Histopathological changes of gill, liver and kidney in Caspian kutum exposed to Linear Alkylbenzene Sulfonate

Naeemi A.[1*]; Jamili S.[1,2]; Shabanipour N.[3]; Mashinchian A.[1] and Shariati Feizabadi S.[4]

Received: December 2011 Accepted: August 2013

Abstract

The histopathological effects of Linear Alkylbenzene Sulfonate on the gill, liver and kidney tissues of the Caspian kutum, *Rutilus frisii kutum* were studied. The fish were exposed to three sublethal concentrations of anionic detergent, Linear Alkylbenzene Sulfonate (LAS) for short term intervals (192 h). Gill, liver and kidney samples were collected after 192 h of exposure to LAS and lesions were analyzed by light microscopy. The histological changes to gills were edema, fusion of lamellae and lamellar aneurism. Some alterations like reduction of the interstitial haematopoietic tissue, tubular shrinkage, degeneration in the epithelial cells of renal tubule and necrosis were observed in the kidney. In the liver tissue, hepatocyte degeneration, congestion and dilation of sinusoid and vacuolar degeneration were seen. It seems that sublethal concentration of LAS may affect sever changes to gill, kidney and liver of *R. frisii kutum* specimens that leads to malfunction of these organs which cause damage to health of the fish.

Keywords: *Rutilus frisii kutum,* Linear Alkylbenzene Sulfonate (LAS), histopathology, gill, liver, kidney

1-Department of Marine Biology, Graduate School of Marine Science and Technology, Science and Research Branch, Islamic Azad University, Tehran, Iran

2- Iranian Fisheries Research Organization, P.O.Box: 14155-6116, Tehran, Iran

3- Department of Biology, Faculty of Science, University of Guilan, Rasht, Iran

4- Department of Chemistry, Science and Research Branch, Islamic Azad University, Guilan, Iran

✱Corresponding author's email: akramnaeemi@yahoo.com

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

Introduction

Linear Alkylbenzene Sulphonate (LAS) is the most widely utilized synthetic anionic surfactant for cleaning products (Hampel et al., 2004), with an annual global production rate of 4 mt (HERA-LAS, 2004). In waste water treatment plants, LAS is removed in up to 99%, but certain amounts of this compound may reach rivers and coastal waters, causing risk to aquatic life (Hampel et al., 2008). Most of domestic sewage flows into the Caspian Sea directly or through rivers without sewage treatment (UNIDO Caspian pollution report, 1998). In cases of untreated wastewater discharge, concentrations of LAS may reach higher concentrations at the impact zone (Hampel et al., 2004).

Water pollution produces pathological alterations in fish. These exposure concentrations may not be lethal for the affected species, but may influence internal functions and structures which under chronic exposure conditions could have effects on the effectiveness of vital functions and processes such as competitive stress and environmental resistance, reproduction, growth, etc (Duft et al., 2003; Hampel et al., 2008).

Histopathological changes have been widely used as biomarkers in the evaluation of the health of fish exposed to contaminants (Thophon et al., 2003). Gill, kidney and liver are responsible for vital functions, such as respiration, excretion and the accumulation and biotransformation of xenobiotics in the fish. One of the great advantages of using histopathological biomarkers in environmental monitoring is that this category of biomarkers allows examining these specific target organs (Gernhofer et al., 2001). Furthermore, the pathological changes discovered in these organs are normally easier to identify than functional ones (Fanta et al., 2003), and serve as warning signs of damage to animal health (Hinton and Laurén, 1990).

One of the most important and economic aquatics of Caspian Sea is Kutum (*R. frisii kutum*) (Babazadeh et al., 2008) which devoted approximately 78% of bony fish harvested (Afraei Bandpei et al., 2011). Research on the histopathological effects of LAS in the gill, kidney and liver of Caspian Sea kutum has not been done. So, this is the first report for assessment of histopathological changes in the gill, kidney and liver of Caspian kutum due to exposure to sublethal concentration of LAS.

.

Materials and methods

The juvenile (0/5- 1 g) Caspian Kutum, *R. frisii kutum* were obtained from rearing unit of Shahid Ansari State, Rasht, Iran. Commercial LAS, a mixture of C10-C13 homologues with all positional isomers except 1-phenyl and an average molecular weight of 343 (sodium salt derivative), was supplied by Paxan Company (Tehran, Iran). Fishes were kept in a stock tank for one week for acclimation.

Exposure Conditions: The fish were tested in a continuous flow-through system, comprising 3 treatments and one blank in 3 replications (totally 12 tanks). The exposure solution was renewed completely each day in order to ensure constant concentrations. The tests were done in 192 h. Physical and chemical factors of water, such as temperature, pH and dissolved oxygen were maintained at appropriate values

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

similar to standard conditions (temperature 20±1°C; pH 7.9 ±0.1; %DO between 60-100%).Three sublethal concentrations (1:20, 1:10 and 1:5 of the 96 h LC50) were chosen, based on the 96 h LC50 value, 11/6 mg L^{-1} (Gholami et al., 2010). So, the fishes exposed to 0/58, 1/16 and 2/32 mg L^{-1} concentrations of commercial LAS. The control tank was kept under the same conditions without adding of surfactant. Sampling was done after 192 h from onset of experiment. Daily Samples of water were taken and measured for ensuring that the concentration of toxicant maintained as near as possible to the nominal value.

Water analysis: Solid-phase Extraction technique with ODS SPE columns (SPE-C18 purchased from Applied Separations) was used for extraction of water samples (50 mL). LAS was eluted from the column with 5 mL of CH3OH (Merck). After evaporation of the solvent, the sample was dissolved in 1 ml of CH3OH (Merck) and transferred to HPLC vials. LAS concentration in water was determined by reversed-phase HPLC with fluorescence detection (Tolls et al., 1999).

Histological analysis: Ten fish per treatment were anesthetized and the gills, livers and kidneys were removed. Tissues were placed in prelabeled, individual glass vials in which Bouin's fixative was added. After 24 h immersion fixation, tissues were transferred to coded glass vials containing 70% ethanol. Tissues dehydrated in concentrations of alcohol, and embedded in paraffin wax for microtome sectioning (5 µm) (Poosti and Marvasti, 1999).Sections were mounted onto slides and after hydration in ethanol series of descending concentration, were stained with standard haematoxylin and eosin (H&S) stain. 10 sections of each tissue from each fish were examined by light microscope (Kazemi and Bahmani, 1998).

Results

Water analysis :Analysis of water samples showed that the concentration of LAS in water was maintained ± 10% of selected concentration.

Control gill tissues: The gills from control specimens showed normal conditions. i.e. fresh red color and the gill lamella were arrange normally (Fig.1).

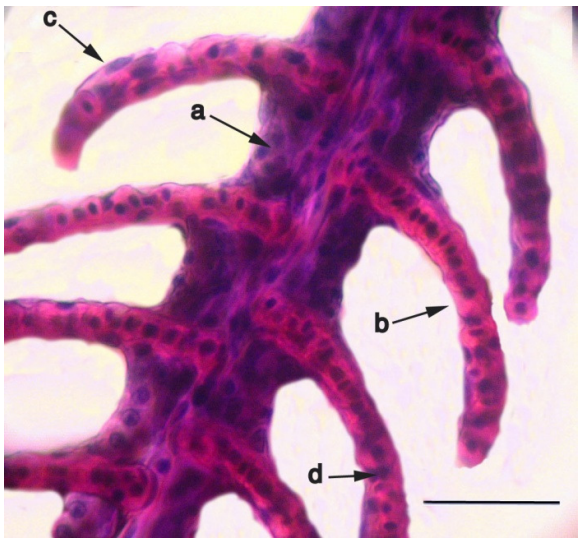


Figure 1: normal Gill tissue of *Rutilus frisii kutum* exposed to 0.00 ppm LAS. H&E.Bar:25µm; (a) primary lamellae (b) secondary lamellae (c) lamellar epithelial cells (d) pillar cell

Treated gill tissues: In the fish exposed to 0.58 mg L^{-1} of LAS for 192 h , hyperplasia of the lamellar epithelial cells, lamellar fusion and edema were observed (Fig. 2). Lamellar aneurysm, increasing the number of epithelial

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

cells and fusion of adjacent lamellae were seen in the gills of fish examined after 192 hours of exposure to 1.16 mg L^{-1} of LAS (Fig.3). In the case of gills that exposed to 2.32 mg L^{-1} LAS, the gills were more damaged that individual lamellae were indistinguishable (Fig. 4).



Figure 2: Gill tissue of *Rutilus frisii kutum* exposed to 0.58 ppm LAS after 192h. H&E. Bar: 25µm. (a) fusion of adjacent lamellae (b) edema

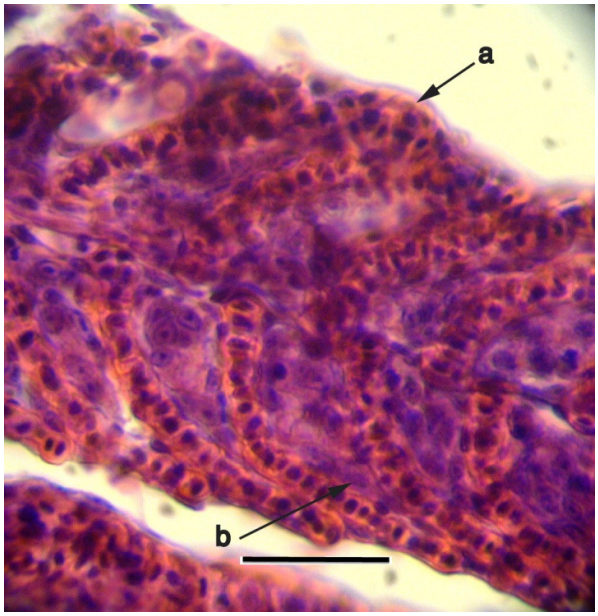


Figure 3: Gill tissue of *Rutilus frisii kutum* exposed to 1.16 ppm LAS after 192h. H&E. Bar: 25µm (a) Lamellar aneurysm (b) fusion of lamellae

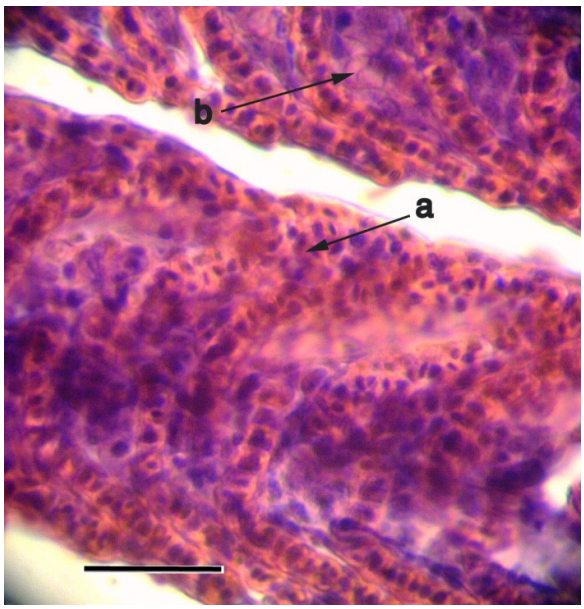


Figure 4: Gill tissue of *Rutilus frisii kutum* exposed to 2.32 ppm LAS after 192h. H&E.Bar:25µm. (a) lamellar aneurysm (b) full fusion of lamellae

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

Control of kidney tissues: Histopathological changes were not observed in the kidney of the control fishes. The structural details of the kidney of control *R. frisii kutum* are shown in Figure 5. Like other teleosts, the basic unit of kidney is composed of a renal corpuscle, Bowman's capsule and glomerulus and various segment of the renal tubules. These tubules are surrounded by an interstitial lymphoid – hematopoietic tissue.

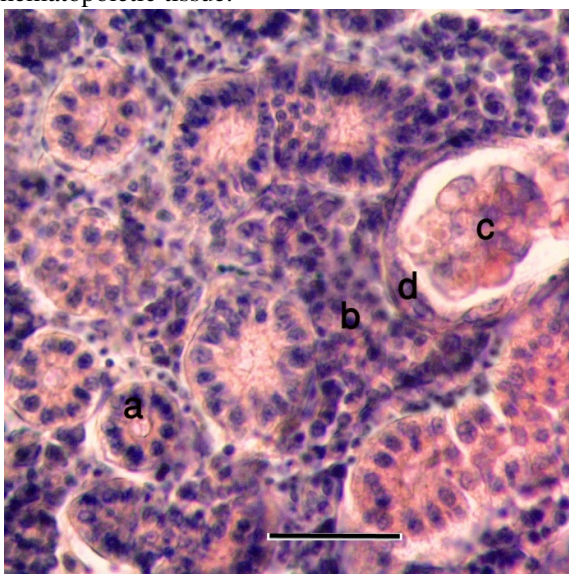


Figure 5: Normal kidney tissue of *R. frisii kutum* exposed to 0.00 mg L^{-1} LAS after 192h. H&E.Bar:25µm; (a) renal tubules (b) hematopoietic tissue (c) glomerulus (d) Bowman's capsule

Treated kidney tissues: The fish exposed to 0.58 mg L^{-1} LAS for 192 h showed a moderate reduction of the interstitial haematopoietic tissue and tubular shrinkage (Fig. 6). These alterations as well as degeneration in the epithelial cells of renal tubule were observed in the fish exposed to1.16 mg L^{-1} of LAS (Fig.7). More reduction of the interstitial haemocytopoietic tissue, necrosis, tubular degeneration and tubular shrinkage were seen at the highest exposure concentration of the LAS (2.32 mg L^{-1}) (Fig.8).

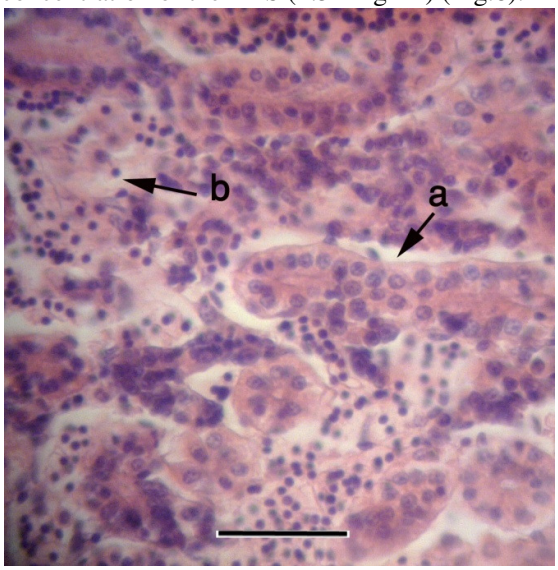


Figure 6: Kidney tissue of *R. frisii kutum* exposed to 0.58 mg L^{-1} LAS after 192h. H&E.Bar:25µm. (a) tubular shrinkage (b) reduction of the interstitial haematopoietic tissue.

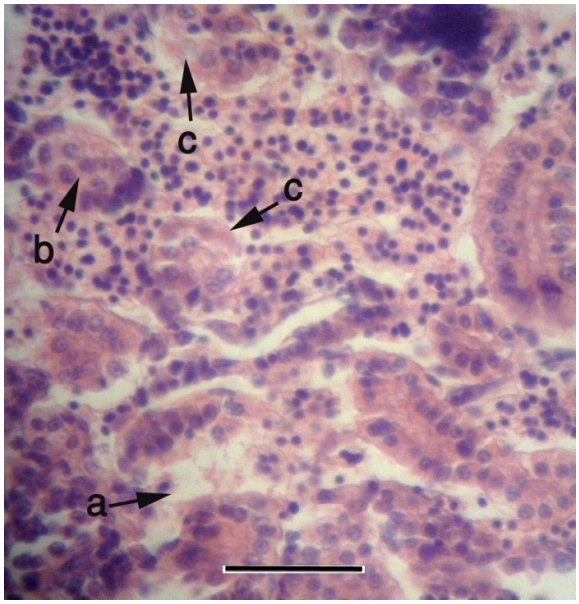


Figure 7: Kidney tissue of *R. frisii kutum* exposed to 1.16 mg L^{-1} LAS after 192h. H&E. Bar:25µm. (a) reduction of the interstitial haematopoietic tissue (b) tubular shrinkage (c) degeneration in the epithelial cells of renal tubule

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

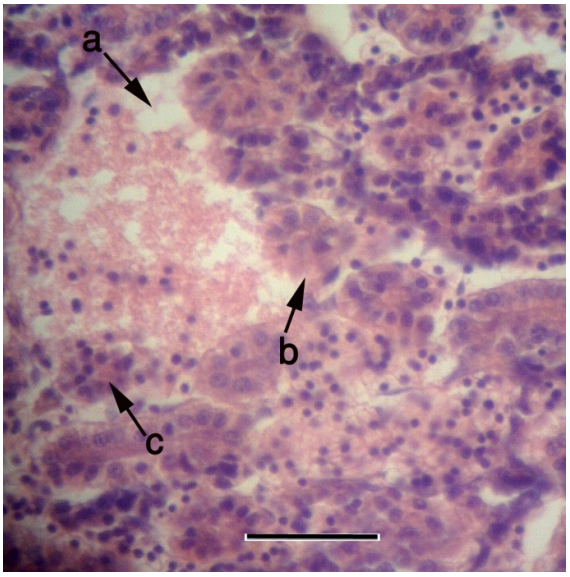


Figure 8: Kidney tissue of *R. frisii kutum* exposed to 2.32 mg L^{-1} LAS after 192h. H&E. Bar: 25µm. (a) necrosis and reduction of the interstitial haematopoietic tissue (b) tubular degeneration (c) luminal occlusion and tubular shrinkage.

Control of liver tissues: Liver histology from control fish is illustrated in Figure 9. No histopathological changes were observed in the liver of the control fish and the hepatic parenchyma shows regular distribution of hepatocytes arranged around the vascular system (sinusoids) (Fig.9). *Treated liver tissues*

In the liver tissues of fish exposed to 0.58 mg L^{-1} LAS, congestion of sinusoid was observed (Fig.10). At exposure concentrations of 1.16 mg L^{-1}, more congestion and dilation of sinusoid were seen (Fig.11). In fish exposed to 2.32 mg L^{-1} of LAS, the histological alterations of liver were more evident and some other changes such as hepatocyte degeneration and vacuolar degeneration were detected (Figs.12 and 13).

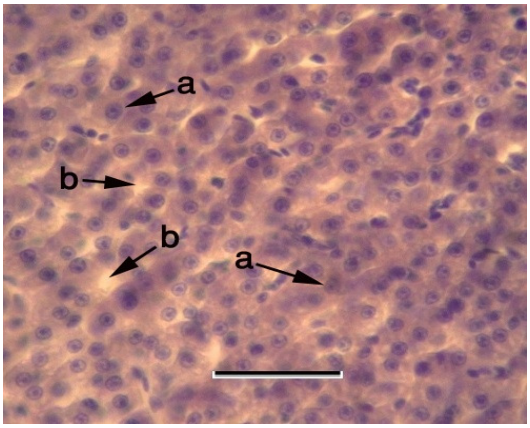


Figure 9: Normal liver tissue of *R. frisii kutum* exposed to 0.00 mg L^{-1} LAS after 192h. H&E.Bar:25µm; (a) hepatocytes with a nucleus (b) sinusoid.

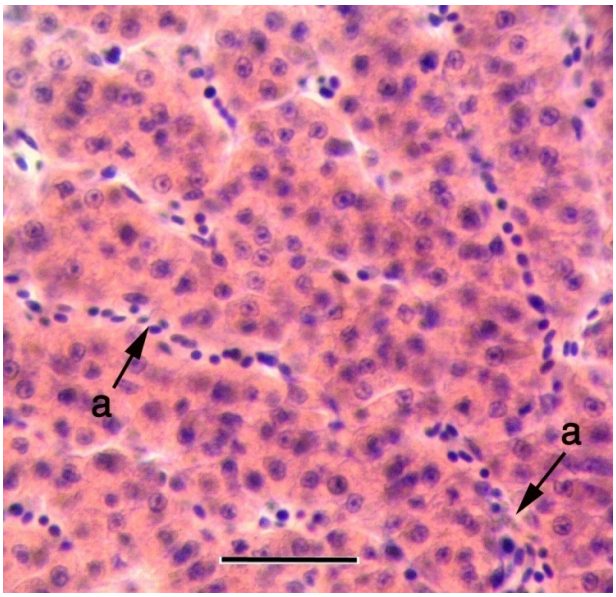


Figure 10: Liver tissue of *R. frisii kutum* exposed to 0.58 mg L^{-1} LAS after 192h. H&E.Bar:25µm. (a) congestion of sinusoid.

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

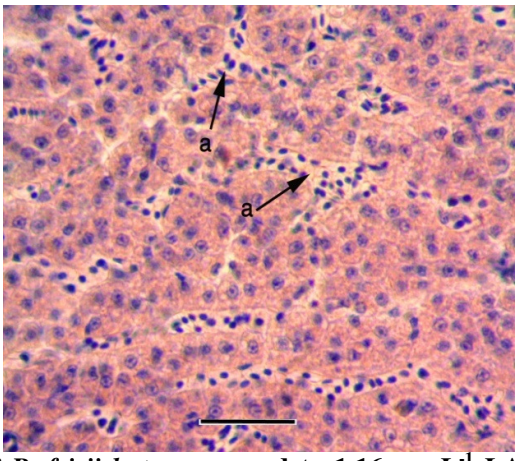


Figure 11: Liver tissue of *R. frisii kutum* exposed to 1.16 mg L^{-1} LAS after 192h. H&E. Bar: 25µm. (a) congestion and dilation of sinusoid.

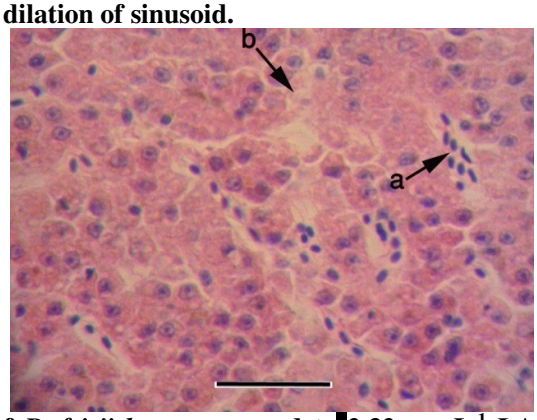


Figure 12: Liver tissue of *R. frisii kutum* exposed to 2.32 mg L^{-1} LAS after 192h. H&E. Bar: 25µm. (a) congestion and dilation of sinusoid (b) hepatocyte degeneration.

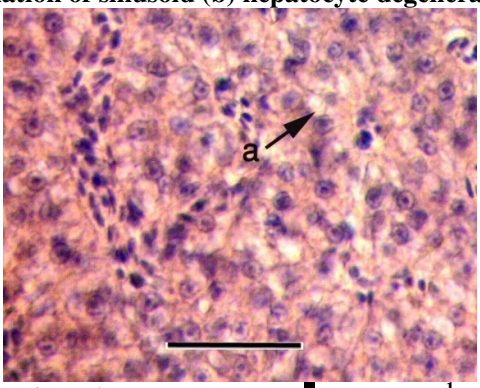


Figure 13: Liver tissue of *R. frisii kutum* exposed to 2.32 mg L^{-1} LAS after 192h. H&E. Bar: 25µm. (a) vacuolar degeneration.

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

Discussion

According to Roberts and Rodger (2001), massive lamellar hyperplasia may leading to secondary lamellar fusion. Thickening of the primary filament epithelial padding due to edema, cellular hyperplasia and fusion of the secondary lamellae are defense mechanisms that could impair blood-water exchange by reducing the surface area of the secondary lamellae that is in contact with the water (Kelly, 2007). Such alterations are non-specific and may be induced by different types of contaminant (Mallatt, 1985). The formation of an aneurysm is related to the rupture of the pillar cells (Martinez et al., 2004) due to a bigger flow of blood or even because of the direct effects of contaminants on these cells. This is a severe type of lesion, recovery from which is more difficult than the epithelial changes (Poleksic and Mitrovic-Tutundzic, 1994).

In the present study several change of gill tissue like hyperplasia, edema, fusion of lamella and lamellar aneurysm were detected in the gills of fish after 192 hours of exposure to three sublethal concentrations of LAS. Some changes in the gill architecture of fish have been observed after acute exposure to a sublethal concentration of LAS, such as epithelial lifting and fusion of gill lamellae and stagnation of gill vessels (Alvarez-Munoz et al., 2009). However hypertrophy, hyperplasia, fusion of adjacent lamella and telengeastases of the gill were noticed when fish exposed to LAS for chronic test (Hampel et al., 2008; Rejeki et al., 2008).

The teleostean kidney is one of the first organs to be affected by contaminants in the water (Thophon et al., 2003) and receives the largest proportion of postbranchial blood. Therefore renal lesions might be expected to be good indicators of environmental pollution (Cengiz, 2006; Hampel et al., 2008). In the kidney of *R. frisii kutum* after acute exposure to LAS, histological changes like reduction of the interstitial haematopoietic tissue, tubular shrinkage, degeneration in the epithelial cells of renal tubule and necrosis were noticed. Reduction of the interstitial haematopoietic tissue and epithelial hyperplasia with luminal occlusion of renal tubules have been reported in kidney of *Solea senegalensis*, when exposed to LAS in a chronic test (Hampel et al., 2008). Degeneration in the epithelial cells of renal tubule and narrowing of the tubular lumen has been shown in kidney of fishes exposed to deltamethrin (Cengiz, 2006). Rosety et al. (2002) reported tubular retraction in the kidney of *Scophthalmus maximus L.* exposed to the sodium dodecyl sulfate. The shrunk lumen of tubules are suggestive of hindered tubular reabsorption (Bhatnagar et al., 2007). Camargo and Martinez (2007) reported some alteration like tubular degeneration and luminal occlusion in the Neotropical fish species *Prochilodus lineatus*, caged in an urban stream. In severe cases, the degenerative process can lead to tissue necrosis (Takashima and Hibiya, 1995). In the present study tissue necrosis in *R. frisii kutum* specimens was seen when exposed to the highest concentrations of LAS.

The organ most associated with the detoxification and biotransformation process is the liver, and due to its function, position and blood supply (Van der Oost et al., 2003), it is

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

also one of the organs most affected by contaminants in the water (Rodrigues and Fanta, 1998). Alterations in the liver may be useful as markers that indicate prior exposure to environmental stressors (Velmurugan et al., 2009). Congestion and dilation of sinusoid, hepatocyte and vacuolar degeneration were detected in *R. frisii kutum* exposed to sublethal concentrations of LAS. Several studies demonstrated that alterations in number, size and shape of the hepatocyte nucleus can be due to contaminants (Paris-Palacios et al., 2000; Camargo and Martinez, 2007; Figueiredo-Fernandes et al., 2007; Hampel et al., 2008).Congestion of sinusoid, shrinkage of hepatocytes, slight atrophy and vacuolar degeneration have been reported in liver of fishes after chronic exposure to LAS (Hampel et al., 2008; Rejeki et al., 2008). Congestion is a blood circulation disturbance due to the increase volume of the blood in the blood capillary. Vacuolar degeneration is known as an acute swelling of the organ (Rejeki et al., 2008).

In this research, the gill, liver and kidney damages were dependent to the concentrations of the LAS. When comparing histopathlogical changes in the three concentrations, 2/32 mg L^{-1} shows more damages than1/16 mg L^{-1}, and 0/58 mg L^{-1} had the lowest irritation. So, the higher concentration of LAS shows more damage than the lower ones. Some other studies have shown this result for LAS (Hampel et al., 2008; Rejeki et al., 2008) and some other pollutants (Cengiz and Nlu, 2002; Regar and Bhatnagar, 2006). The tissue damages of the fish exposed to LAS detergent may be due to the accumulation of the detergent in them (Rejeki et al., 2008).

All the histopathological observation indicated that exposure to sublethal concentrations of LAS caused destructive effect in the gill, kidney and liver tissues of *R. frisii kutum*. However, complementary studies are necessary for a better understanding of its deleterious effects.

References

Afraei Bandpei, M.A., Mashhor, M., Abdolmaleki, S.H., Najafpour, S.H., Bani, A., Pourgholam, R., Fazli, H., Nasrolahzadeh, H., and Janbaz, A.A., 2011. The Environmental effect on spawning time, length at maturity and fecundity of kutum (*Rutilus frisii kutum*, Kamensky, 1901) in Southern Part of Caspian Sea, Iran. *Iranica Journal of Energy & Environment*, 2 (**4**): 374-381.(in Persian)

Alvarez-Munoz, D., Gomez-Parra, A., Blasco, J., Sarasquete, C. and Gonzalez-Mazo, E., 2009. Oxidative stress and histopathology damage related to the metabolism of dodecylbenzene sulfonate in Senegalese sole. Chemosphere, 74(**9**):1216-23.

Babazadeh, M., Kousha, A. and Safari, R., 2008. Changes in Nutritional Factors of Freezed Kutum (*Rutilus frisii kutum*). *World Journal of Zoology*, 3,51-53.

Bhatnagar, C., Bhatnagar, M. and Regar, B.C., 2007. Fluoride-induced histopathological changes in gill, kidney, and intestine of freshwater teleost. *Labeo rohita*. Research report Fluoride, 40(**1**):55–61.

Camargo, M. M. P. and Martinez, C. B. R., 2007. Histopathology of gills, kidney and liver

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

of a Neotropical fish caged in an urban stream. *Neotropical Ichthyology*, 5(**3**):327-336.

Cengiz, E.I., 2006. Gill and kidney histopathology in the freshwater fish *Cyprinus carpio* after acute exposure to deltamethrin. *Environmental Toxicology and Pharmacology*, 22, 200–204.

Cengiz, E.I. and Nlu, E.U., 2002. Histopathological changes in the gills of Mosquitofish, *Gambusia affinis* exposed to Endosulfan. *Bulletin of environmental contamination and toxicology*, 68,290–296.

Duft, M., Schulte-Oehlmann, U., Weltje, L., Tillmann, M. and Oehlmann, J., 2003. Stimulated embryo production as a parameter of estrogenic exposure via sediments in the freshwater mudsnail *Potamopyrgus antipodarum*. *Aquatic Toxicology*, 64, 437-449.

Fanta, E., Rios, F. S., Romão, S., Vianna, A. C. C. and Freiberger, S., 2003. Histopathology of the fish *Corydoras paleatus* contaminated with sublethal levels of organophosphorus in water and food. *Ecotoxicology and Environmental Safety*, 54, 119-130.

Figueiredo-Fernandes, A., Ferreira-Cardoso, J.V., Garcia-Santos, S., Monteiro, S.M. and Carrola, J., M., 2007. Histopathological changes in liver and gill epithelium of Nile tilapia, *Oreochromis niloticus*, exposed to waterborne copper. *Pesquisa Veterinária Brasileira,* 27**(3):**103-109.

Gernhofer, M., Pawet, M., Schramm, M., Müller, E. and Triebskorn, R., 2001. Ultrastructural biomarkers as tools to characterize the health status of fish in contaminated streams. *Journal of Aquatic Ecosystem, Stress and Recovery*, 8,241-260.

Gholami, M., Fatemi, M.R., Falahi, M., Esmaili, A. and Mashinchiyan, A., 2010. Effects of heavy metals (Copper and Cadmium) and detergent (LAS) on white fish fry *Rutilus frisii kutum*. *Research Journal of Environmental Toxicology*, 4,231-236.

Hampel, M., Ortiz-Delgado, JB., Moreno-Garrido, I., Sarasquete, C. and Blasco, J., 2004. Sublethal effects of linear alkylbenzene sulphonate on larvae of the seabream (*Sparus aurata*): histological approach**.** *Histology and Histopathology,* 19**,**1061-1073.

Hampel, M., Ortiz-Delgado, J.B., Sarasquete, C. and Blasco, J., 2008. Effects of sediment sorbed linear alkylbenzene sulphonate on juveniles of the Senegal sole, *Solea senegalensis*: toxicity and histological indicators. *Histology and histopathology***,** 23, 87-100.

HERA-LAS. Human and Environmental Risk Assessment LAS CAS No 68411-30-3. Version No. 9., 2004. (http://www.heraproject.com/Index.cfm).A.I.S. E. Association Internationale de la Savonnerie, de la Détergence et des Produits d'Entretien (www.aise-net.org), Cefic. European Chemical Industry Council (www.cefic.org).

Hinton, D.E. and Laurén, D.J., 1990. Integrative histopathological approaches to detecting effects of environmental stressors in fishes. *American Fisheries Society Symposium*, 8,51-66.

Kazemi, R. and Bahmani, M., 1998. Instructions for preparing and staining tissues for histological studies. International Institute of sturgeon Dr Dadman. 16P.(in persian)

Kelly, J.M., 2007. Ecotoxicological assessment of juvenile northern pike inhabiting lakes downstream of a Uranium mill. Master thesis,University of Saskatchewan.144P.

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

Mallatt, J., 1985. Fish gill structural changes induced by toxicants and other irritants: a statistical review. *Canadian Journal of Fisheries and Aquatic Science*. 42,630-648.

Martinez, C.B.R., Nagae, M.Y., Zaia, C.T.B.V. and Zaia, D.A.M., 2004. Morphological and physiological acute effects of lead in the neotropical fish *Prochilodus lineatus*. *Brazilian Journal of Biology*, 64(**4**):797-807.

Paris-Palacios, S., Biagianti-Risbourg, S. and Vernet, G., 2000. Biochemical and (ultra) structural hepatic perturbation of *Brachydanio rerio* (Teleostei, Cyprinidae) exposed to two sublethal concentrations of copper sulphate. *Aquatic Toxicology*, 50,109-124.

Poleksic, V. and Mitrovic-Tutundzic, V., 1994. Fish gills as a monitor of sublethal and chronic effects of pollution.Oxford, Fishing News Books. pp.339 – 352.

Poosti, A.S. and Marvdasti, S., 1999. Atlas of fish histology (normal and pathological forms). Tehran University Press, 328P.(in persian)

Regar, B.C. and Bhatnagar, C., 2006. Histopathological changes in the gill architecture of freshwater teleosts, *Labeo rohita* exposed to sodium fluoride. *Journal of Herbal Medicine and Toxicology*, 1,35-41.

Rejeki, S., Desrina, D. and Mulyana, A.R., 2008. Chronic affects of detergent surfactant (Linear Alkylbenzene Sulfonate / LAS) on the growth and survival rate of sea bass (*Lates calcalifer Bloch*), larvae Journal of Coastal Development.213P.

Roberts, R.J. and Rodger, H.D., 2001. The pathophysiology and systematic pathology of teleosts. in fish pathology (3eds.). Ronald J. Roberts. Harcourt Publishers Limited, London.

Rodrigues, E. L. and Fanta. E., 1998. Liver histopathology of the fish *Brachydanio rerio* after acute exposure to sublethal levels of the organophosphate Dimetoato 500. *Revista Brasileira de Zoologia*, 15,441-450.

Takashima, F. and Hibya, T., 1995. An atlas of fish histology: normal and pathological features. 2nd ed., Tokyo, Kodansha.195P.

Thophon, S., Kruatrachue, S., Upathan, E., Pokethitiyook, P., Sahaphong, S. and Jarikhuan, S., 2003. Histopathological alterations of white seabass, *Lates calcarifer* in acute and subchronic cadmium exposure. *Environmental Pollution,* 121,307-320.

Tolls, J., Haller, M. and Sijm, D.T.H.M., 1999. Extraction and Isolation of Linear Alkylbenzenesulfonate and Its Sulfophenylcarboxylic Acid Metabolites from Fish Samples. *Analytical chemistry*, 71,5242–5247.

UNIDO Caspian pollution report, 1998. Main sources of industrial pollution Available via. http://www.caspianenvironment.org/ Main sources of industrial pollution.

Van der Oost, R., Beber, J. and Vermeulen, N. P. E., 2003. Fish bioaccumulation and biomarkers in environmental risk assessment: a review. *Environmental Toxicology and Pharmacology*, 13,57-149.

Velmurugan, B., Selvanayagam, M., Cengiz, E.L. and Unlu, E., 2009. Histopathological Changes in the Gill and Liver Tissues of Freshwater Fish, *Cirrhinus mrigala* Exposed to Dichlorvos, *Brazilian archives of biology and technology*, 5,1291-1296.

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]

تغییرات بافت شناسی آبشش، کبد و کلیه ماهی سفید دریای خزر در معرض آلاینده آلکیل بنزن سولفونات خطی

اکرم سادات نعیمی*[1]، شهلا جمیلی[1و2]، نادر شعبانی پور[3]، علی ماشینچیان [1] و شهاب شریعتی فیض آبادی[4]

تاریخ دریافت: آذر ۱۳۹۰ تاریخ پذیرش:مرداد ۱۳۹۲

چکیده:

در تحقیق حاضر اثرات بافت شناسی آلاینده آلکیل بنزن سولفونات خطی (Linear Alkylbenzene Sulfonate) بر بافتهای آبشش، کبد و کلیه ماهی سفید دریای خزر (Rutilus frisii kutum) مطالعه گردید. ماهیان در معرض سه غلظت تحت کشنده شوینده آنیونی، آلکیل بنزن سولفونات خطی، به مدت ۱۹۲ ساعت قرار گرفتند. نمونه گیری از بافت های آبشش، کبد و کلیه پس از قرارگیری در معرض آلاینده LAS انجام شد و آسیب ها توسط میکروسکوپ نوری بررسی گردیدند. تغییرات بافت شناسی آبشش شامل ادم، اتصال لاملاها و آنوریسم بودند. در بافت کلیه کاهش بافت بینابینی، چروکیدگی توبولها ، دژنراسیون سلولهای اپی تلیالی توبولها و نکروز مشاهده گردید. بافت کبد دژنراسیون هپاتوسیتها، پرخونی و تورم در سینوزوئیدها و دژنراسیون واکوئولی را نشان داد. طیق نتایج به نظر می رسد که غلظت تحت کشنده آلکیل بنزن سولفونات خطی(LAS) تغییرات شدیدی را در بافتهای آبشش، کبد و کلیه گونه ماهی سفید دریای خزر ایجاد میکند که می تواند منجر به اختلالات عملکردی این اندامها و در نهایت باعث آسیب به سلامت ماهی گردد.

کلمات کلیدی: ماهی سفید دریای خزر (*Rutilus frisii kutum*)، آلکیل بنزن سولفونات خطی، آسیب بافتی، آبشش، کبد و کلیه

۱-گروه بیولوژی دریا، واحد علوم وتحقیقات، دانشگاه آزاد اسلامی، تهران، ایران

۲- مرکز تحقیقات شیلات ایران، تهران، ایران

۳- گروه زیست شناسی، دانشکده علوم پایه، دانشگاه گیلان

۴- گروه شیمی، واحد علوم وتحقیقات، دانشگاه آزاد اسلامی، گیلان، ایران

آدرس الکترونیکی نویسنده مسئول: akramnaeemi@yahoo.com

[Downloaded from jifro.ir on 2026-10-30]

[DOR: 20.1001.1.15622916.2013.12.4.15.4]