 1969), it has been suggested that values more than maximum levels of TBARs (3-4mg malonaldehyde/kg) indicates lipid oxidation and loss of quality in fish flesh (Taskaya et al., 2003). In the present study, TBA content for both the control and treated sticks of rainbow trout were much lower than proposed limit through the 12-days storage.

Examination of free fatty acids deemed to be important since it has been proved that accumulation of FFA is related to some extent to texture deterioration by interacting with proteins (Mackie, 1993; Sikorski & Kolakowska, 1994) and to lipid oxidation enhancement (Miyashita & Takagi, 1986;

Yoshida *et al.*, 1992). In this experiment, the free fatty acids content evolution was quite similar under both sodium salts treatments and the control up to the sixth day of storage, showing a slight increase with time. Afterwards, a lower FFA formation was observed for fish samples treated with NaA than in their counterparts immersed in NaL, NaC and distilled water as control.

Significant effect of storage time on acceptability scores verified in control and sodium salt-treated (2.5% w/v) rainbow trout sticks after 12-days storage at 4°C. All sensory scores (odor, flavor and color) for rainbow trout samples treated with sodium organic salt solutions and under control treatment showed a decreasing acceptability pattern. In the current study, sticks treated with NaC and NaA solutions had flavor scores that were significantly (P<0.05) distinguishable from the control and NaLtreated samples at the end of storage period. Results of color evaluation of the present study confirmed those of Al-Sheddy's reports (1999) about a significant (P<0.05) reduction in surface discoloration of camel meat dipped in sodium acetate (10% w/w) when compared to controls and other samples. Mendonca et al. (1989) demonstrated that the presence of sodium acetate with acetic acid improved color quality of pork chops than the use of acetic acid alone. Odor change was detected in sticks treated with NaA and the panelists characterized the new odor as mild acetic acid-like even at the first day of storage (P<0.05). By the end of storage, only samples treated with NaA solution remained acceptable and continued to exhibit desirable odor as mild acetic acidlike odor in the present trail. A previous study demonstrated that camel meat treated with sodium acetate (10% w/w) alone or combined with Bifidobacteria (5%) had significantly the least strong off-odors than other samples after 9-days of storage at 4°C (Williams et al., 1995). Based on scores given to sensory attributes, a shelf life of 9 (NaA), 6 (NaC) and 3 (NaL and Control) days were achieved in this experiment. Manju (2007) reported that vacuum-packaging in conjuncttion with 2% sodium acetate can be safely used to extend the shelf life of Pearlspot samples up to 15 days at 2°C. Results from previous study proved that a combination of sodium acetate (dip treatment) and Bifidoshelf life bacteria extended the of refrigerated catfish fillets.

This study demonstrates that NaA is the best chemical treatment to prolong the shelf-life of stored rainbow trout sticks in refrigeration at 4°C at least by 9 days compared to other treatments. To evaluate the effect of sodium acetate on the sensory quality of fish, further studies such as microbiological analysis is proposed, which has not been investigated in the present experiment.

Acknowledgments

We acknowledge Mr. Mirfendereski for careful reading of this manuscript. We are also grateful to all staff working at our university for their help and providing facilities during the trail.

References

- AI-Sheddy, I., AI-Dagal, M. and Bazaraa, W.A., 1999. Microbial and sensory quality of fresh camel meat treated with organic acid salts and/or Bifidobacteria. Journal of Food Science, 64(2):336-339.
- Amanatidou, A., Schlüter, O., Lemkau, K., Gorris, L.G.M., Smid, E.J. and Knorr, D., 2000. Effect of combined application of high pressure treatment and modified atmospheres on the shelf life of fresh Atlantic salmon. Innovative Food Science & Emerging Technologies, 1: 87–98.
- **Aubourg, S., 2001.** Damage detection in horse mackerel (*Trachurus trachurus*) during chilled storage. Journal of the American Oil Chemists' Society (JAOCS). **78:**857–862.
- Cann, D.L., Smith, G.L. and Houston, N.G., 1983. Further studies on marine fish stored under modified atmosphere packaging. Aberdeen: Ministry of Agriculture Fisheries and Food, Torry Research Station, Scotland. 322P.
- Chytiri, S., Chouliara, I., Savvaidis, I.N. and Kontominas, M.G., 2004. Microbiological, chemical, and sensory assessment of iced whole and filleted aquacultured rainbow trout. Food Microbiology, 21:157–165.
- Egan, H., Krik, R.S. and Sawyer, R., 1997.

 Pearson's chemical Analysis of food. (9th edn.). Churchil Livingstone. Edinburgh, UK. pp.609-634.
- González-Fandos, E., Villarino-Rodríguez, A.,
 García-Linares, M.C., García-Arias, M.T.
 and García-Fernández, M.C., 2005.
 Microbiological safety and sensory characteristics of salmon slices (salmo salar)

- processed by the sous vide method. Food Control, **16:**77–85.
- Gram, L., Wedell-Neergaad, C. and Huss, H.H., 1990. The bacteriology of fresh and spoiling Nile perch. International Journal of Food Microbiology, 10:303-316.
- Karmer, A. and Twigg, B.A., 1966.

 Fundamental of quality control for the food industry. 2nd edition, AVI. Pennsylvania, USA. 505P.
- Lanier, T.C., 1992. Measurement of surimi composition and functional properties. 1123-1163. *In*: Surimi Technology. (Eds., T.C. Lanier and C.M. Lee) Macrel Decker, Inc., New York. USA, 3441P.
- Lee, S., Decker, E.A. and Faustman, C., 2005. The effects of antioxidant combinations on color and lipid oxidation in n-3 oil fortified ground beef patties. Meat Science, 70(4):pp.683-689.
- Manju, S., Jose, L., Srinivasa Gopal, T.K., Ravishankar, C.N. and Lalitha, K.V., 2007. Effects of sodium acetate dip treatment and vacuum-packaging on chemical, microbiological, textural and sensory changes of Pearlspot (*Etroplus suratensis*) during chill storage. Food Chemistry, 102:27-35.
- **Mackie, I., 1993.** The effects of freezing on flesh proteins. Food Reviews International, **9:**575–610.
- Mendonca, A.F., Molins, R.A., Kraft, A.A. and Walker, H.W., 1989. Microbiological, chemical, and physical changes in fresh vacuum-packaged pork treated with organic acids and salts .Journal of Food Science, 54:18-21.

- Miyashita, K. and Takagi, T., 1986. Study on the oxidative rate and proxidant activity of free fatty acids. Journal of the American Oil Chemists Society, 63:1380–1384.
- Natseba, A., Lwalinda, I., Kakura, E.,
 Muyanja, C.K. and Muyonga, J.H., 2005.
 Effect of pre-freezing icing duration on quality changes in frozen Nile perch (*Lutes niloticus*). Food Research International, 38:469-474.
- **Nettlon, J.A., 1995.** Omega-3 fatty acids and health. Chapman &Hall. New York, USA. pp.18-27.
- Pirini, M., Gatta, P.P., Testi, S., Trigari, G. and Monetti, P.G., 2000. Effect of refrigerated storage on muscle lipid quality of sea bass (*Diaentrachus labrax*) fed on diets containing different levels of vitamin E. Food Chemistry, **68**:289-293.
- Puwasyien, P., Judprasong, K., Kettwan, E., Vasanachitt, K., Nakngamanong, Y. and Bhattacharjee, L., 1999. Proximate composition of raw and cooked Thai freshwater and marine fish. Journal of Food Composition and Analysis, 12:9-16.
- Richards, M. and Hultin, H., 2002.

 Contribution of blood and blood components to lipid oxidation in fish muscle. Journal of the Agriculture and Food Chemistry, 50:555-564.
- **Sallam, Kh.I., 2007.** Antimicrobial and antioxidant effects of sodium acetate, sodium lactate and sodium citrate in refrigerated sliced salmon. Food Control, 18(5):566-575.
- **SAS Institute, 1996.** SAS/STAT guide for personal computers, version 6.1. Cary, NC: SAS Institute Inc.

- Scaife, J.R., Onibi, G.E., Murray, I., Fletcher, T.C. and Houlihan, D.F., 2000. Influence of α-tocopherol acetate on the short-and long-term storage properties of fillets from Atlantic salmon *Salmo salmo salar*-fed a high lipid diet. Aquaculture. Nutrition, 6:65-71.
- Schormüller, J., 1969. Handbuch der lebensmittelchemie (Band IV). Berlin, Heidelberg: Springer-Verlag. 177P.
- Sikorski, Z. and Kolakowska, A., 1994.
 Changes in protein in frozen stored fish. *In*:
 (Z. Sikorski, B. Sun Pan, & F. Shahidi eds.), Seafood proteins. Chapman & Hall.
 New York, USA. pp.99–112.
- Simeonidou, S., Govaris, A. and Vareltzis, K., 1998. Quality assessment of seven Mediterranean fish species during storage on ice. Food Research International, 30:479–484.
- **Steffens, W., 1997.** Effects of variation in essential fatty acids in fish feeds on nutritive value of freshwater fish for humans. Aquaculture, **151**:97-119.
- **Taskaya, L., Duygu, S.C. and Kilinc, B., 2003.** Quality changes of fish burger from rainbow trout during refrigerated storage. European Journal of Fisheries & Aquatic Science, 20(1-2):147-154.
- Venugopal, V. and Shahidi, F., 1996. Structure and composition of fish muscle. Food Reviews International, 2(2):175.
- Williams, SK., Rodrick, GE. and West, RL., 1995. Sodium lactate affects shelf life and consumer acceptance of fresh catfish (*Ictalurus nebulosus*, *marmoratus*) under stimulated retail conditions. Journal of Food Science, 60:636–639.

Yoshida, H., Kondo, I. and Kajimoto, G., 1992. Participation of free fatty acids in the oxidation of purified soybean oil during microwave heating. Journal of the American Oil Chemists Society, 69:1136–1140.

Zhuang, R.Y., Huang, Y.W. and Beuchat, L.R., 1996. Quality changes during refrigerated storage of packaged shrimp and catfish fillets treated with sodium acetate, sodium lactate or propyl gallate. Journal of Food Science, 61:241–244.

خواص آنتی اکسیدانتی استات سدیم، سیترات سدیم ولاکتات سدیم بر اکسیداسیون چربی استیکهای قزل آلای (Onchorhynchus mykiss)

طی نگهداری در یخچال (۴ درجه سانتیگراد)

سارا حقپرست*؛ حدیثه کشیری؛ بهاره شعبانپور و محمد هادی

پهلوانی

تاریخ دریافت: اردیبهشت ۱۳۸۷

ینیرش: خرداد ۱۳۸۸

چکیده

Onchurhynchus) و پس از غوطهوری در محلولهای استات سدیم، سیترات سدیم و لاکتات سدیم (1 N درصد) در مقایسه با (1 mykiss) و پس از غوطهوری در محلولهای استات سدیم، سیترات سدیم و لاکتات سدیم (1 N درصد) در مقایسه با تیمار شاهد (غوطهوری در آب مقطر) صورت گرفت. آزمای شات تعی بین TBA 1 FFA 1 PH و 1 N 1 PH نمونههای در روزهای صفر، 1 N 1 Ph 1 Ph 1 PH 1 PH 1 PH نمونههای فوطهور شده در شاهد بطور معنی داری بالاتر از سای 1 Ph 1 PH

کــلـمــات کــلـیــدی: قزل آلای رنگین کمان، نمکهای سدیم، آنتی اکسیدانت، اکسیداسیون چربی، نگهداری در یخچال

دانشگاه علوم کشاورزي و منابع طبیعي گرگان، گرگان صندوق پستي: ۳۸۹

^{&#}x27; پست الكترونيكي نويسندة مسئول: sarah_haghparast@yahoo.com