The growth, survival rate and reproductive characteristics of Artemia urmiana fed by Dunaliella tertiolecta, Tetraselmis suecica, Nannochloropsis oculata, Chaetoceros sp., Chlorella sp. and Spirolina sp. as feeding microalgae

Mohebbi F.^{1*}; Hafezieh M.²; Seidgar M.¹; Hosseinzadeh Sahhafi H.²; Mohsenpour Azari A.¹; Ahmadi R.¹

Received: February 2014 Accepted: November 2014

Abstract

This study was performed to compare the efficiency of six microalgae namely Dunaliella tertiolecta, Tetraselmis suecica, Nannochloropsis oculata, Chaetoceros sp., Chlorella sp. and Spirolina sp. on the growth, survival rate and reproduction efficacy in Artemia urmiana in laboratory conditions. Artemia cysts were harvested from Urmia Lake and hatched according to the standard method. Live microalgae were cultured using the f/2 culture medium. Artemia survival was determined in treatments on days 8, 11, 14, 17 and 20. A highly significant difference (p<0.01) were found among three microalgae in terms of length growth, survival rates and reproduction characteristics in A. urmiana. In spite of higher length growth of A.urmiana fed on N. oculata than A. urmiana fed by T. suecica but survival and reproduction in the latter was better than the first treatment. In general, D. tertiolecta was more efficient than other microalgae examined in the present study on A. urmiana concerning not only to growth and survival but also to reproduction mode. So, it is preferred to feed A. urmiana.

Keywords: Artemia urmiana, Microalgae, Length growth, Survival rate

¹⁻Iranian Fisheries Science Research Institute, Natinal Artemia Research Center, Agricultural Research Education and Extension Organization (AREEO), Urmia, Iran.

²⁻Iranian Fisheries Science Research Institute, Agricultural Research Education and Extension Organization (AREEO), Tehran, Iran.

^{*} Corresponding author's Email: mohebbi44@gmail.com

Introduction

Potentially, Artemia is an excellent feed for fish and crustaceans (Sorgeloos, 1980). The brine shrimp Artemia is probably the most popular live diet in aquaculture. Artemia is a non-selective filter feeder. It is able to use all nutrients that are smaller than its mouth. Various factors affect Artemia's filtration rate, ingestion, digestion and feeding behavior. These factors include the quality and quantity of feed such as floatability, minimum solubility in water, digestibility and size and so on (Sorgeloos et al., 1998).

Due to its particular biological characteristics, Artemia can be fed on different diets, from live microalgae to microcapsules and waste products from the food industry (Lavens and Sorgeloos, 1991). Microalgae strains are recognized as excellent sources of proteins, carbohydrates, lipids, vitamins. and as food and feed additives. Nannochloropsis sp. is well known as a source of EPA, an important polyunsaturated fatty acid (Radhakrishnan et al., 2009). Chlorella sp. is also recognized as a source of EPA.

The bioencapsulation technique provides interesting opportunities for using *Artemia* biomass not only as food attractant, but also as carrier for administration of various products to the predator, such as essential nutrients, pigments, hormones, and prophylactic or therapeutic agents (Léger *et al.*, 1986, Majack *et al.*, 2000, Malpica Sanchez *et al.*, 2004).

Suitable algal species for filter-feeding organisms such as *Artemia* are selected according to mass culture potential, cell size, digestibility and nutritional value (Hafezieh, 2004).

Diatoms are considered good sources of highly unsaturated fatty acids, especially of 20:5ω-3 (Lora-Vilchis and Voltolina, 2003). contrast, chlorophytes are rich in C16 and C18 fatty acids (Dunstan et al., 1992; Brown et al., 1997;), and in particular Chlorella has also a high content of carotenoids and ascorbic acid (Czygan, 1968; Merchie et al., 1995), which might be of importance for especially growth and long-term enhancement of the food quality of Artemia.

In natural habitats, microalgae form the main food source for *Artemia*. In Urmia Lake, for example, the microalga *Dunaliella* is the dominant species of the lake microalgal flora and composes more than 90% of algal density (Mohebbi *et al.*, 2009; Mohebbi, 2010). Obviously, *Artemia* often feeds on *Dunaliella* in most of its natural habitats.

Considering the substantial growth of aquaculture activities, it is useful to study more about microalgae suitability for *Artemia* feeding. Besides, studies on native *Artemia* populations represent an alternative for the exploitation of natural resources, also favoring the development of the local aquaculture industry. While there are so many studies on the effect of different algae on various *Artemia* strains, there are

few studies related to *A. urmiana* fed on various microalgae. The purpose of this study was to investigate and compare the effects of various algae on the growth, survival rate and reproduction of *A. urmiana*, and to determine the most appropriate algal species for *A. urmiana* in laboratory conditions.

Materials and methods

Microalgae culture

Stock culture of *T. suecica* was provided from the Persian Gulf Ecology Research Institute in Bandar Abbas (Iran). *N. oculata* was sent from Aquaculture Research Institute of South (Ahvaz, Iran).

Live microalgae were cultured using the f/2 culture medium (Guillard, 1975). A volume of 20 mL sea water (20-24 ppt) was poured into twenty five 75-mL test tubes and 40 µL of f/2 medium was added to each tube. When the tubes were cooled, 1-2 drops of vitamin solution was added to each tube. A little of alga was removed from stock culture by forceps and transferred into test tubes. The tubes were placed in suitable condition and stirred several times daily. After a few days, the tubes went green. Then the alga of each tube was transferred into a 250-mL or 500-mL flasks which contained the f/2 medium and vitamin. Similarly, this cycle was repeated until the algae were finally transferred into 30-L plastic bags and 100-L tanks. When the algal density reached a maximum level, aeration was interrupted. Then, the algal solution was concentrated more by cooling in the

refrigerator. The concentrated alga was diluted up to a determined level (18× 10⁶ cells/mL) before use for *Artemia* feeding. The density of the alga was determined using a Neubar slide and a Nikon ECLIPSE 50i microscope.

Artemia culture

Artemia cysts were harvested from Urmia Lake and hatched according to Sorgeloos et al. (1986). Artemia were starved during the first 24 hr in order to allow yolk resorption (Teresita and Leticia, 2005). Newly hatched larvae were enumerated and 500 larvae were placed in one conical vessel (4 repeats from each treatment) that contained 1000 cc of water with 80 ppt salinity. The vessels were placed in the incubator with $25 \pm 1^{\circ}$ C temperature (Boone and Bass-Becking, 1931).

Brine shrimp nauplii were experimentally kept under the following culture conditions: 25±2.5°C temperature, 30 ± 1.3 ppt salinity. 8.0 ± 0.4 pH and >5 mg L⁻¹ dissolved oxygen. Feeding the larvae was started according to Coutteau et al. (1992) 24 hr after hatching of the cysts. The used food composed of the algae D. tertiolecta, T. suecica and N. oculata. At the beginning, Artemia density was one larva per 2 mL of water which was reduced to one Artemia per 3 mL and one Artemia per 4 mL on days 8 and 14, respectively (Boone and Bass-Becking, 1931).

On days 8, 11, 14, 17 and 20, ten animals were taken out from each container (30 per treatment) and

measured (from the naupliar eye to the telson; (Amat, 1980) using Motic Images plus 2.0 software.

Artemia survival percentages were determined in three treatments on days 8, 11, 14, 17 and 20 (Cruz *et al.*, 1993).

When the Artemia were grown as adults, 30 females and 30 males were randomly selected and transferred into cylindrical bottom-conical small vessels named falkons (one female and one male Artemia in each falkon). In order to control the falkons' temperature, they were placed in special boxes (Racks) which in turn were put in aquariums with 25°C temperature (Boone and Bass-Becking, 1931). For each Artemia one drop of the enumerated algae (18× 10⁶ cells/mL) was daily added into the falkons. The water content of the falkons was changed daily. At the same time, the probable produced cysts or larvae were counted using a WILD M₃C model stereomicroscope (Mohammadyari, 2002). The type and number of offspring, the reproduction rate in the study period, the day of first reproduction, the interval between the two consecutive reproductions were calculated for each pair of Artemia.

One way analysis of variance (ANOVA) and Duncan test were used to compare the average of properties. All diagrams were produced in Excell 2007.

Results

A significant difference (*p*<0.01) was observed between length growths of *A. urmiana* fed on three different microalgae so that, the *A. urmiana* fed

on *D. tertiolecta* and *T. suecica* showed the highest and the lowest length growth, respectively (Fig. 1). In the study period (20 days) the mean of length growths were 5.171 mm, 4.555 mm and 3.131 mm 4943.44 mm and 4820.024 mm in *A.urmiana* fed on microalgae *Dunaliella tertiolecta*, *N. oculata* and *T. suecica*, *Chaetoceros* sp., *Chlorella* sp. and *Spirolina* sp. respectively (Table 1).

Survival indicated significant difference (p<0.05) between Artemia fed on N.oculata than those fed on D. tertiolecta and T. suecica so that Artemia fed on N. oculata showed lower survival percentages than the two latter treatments (Fig. 2). Besides, the Artemia fed on T. suecica contained lower survival rates than those fed on D. tertiolecta, though this difference was not statistically significant (Fig.2). The survival rate in various days of the experiment showed significant difference between days 8 and 11 and between days 11 and days 14, 17 and 20 (p<0.05).

There was no significant difference in survival percentages among repeats in three different microalgae. However, survival percentages among various days of the experiment suggested that it was higher in Artemia fed on D. tertiolecta than Artemia fed on T. suecica which in turn was higher than those fed on Nannochloropsis oculata (p<0.01, Table 2). On the other hand, Spirolina sp. induced the highest mortality in A. urmiana (Fig 2). Also, A. urmiana fed by Chaetoceos sp. and Chlorella sp. indicated relatively similar survival patterns (Fig. 2). This pattern of survival was similarly observed on days 8, 11, 14, 17 and 20 of the experiment.

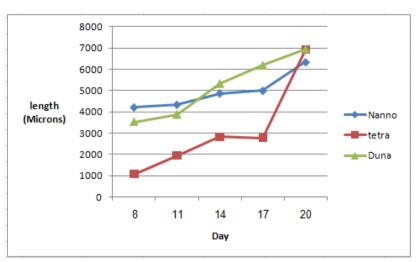


Figure 1: Length growth of Artemia urmiana treated by three microalgae feeds.

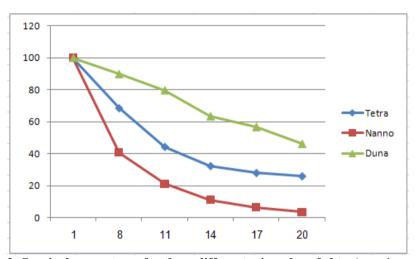


Figure 2: Survival percentages for three different microalgae fed to Artemia urmiana.

Table 1: Means of length growth between Artemia urmiana fed on different microalgae (p<0.01).

		8 1 /
Microalga	N	Mean of length growth (mm) ±Std.Deviation
Tetraselmis suecica	152	3131.14±1447.033
Nannochloropsis oculata	66	4555.47 ± 719.085
Chaetoceros sp.	81	4943.44±542.92
Chlorella sp.	75	4820.024±552.82

Cysts and nauplius production were only observed in *A. urmiana* fed on *D. tertiolecta* and *T. suecica. Chaetoceos* sp. and *Chlorella* sp. . In other words, *A.urmiana* fed on *N. oculata* and *Spirolina* sp.did not mature to produce cysts or naplius. The comparison of cysts and nauplius production between

A. urmiana fed on D. tertiolecta and T. suecica indicated a significant difference (p<0.01). A. urmiana fed on D. tertiolecta produced much more cysts and naplius than the A. urmiana fed on T. suecica (Table 3). The mean cysts production in A. urmiana treated with D. tertiolecta and T. suecica

Chaetocerus sp. and Chlorella sp. were 12.87, 2.47, 1.2 and 1.6 cysts over the experiment period, respectively. Also, A. urmiana fed on D. tertiolecta and T. suecica Chaetocerus sp. and Chlorella sp. produced 8.36, 2.60, 1.19 and 1.50nauplius in the experiment period respectively. Significant differences were observed between A. urmiana fed on D. tertiolecta and T. suecica in terms of the number of reproductions in the

study period and the day of first reproduction (p< 0.01), but these two treatments did not indicate any significant differences with regard to the interval between two consecutive reproductions.

There was a significant difference (p<0.01) only between repeats 1 and 3 in *A. urmiana* fed on *D. tertiolecta*. Other repeats did not indicate any significant differences in terms of cysts and nauplius production.

Table 2: Means of survival rates for Artemia urmiana fed on different microalgae.

Days	mean±Std.Deviation
8	482.75 ± 0.00
	264.25 ± 0.00
	132.25 ± 0.00
	107.50 ± 0.00
20	91.75 ± 0.00
8	342.25± 30.66
11	221.50 ± 4.79
14	124.25 ± 76.63
17	106.50 ± 68.07
20	98.50 ± 63.84
8	204.00 ± 26.14
11	106.25 ± 42.94
14	55.75 ± 16.52
17	32.00 ± 6.27
20	17.75 ± 3.30
8	75.1±7.3
	54.3 ± 5.2
	42.9±7.1
	32.5±6.9
20	23.9±4.6
8	73.1±4.1
	52.1±6.4
	42.5±7.5
	33±6.3
20	23.5±5.7
8	7.1±2.1
	1.45 ± 1.1
	0.4 ± 0.6
	0.4±0.0 0.0±0.0
20	0.0±0.0 0.0±0.0
	8 11 14 17 20 8 11 14 17 20 8 11 14 17 20 8 11 14 17 20 8 11 14 17 20 8 11 14 17 20 8 11 14 17 20 8 11 14 17 20

Table 3: Cysts and nauplius production in Artemia urmiana fed by different microalgae.

Microalga	repeat	Cysts	Nauplius
	-	(mean±Std.Deviation)	(mean±Std.Deviation)
Dunaliella tertiolecta	1	12.975± 12.057	9.077± 9.076
	2	13.110 ± 5.030	11.306 ± 7.235
	3	10.183 ± 6.357	14.816 ± 7.504
Tetraselmis suecica	1	3.304 ± 2.944	2.819± 4.389
	2	2.829 ± 2.251	2.799 ± 2.764
	3	1.355 ± 1.247	2.235 ± 2.586
Chaetocerus sp.	1	1.255±1.235	1.191±1.242
	2	0.812 ± 0.682	1.352±1.110
	3	0.954 ± 0.825	1.542±1.365
Chlorella sp.	1	1.626±1.411	1.547±1.324
	2	1.360±1.032	1.881±1.547
	3	1.502±1.361	1.425±1.256

Discussion

It is well accepted that Artemia is the most widespread live food item used in the production of shrimp, prawn and fish larval stages. The organism can be used in different forms in hatcheries and nurseries, e.g. decapsulated cysts, nauplii, metanauplii, juvenile and adult stages, and frozen and freeze-dried Artemia biomass. Artemia biomass is nowadays more frequently used for specific stages of aquatic species as it enhances production characteristics and resistance and/or overall stress decreases cannibalism in dolphin fish and lobster larviculture (Lavens and Sorgeloos, 1991).

The quality of microalgae diets for Artemia has been the object of several studies (e.g. Sick, 1976; Johnson, 1980; Fábregas et al., 1996, 1998) with different results, depending on the microalgae, species of culture conditions, and possibly the species of Artemia used for the feeding experiments.

Maldonado-Montiel and Rodríguez-Canché (2005) reared a Mexican local Artemia with rice bran (days 1- 6) and microalga T. suecica (days 7-15). They reported 79% survival rate at the end of trial which was higher than the value observed on day 14 in our study. They also measured a mean length of 5.34mm for Artemia at the end of their experiment (day 15). This value was higher than that in our study for which we obtained amean length of 3.01mm for A. urmiana fed on T. suecica on day 14. These differences may be attributed either to Artemia species or to Mexican tropical climate, sharply different than ours.

The results of the present study confirmed those obtained by Voojodzadeh *et al.* (2007) who found that *A. urmiana* fed with *N. oculata* did not produce any cysts or larvae even though they were reared until day 30. However, our study indicated that *A. urmiana* fed on *T. suecica* had the lowest length growth among treatments

which was not consistent with the work of Voojodzadeh *et al.* (2007).

A.urmiana fed with Spirolina sp. had the lowest (18.5%) survival rate and indicated statistically significant difference with other algae examined in this study (P< 0.00). this was due to large size of this alga which was unsuitable for Artemia. In fact, Spirolina sp. should be powdered before was fed to Artemia (Garcia-Ulloa and Garcia-Olea, 2004).

On the other hand, Fabregas et al. (1996) evaluated T. suecica nutritional value on Artemia's total growth, survival reproduction and characteristics in different culture concentrations. They obtained the best results when Artemia were fed on T. grown nutrient suecica at a concentration of 8 mg atom N 1-1. This concentration was relatively higher than that of T. suecica concentration we used in our study. Therefore, we may attribute the lower length growth of A.urmiana fed by T. suecica to lower concentration of this microalga.

Study conducted by Hafezieh (2004) indicated that the application of *Chaetoceros* sp. as live food for *A.urmiana* had significantly different effect on body length than *Chlorella* sp. which confirms our study. However, in our study *Chaetoceros* sp. had higher effect on Artemia body length than *Chlorella* sp. that was reverse to Hafezieh (2004).

In spite of the fact that T. suecical induced lower growth (mean length = $3131.14 \mu m$) in A. urmiana than N.

oculata (mean length = $4555.47 \mu m$) in our study, but reproduction outcome was better than A. urmiana fed on N. oculata (Table 3). This suggested that suecica had higher effects in differentiating sexual capabilities in A. urmiana than N. oculata. As shown in Fig.1, A. urmiana fed on T. suecica indicated a lower growth rates than A. urmiana fed on N. oculata on days 8, 11, 14 and 17. However, the growth rate of A. urmiana fed on T. suecica was higher than A. urmiana fed on N. oculata from day 17 to 20 (Fig. 1). This suggested that A. urmiana fed on T. suecica grew to adults at the end of the trial period (day 20), but A. urmiana fed on N.oculata did not reach the length or differentiation that could produce cysts nauplius. The comparison reproduction characteristics between A. urmiana fed on D. tertiolecta and T. suecica showed that D. tertiolecta had reproduction outcomes A.urmiana than T.suecica.

We can conclude that D. tertiolecta has higher potential in creating better reproductive characteristics urmiana than other algae. respect, T. suecica is located after D. tertiolecta and before Chlorella sp. and Chaetoceros sp. is at the end of this list. In general, the results of the present study indicated that D. tertiolecta had higher efficiency than the two other microalgae on A. urmiana in terms of length growth, survival rates and reproduction outcomes. Therefore, D. tertiolecta is suggested as a preferable food for A. urmiana. Hannah et al. (2013) evaluated the nutritional value of four microalgae namely Chaetoceros Skeletonema calcitrans. coastaum. Dunaliella salina and D. bardawil for Artemia sp. nauplii. They concluded that among the four microalgae tested, D. salina could be used as a potential live feed to improve the nutritional status of Artemia sp. as nauplii. Their finding was notconsistent with our results and they suggested that another species of Dunaliella (i.e. D. tertiolecta) was preferable food source for Artemia.

In the natural habitat of *A.urmiana* (i.e. Urmia Lake) *Dunaliella* spp compose more than 90% of the total algal density (Mohebbi, 2010).

References

- Amat D.F. 1980. Differentiation in Artemia strains from Spain. In: The brine shrimp Artemia. Vol. 1.
 Persoone, G.; Sorgelooa, P.; Roels, O. and Jaspers, E. (Eds). Univesa Press. Wettern. Belgium. 19-39.
- **Boone, E. and Bass-Becking, L.G.M.,** 1931. Salt effects on eggs and nauplii of *Artemia salina* L., *Journal of General Physiology*, 14(6), 753-763.
- **Brown, M.R., Jeffrey, S.W., Volkman, J.K. and Dunstan, C.A., 1997**. Nutritional properties of microalgae for mariculture. *Aquaculture,* 151, 315-331.
- Coutteau, P., Brendonck, L., Lavens, P. and Sorgeloos, P., 1992. The use of manipulated baker's yeast as an algal substitute for the laboratory

- culture of Anostraca . *Hydrobiologia* 234, 25-32.
- Czygan, F., 1968. Sekundar-Carotinoide in Grünalgen. I. Chemie, Vorkommen und Faktoren welche die Bildung dieser Polyene beinflussen. *Archives of Microbiology*, 61, 81-102.
- Dunstan, G.A., Volkman, J.K., Jeffrey, S.W. and Barrett, S.M., 1992. Biochemical composition of microalgae from the classes Chlorophyceae and Prasinophyceae. 2. Lipid classes and fatty acids. *Journal of Experimental Marine Biology and Ecology*, 161,115-134.
- Fábregas, J., Otero, A., Morales, E., Cordero, B. and Patiño, M., 1996. Tetraselmis suecica cultured in different nutrient concentrations varies in nutritional value to Artemia. Aquaculture, 143,197-204.
- Fábregas, J., Otero, A., Morales, E., Arredondo- Vega, B.O. and Patiño, M., 1998. Modification of the nutritive value of *Phaeodactylum tricornutum* for *Artemia* sp. in semicontinuous cultures. *Aquaculture*, 169, 167-176.
- Garcia Ulloa, M. and Garcia Olea, J. 2004. Reproductive performance of the Guppy fish Peocilia reticulate (Peters, 1859) fed with live Artemia franciscana cultured with inert and live diets. Avences en Investigacion Agropecuarina, Vol. 8, No. 003, pp. 1-7.
- Guillard, R.R.L. 1975. Culture of phytoplankton for feeding marine invertebrates. pp 26-60. In Smith W.L. and Chanley M.H (Eds.)

- Culture of Marine Invertebrate Animals. Plenum Press, New York, USA.
- **Hafezieh, M. 2004.** Effect of Chaetocerus, chorella as food on growth and survival rate of *Artemia urmiana*, 64, 76-80.
- Hannah, C., Mani, M. and Ramasamy, R. 2013. Evaluation of the Biochemical Composition of Four Marine Algae and Its Nutritional Value for Brine Shrimp. IOSR Journal of Pharmacy and Biological Sciences, 6, 3, 47-51.
- Johnson, D.A., 1980. Evaluation of various diets for optimal growth and survival of selected life stages of Artemia. In: The brine shrimp Artemia (G. Persoone, P. Sorgeloos, O. Roels and E. Jaspers, eds.), Universa Press, Wetteren, Belgium, pp. 185-191.
- Lavens, P. and Sorgeloos, P., 1991.

 Production of *Artemia* in culture tanks.. In: Browne, R.A.; P. Sorgeloos and C.N.A. Trotman (Eds). *Artemia biology*. CRC press, Inc. Boca Ratón, Florida, USA., 317-350.
- Léger, P.H., Bengtson, D.A., Simpson, K.L. and Sorgeloos, P., 1986. The use and nutritional value of *Artemia* as a food source. *Oceanography and Marine Biology Annual Review*, 24, 521-623.
- Lora-Vilchis, M. C. and Voltolina, D. 2003. Growth and Survival of Artemia franciscana (Kellogg) Fed with Chaetoceros muelleri Lemmerman and Chlorella capsulata Guillard. Rev. Invest. Mar. 24(3):241-246.

- Majack, T.J., Rust, M.B., Massee, K.C., G.W., Kissil, Hardy, R.W. and Peterson, M.E., 2000. Bioencapsulation of erythromicin using adult brine shrimp, *Artemia franciscana* (Latreille). *Journal of Fish Diseases*, 23, 71-76.
- Maldonado-Montiel T.D.N.J. and Rodríguez-Canché L.G., 2005. Biomass production and nutritional value of *Artemia* sp. (Anostraca: Artemiidae) in Campeche, México. *Revista de Biología Tropical*, 53 (3-4), 447-454.
- Malpica Sanchez, A., T., Castro Barrera, Sandoval Trujillo, H., Castro Mejía, J., DeLara Andrade R. and Castro Mejía, G., 2004. Composición del contenido de ácidos grasos en tres poblaciones mexicanas de Artemia franciscana de aguas epicontinentales. Revista de Biología Tropical, 52, 297-300.
- Merchie, G., Lavens, P., Dhert, P., Dehasque, M., Neils, H., Leenheer, D.A. and Sorgeloos P., 1995. Variation of ascorbic acid content in different live food organisms. *Aquaculture*, 134(3-4), 325-337.
- Mohammadyari, A., 2002. Biometric, morphologic and life cycle studies on three Artemia populations from Iran. M.Sc Thesis. Biology Departement, Scince faculty, Tehran University. Tehran, Iran.
- Mohebbi, F., Esmaeili L., Negarestan H. and Ahmadi R., 2009. Dynamics of phytoplankton population in Urmia Lake: Consequences on *Artemia*. Proceedings of the

- International Symposium/ Workshop on Biology and Distribution of *Artemia*. Urmia, Iran.
- Mohebbi, F., 2010. The Brine Shrimp *Artemia* and hypersaline environments microalgal composition: a mutual interaction. *International Journal of Aquatic Science*, 1(1), 19-27.
- Radhakrishnan. E.V., Rekha Chakraborty, D., Thangaraja, R. and Unnikrishnan, C., 2009. Effect of Nannochloropsis salina on the survival and growth of phyllosoma of the tropical spiny lobster, **Panulirus** homarus L. under laboratory conditions. Journal of the Marine Biological Association of India, 51 (1), 52 - 60.
- Rocha, M. S., Garcia, E.C. and Henriques, H.F., 2003.

 Biomolecular Engineering, 20, 237-/242
- **Sick, L.V., 1976**. Nutritional effect of five species of marine algae on the growth, development and survival of the brine shrimp *Artemia salina*. *Marine Biology*. 35,69-78.
- **Sorgeloos, P., 1980.** Life history of the brine shrimp, Artemia. In: The brine shrimp Artemia. Universa Press, Wettern, Belgium, ixx-xxii.
- Sorgeloos P., Lavens P., Léger Ph., Tackaert W. and Versichele D., 1986. Manual for the culture and use of brine shrimp *Artemia* in aquaculture. Laboratory of Mariculture, State University of Ghent, Belgium. 319P.

- Sorgeloos, P. Coutteau, P., Dhert, P. Merchie, G. and Lavens, P., 1998. Use of brine shrimp Artemia spp., in larval crustacean nutrition; A review. Reviews in Fisheries Science, 6, 55-68.
- Teresita D.N.J. Maldonado-Montiel and Leticia G. Rodríguez-Canché. 2005. Biomass production and nutritional value of Artemia sp. (Anostraca: Artemiidae) in Campeche, México. Rev. Biol. Trop. (Int. J. Trop. Biol.) Vol. 53 (3-4): 447-454.
- Vojoodzadeh, H., Ghezelbash, F., Riahi, H. and Manaffar, R., 2007. A study on the growth and survival rates of three different Artemia with microalgae species Nannochloropsis oculata Dunaliella tertiolecta and **Tetraselmis** suecica. Iranian Journal of Fisheries Science. 4, 143-152.