Research Article

Cultivations of *Arthrospira maxima* (Spirulina) using ammonium sulfate and sodium nitrate as an alternative nitrogen sources

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Abstract

Arthrospira (Spirulina) has been considered as an attractive microalgae in all aspects of human life including medicine, cosmetics, and food. Nitrogen source is an important cost-saving factor in large-scale cultivation. In the present study, the cultivation of *S. maxima* was studied by replacing the basic-nitrogen source of Zarrouk’s medium (2.5 gL\(^{-1}\)) with concentration ranges of 0-10 gL\(^{-1}\) for sodium nitrate and 0-5 gL\(^{-1}\) for ammonium sulfate in terms of biomass and phycobiliproteins production. Biomass and phycobiliprotein growth of different nitrogen sources have shown different effects on growth. The changes in the amount of cell dry weight as a function of sodium nitrate did not show significant changes relating to its concentration. In case of ammonium sulfate, the cell dry weight of *S. maxima* without nitrogen source was 0.835 gL\(^{-1}\) during five days of cultivation. Moreover, phycocyanin and allophycocyanin contents were 0.053 and 0.072 mgL\(^{-1}\), respectively, while phycobiliproteins content and cell dry weight were decreased by increasing further concentration. There was a significant difference among the culture mediums containing ammonium sulfate and without nitrogen source in terms of concentration of biomass and phycobiliprotein. The highest and lowest results for cell dry weight and phycobiliprotein production were obtained from the treatment with nitrogen starvation and 5 gL\(^{-1}\) ammonium sulfate, respectively. Finally, nitrogen starvation was proved as a feasible way to grow and could be good candidate for biomass growth and phycobiliprotein.

Keywords: *Arthrospira maxima*, Nitrogen sources, Ammonium sulfate, Sodium nitrate

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Introduction
Natural pigments are current interest in the market because of their color functions and physiological activity compared with synthetic colorant agents (Sigurdson et al., 2017). Furthermore, there is a massive demand for natural coloring agents in industries like food, drug, and cosmetics (Clydesdale, 1993; Wissgott and Bortlik, 1996). Most recently, the feasibility of natural pigments extraction from microalga has attracted considerable interest worldwide, while the use of most or all synthetic colorants has been banned by FDA (Kevin B. Hicks et al., 1992; Beatriz et al., 2015).

Among these pigments, applications of phycocyanin and allophycocyanin as a natural color from alga have massive values in nutraceutical, cosmetics, and pharmaceutical industries owing to their health benefits (Kuddus et al., 2013). Currently, Spirulina is the widely available microalga source that contains a high value of phycocyanin and allophycocyanin pigments (Park et al., 2018). Among various species of this algae, S. platensis and S. maxima are the two most important species (Sánchez et al., 2003). S. maxima is a photosynthetic, filamentous, and multicellular blue-green cyanobacterium with a long history of use as food (Ashton Acton, 2013). Traditionally, S. maxima was harvested from the Texcoco lake for human consumption (Vo et al., 2015).

According to researchers, Spirulina production is mainly depending on nutrient availability, temperature, and light (Carvalho and Malcata, 2003; Kumar et al., 2011). Previously, a wide number of media have been developed for the Spirulina biomass growth like modified Zarrouk (Rajasekaran et al., 2016), revised medium (RM6) (Raoof et al., 2006), CFTRI, JPJM (Salunke et al., 2016), Convy, f/2, BG-11 (Dineshkumar et al., 2016), and Bangladesh medium (Khatun et al., 1970). Moreover, Zarrouk’s medium has been known as the conventional standard medium for Spirulina culture (Zarrouk, 1966; Raoof et al., 2006).

Nitrogen and carbon sources are the most cost-effective factor in the growth and pigment production of Spirulina (Soletto et al., 2005; Çelekli and Yavuzatmaca, 2009). Sodium nitrate is the common nitrogen source that has been used for Spirulina cultivation (Cost et al., 2001). The replacement of basic nitrogen sources of culture medium with cheaper materials such as urea, ammonium sulfate, and ammonium chloride have been previously studied in S. platensis cultivation (Carvalho et al., 2004; Bezerra et al., 2008; Ferreira et al., 2010; Avila-Leon et al., 2012; Matsudo et al., 2012).

To the best of our knowledge, biomass and pigment production of S. maxima has not been widely studied in evaluating the effect of nitrogen sources. Therefore, the present study aimed to evaluate the effect of two different nitrogen sources (sodium nitrate and nitrogen sulfate) at different concentrations on the specific growth
rate, cell dry weight, and phycobiliproteins production.

Materials and methods
Microorganism and Culture condition: S. maxima strain CIB79 was obtained from National Polytechnic University, (IPN, Mexico). The experiment was carried out at laboratory temperature using white fluorescent lamps (4,500 lux). The cultures were incubated under continuous illumination and aeration with air pump AC-9602 (RESUN, Mexico) during the process of growth. We studied the influence of two different nitrogen sources (i.e., sodium nitrate and ammonium sulfate) at various concentrations and cultivation time (Table 1) to estimate growth and phycobiliproteins production in the pure culture of Zarrouk’s medium. Nitrogen sources were replaced by the basic-nitrogen source of Zarrouk’s medium (2.5 g L⁻¹). Nitrogen sources were bath added in 125 ml flask with 40 ml of S. maxima stock (Initial OD₆₇₄ ≥ 0.4) during 8 days of cultivation. All experimental cultures were set up in triplicate. The experimental data was obtained as the mean ± SEM value for triplicate cultures in all treatments. The pH was monitored by a pH21 pH/mV meter (HANNA model).

<table>
<thead>
<tr>
<th>Treatment No.</th>
<th>Nitrogen source</th>
<th>Nitrate source concentration</th>
<th>Unit</th>
<th>Cultivation time (day)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Sodium Nitrate</td>
<td>1.25</td>
<td>gL⁻¹</td>
<td>8</td>
</tr>
<tr>
<td>3</td>
<td>Sodium Nitrate</td>
<td>2.5</td>
<td>gL⁻¹</td>
<td>8</td>
</tr>
<tr>
<td>4</td>
<td>Sodium Nitrate</td>
<td>5</td>
<td>gL⁻¹</td>
<td>8</td>
</tr>
<tr>
<td>5</td>
<td>Sodium Nitrate</td>
<td>10</td>
<td>gL⁻¹</td>
<td>8</td>
</tr>
<tr>
<td>6</td>
<td>Sodium Nitrate</td>
<td>0</td>
<td>gL⁻¹</td>
<td>8</td>
</tr>
<tr>
<td>7</td>
<td>Ammonium Sulfate</td>
<td>1.25</td>
<td>gL⁻¹</td>
<td>5</td>
</tr>
<tr>
<td>8</td>
<td>Ammonium Sulfate</td>
<td>2.5</td>
<td>gL⁻¹</td>
<td>5</td>
</tr>
<tr>
<td>9</td>
<td>Ammonium Sulfate</td>
<td>5</td>
<td>gL⁻¹</td>
<td>5</td>
</tr>
</tbody>
</table>

Estimation of biomass growth:
The optical density was determined with a Multiscan Go spectrophotometer (Thermo SCIENTIFIC, England) by measuring the absorbance of the medium at a provided wavelength. To measure the biomass and phycobiliproteins production, the cell dry weight of S. maxima was calculated by the equation of Y (gL⁻¹) = 0.58X-0.0201 (correlation coefficient (r) = 0.997), where Y is the cell dry weight of biomass and X is the optical density at 674 nm. We took the samples of cultures to analyze growth and phycobiliproteins contents at specific time intervals. The pigment concentrations of phycocyanin (PC) and allophycocyanin (APC) were calculated using the following equations by Bennett and Bogorad (Bennett and Bogorad, 1973).
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\[
PC (\text{gL}^{-1}) = \frac{(A_{620} - 0.474 A_{652})}{5.34} \quad (1)
\]

\[
APC (\text{gL}^{-1}) = \frac{(A_{652} - 0.208 A_{620})}{5.09} \quad (2)
\]

Where \( A_{620} \) and \( A_{652} \) are absorbance at the wavelength of 620 and 652, respectively.

Furthermore, the maximum specific growth rate (\( \mu_{\text{max}} \)) and the minimum doubling time (\( t_d \)) were determined by nonlinear regression of the logarithmic growth phase by plotting cell dry weight versus time using Wolfram Mathematica software version 11.3. Moreover, the nitrogen cell conversion factor of \( S. \) maxima in different treatments was calculated using the procedure of Danesi et al. (2011).

\[
Y_{X/N} = \frac{(X_m - X_i) V}{N_t} \quad (3)
\]

Where:

- \( Y_{X/N} \): Nitrogen-cell conversion (\text{gg}^{-1})
- \( X_m \): Maximum dry weight (\text{gL}^{-1})
- \( X_i \): Initial cell dry weight (\text{gL}^{-1})
- \( V \): The total volume of cultivation (L)
- \( N_t \): The total quantity of added nitrogen (g)

All graphic designs and pigment calculations in this study were performed using the GraphPad Prism 8 software.

**Phycobiliproteins Isolation:**

The cell walls were ruptured using repeated five freezing and thawing cycles for phycobiliproteins isolation (Sarada et al., 1999). Also, the samples were frozen at -20°C for 1 h and then were thawed at room temperature for 45 min. After freezing and thawing the samples up to five cycles, they were centrifuged (The Velocity 14/14 R Refrigerated centrifuge, China) at 10,000 rpm for 10 min at 4°C, where the supernatant was collected for phycobiliproteins determination.

**Results**

The impact of two different nitrogen sources was studied by replacing the basic-nitrogen source of Zarrouk’s medium (2.5 gL\(^{-1}\)) with different concentration of sodium nitrate and ammonium sulfate. Fig. 1 shows the changes in biomass growth by cell dry weight (panel A and B) and phycobiliproteins pigment (panel C and D) during the 8 days of cultivation time at different sodium nitrate concentration. Changes of cell dry weight (i.e., 1.209, 1.200, 1.010, 1.001, 0.992) gL\(^{-1}\) were monitored by increasing the concentration of sodium nitrate from 0 to 10 gL\(^{-1}\) (Fig. 1, panel B) at the final day of cultivation. Also, it was observed that the highest biomass growth (1.209 gL\(^{-1}\)) occurred in the nitrate-free medium. The cell dry weight was decreased with an increase in the amount of nitrogen in the culture medium. Although cell dry weight did not show significant changes related to concentration, even this smallest possible change is very important to reduce the production cost of scaling up. The highest phycocyanin and allophycocyanin production (0.066 and 0.088 mg/L) also occurred in the medium with sodium nitrate starvation (Fig. 1, panels C and D).
Treatment 5 showed a negligible change compared to treatments 3 and 4. Fig. 2 represents the effect of sulfate ammonium at the different concentration on biomass growth by cell dry weight (panels A and B) and phycobiliproteins pigment (panels C and D) during the cultivation time. In this case, the experiment was stopped due to a decrease in microalgae growth on day 5 when the medium culture was supplemented with ammonium sulfate. Although increasing the concentration of sodium nitrate do not dramatically change the accumulation of phycobiliprotein.

Allophycocyanin content was about 0.072, 0.044, 0.035, and 0.026 mgL\(^{-1}\) when cultured in the growth medium treatment 6, treatment 7, treatment 8, and treatment 9, respectively (Fig. 2, panels C and D).

Moreover, the highest and lowest phycocyanin reductions were observed about 79% and 44% after 5 days of cultivation in culture medium treatments 9 and 7, respectively.

Figure 1: The effect of sodium nitrate concentrations on cellular growth (panel A), cell dry weight (panel B), phycocyanin (panel C) and allophycocyanin content (panel D) during cultivation time. (∗: 0 mM, ⊥: 1.25 gL\(^{-1}\)mM, ◊: 30 mM, ▼: 60 mM, ◆: 120 mM). Values are the mean ± SD of three replicates.
Figure 2: The effect of sulfate ammonium concentration on cellular growth (panel A), cell dry weight (panel B), phycocyanin (panel C) and allophycocyanin content (panel D) during cultivation time (*: 0 gL⁻¹, ◊: 1.25 gL⁻¹, ○: 2.5 gL⁻¹, ▼: 5 gL⁻¹). Values are the mean ± SD of three replicates.

On the basis of the obtained results, it is clear that the changes in biomass growth could influence phycobiliprotein content. Fig. 3 exhibits the effect of sulfate ammonium at different concentrations by changing in medium color at several time-point during cultivation time. The color of matured S. maxima in treatment 6 altered from light to dark green (flask with nitrate-free), while it could not grow in other treatments of this group.

There was a significant difference among the cultures containing ammonium sulfate and without nitrogen source in terms of concentration of biomass and phycobiliprotein.

The dead cell of S. maxima was recognized in treatment 9 when the color of the culture medium was suddenly changed from green light to pale yellow and decolorized cell on the fifth day of cultivation.
Figure 3: Changes in medium’s color during cultivation day (from right to left show four different concentrations of ammonium sulfate of 0, 1.25, 2.5 and 5 gL⁻¹, respectively)

The changes in final cell dry weight and nitrogen-cell conversion factors related to nitrogen sources at different concentrations are presented in Table 2. Variations in the ammonium sulfate concentrations did not show a significant difference in a pattern throughout the growth and the cell dry weight values. The final cell dry weight of treatment 9 was the lowest when ammonium was only twice higher than the control concentration (2.5 gL⁻¹) of the total nitrogen. According to Table 2, the medium without nitrate source seemed more adequate than the others for growth. The nitrogen-cell conversion factor was decreased and cell dry weight decreased, suggesting the negative impact of ammonium sulfate when added to the medium.

The maximum specific growth rate ($\mu_{\text{max}}$), doubling time ($t_d$), and correlation coefficient ($r$) were studied as a function of nitrogen source according to Table 3.

Table 2: Final cell dry weight and nitrogen-cell conversion factor of S. maxima in different treatments.

<table>
<thead>
<tr>
<th>Treatment No.</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
</tr>
</thead>
<tbody>
<tr>
<td>Final cell dry weight (gL⁻¹)</td>
<td>1.209</td>
<td>1.200</td>
<td>1.010</td>
<td>1.001</td>
<td>0.992</td>
<td>0.835</td>
<td>0.143</td>
<td>0.088</td>
<td>0.065</td>
</tr>
<tr>
<td>Nitrogen cell conversion factor (gg⁻¹)</td>
<td>ND</td>
<td>0.776</td>
<td>0.306</td>
<td>0.151</td>
<td>0.074</td>
<td>ND</td>
<td>N</td>
<td>N</td>
<td>N</td>
</tr>
</tbody>
</table>

ND: Nitrogen source was not added in the medium
N: Negative amount

S. maxima samples were grown successfully in 5 out of 9 treatments. According to Table 3, the highest maximum specific rate (0.287) was found at 0 gL⁻¹ sodium nitrate at a smaller doubling time (2.43). Nevertheless, there were no statistical differences in this group. We recognized that ammonium utilization causes the pH to drop below the normal
range of microalga growth to sustain further growth (pH ≤ 5) in treatment 9. Moreover, pH instability in the presence of ammonium in the medium culture can prevent microalga growth. As a result, the ammonium source shows the lowest performance in biomass growth and phycobiliproteins production.

Table 3: Maximum specific growth rate ($\mu_{\text{max}}$), doubling time ($t_d$) and correlation coefficient ($r$) at different treatment.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>$\mu_{\text{max}}$ (day$^{-1}$)</th>
<th>$t_d$ (day)</th>
<th>$r$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.287</td>
<td>2.43</td>
<td>0.9943</td>
</tr>
<tr>
<td>2</td>
<td>0.260</td>
<td>2.67</td>
<td>0.9968</td>
</tr>
<tr>
<td>3</td>
<td>0.259</td>
<td>2.68</td>
<td>0.9980</td>
</tr>
<tr>
<td>4</td>
<td>0.256</td>
<td>2.73</td>
<td>0.9955</td>
</tr>
<tr>
<td>5</td>
<td>0.255</td>
<td>2.74</td>
<td>0.9968</td>
</tr>
<tr>
<td>6</td>
<td>0.248</td>
<td>2.80</td>
<td>0.9965</td>
</tr>
<tr>
<td>7</td>
<td>N</td>
<td>N</td>
<td>0.9907</td>
</tr>
<tr>
<td>8</td>
<td>N</td>
<td>N</td>
<td>0.9848</td>
</tr>
<tr>
<td>9</td>
<td>N</td>
<td>N</td>
<td>0.9893</td>
</tr>
</tbody>
</table>

N: negative amount

Discussion

Nitrogen source is an important issue since $S.\ maxima$ response can vary with different sources of nitrogen (Bezerra et al., 2008; Ferreira et al., 2010; Avila-Leon et al., 2012; Matsudo et al., 2012). In this regard, investigating the effect of nitrogen concentration could help to understand the effect of nutrient factor on production of microalga. Microalgae can utilize inorganic nitrogen in any of the three available forms like nitrate, nitrite, and ammonium by reducing the oxidized nitrogen (Cai et al., 2013). In the present study, ammonium sulfate and sodium nitrate were used as supplemental nitrogen in the medium. Cost et al. (2001) reported that the highest biomass was obtained in sodium nitrate, ammonium nitrate, and urea, in the order of their appearance (Cost et al., 2001). As a result, it was determined that the sodium nitrate at different concentrations introduced into the culture medium had no apparent positive effect on phycocyanin and allophycocyanin content. They also reported that the optimal cell dry weight was 1.992 gL$^{-1}$ in the growth medium containing 0.03 M sodium nitrate for production of $S.\ platensis$ during 672 hours (28 days) cultivation (Cost et al., 2001). We have determined that the cell dry weight decreased by increasing the concentration of sodium nitrite from 0 to 10 gL$^{-1}$, although the sodium nitrate at different concentrations introduced into the culture medium had no apparent positive effect. On the other hand, it was reported that the concentration of sodium nitrate in Zarrouk medium (2.5 gl$^{-1}$) reduces without losing the productivity as an important cost-saving factor in large-scale mass production of $S.\ platensis$ (Colla et al., 2007). Furthermore, de Castro et al. (2015) proved that sodium nitrate concentration in the Zarrouk’s medium can be reduced by increasing
the biomass growth while a higher biomass production depends on the bicarbonate sodium amount (de Castro et al., 2015). According to Tedesco and Duerr (1989), total lipid content was increased with nitrogen starvation in the Zarrouk’s medium, but the content of fatty acid was decreased in *S. platensis*. Moreover, they reported an increase in biomass growth during the increasing time of nitrogen starvation (Tedesco’ and Duerr, 1989). Similarly, in this study, the cell dry weight and pigment content were increased when nitrogen was limited. Furthermore, Abd El-Baky (2003) found that nitrogen concentration reduction in *S. platensis* culture decreases phycocyanin, chlorophyll and protein contents due to break-downing the chloroplasts which was not in accordance with our findings (El-Baky et al., 2003). However, nitrogen starvation increased the lipid fraction of some microalgae such as *S. platensis* (Illman et al., 2000; Uslu et al., 2011; Ak et al., 2015). Moreover, a research showed that the highest biomass content of *Spirulina platensis* without nitrogen source during 31 days of cultivation may be obtained while in contrast to the present research, nitrogen content had no significant effect on the biomass production enhancement (Uslu et al., 2011). On the basis of *Spirulina platensis*, a sharp decrease on biomass and phycocyanin production was reported when the concentration of sodium nitrate increased to 4.5 gL$^{-1}$. However, the maximum biomass concentration and phycocyanin productivity were observed in the batch medium containing 3.5 gL$^{-1}$ than the classic amount of Zarrouk’s medium in case of *Spirulina platensis* (Kaewdam et al., 2019).

Similar to our result, Cost et al. (2001) reported that it is impossible to utilize ammonium sulfate as an alternative nitrogen source instead of the classic nitrate-based of Zarrouk’s medium for the growth of *S. platensis*. The results showed that biomass growth was decreased by increasing the concentration of ammonium sulfate (Cost et al., 2001). Based on sulfate ammonium, it is suggested that the fed-batch supply of ammonium sulfate under pH control and varying the feeding time (time = 7–15 days) on a modified Schlösser’s medium leads to the best results in terms of biomass growth, protein, lipid, and carbohydrate content. Moreover, day 13 was selected as the best condition for adding ammonium sulfate on medium (Rodrigues et al., 2011).

Moreover, the ammonium sulfate rapidly dissolves into the ammonium and sulfate forms after addition to the culture medium (International Plant Nutrition Institute (IPNI)). Moreover, we also showed that *S. maxima* growth in the culture medium is associated with color variation. Also, the ammonium may be susceptible to gaseous loss in alkaline conditions. Then, ammonium component was converted in to the nitrite by nitrification process ($2\text{NH}_4^+ + 3\text{O}_2 \rightarrow 2\text{NO}_3^- + 2\text{H}_2\text{O} + 4\text{H}^+$) during
the cultivation period (Arsalan Sepehri and Sarrafzadeh, 2019). Although ammonia was converted into the nitrite in the presence of oxygen during the first day of cultivation, converting the produced nitrites into the nitrate form needs more oxygen (Barth et al., 2020). Hence, the ammonium sulfate typically has a pH in range of 5 to 6, which is lower than the pH balance of the microalga growth. Furthermore, the pH of culture medium was ultimately decreased by volatilization of acidity (H⁺). However, this level is not the desired working pH and may lead to death if the microalga is not adequately alkaline (in the range of 8 to 10). The experimental results of this study showed the ability of microalgae death to utilize ammonium and pH drop (≅ 5.3) in medium culture.

Based on the other study, the supply of ammonium as a nitrogen source in the Zarrouk’s medium for cultivating *Spirulina platensis* was shown to be effective either to produce biomass or to increase its CPC content. Notwithstanding, the microalgae utilize ammonium on excess CO₂ by aeration during the biomass growth to control growth pH at 9.0-9.5 (Kaewdam et al., 2019). Moreover, we also showed that *S. maxima* growth in the culture medium related to the observation of changing color from dark green to white during the first five days of cultivation. Besides this, ammonium sulfate did not allow *S. maxima* to settle down and grow, as well as preventing sodium nitrate as a nitrogen source. Therefore, a negative specific growth rate (**μ**ₘₐₙₙ) and doubling time (**t**ₐ) were observed when ammonium sulfate was available in the medium. In this process, using excess bicarbonate with ammonium may be useful while using ammonium can be the principal cause of the reduction in alkalinity, which impacts pH stability and then biomass growth capacity. Another result of this study was that phycocyanin and allophycocyanin contents of *S. maxima* were rapidly declined by increasing the ammonium sulfate concentration in the medium.

Among the nutrients required for the growth of *S. maxima*, nitrogen is not considered as an important element for the synthesis of phycobiliprotein according to our findings, while it is almost necessary for the synthesis of proteins and other cellular components (Khazi et al., 2018). It has been also reported that nitrogen is essential factor for synthesis of the amino acids and the excess nitrogen can cause an increase in the protein content of Spirulina. The same finding showed that lipid content could remained constant at all concentration of nitrogen (Piorreck et al., 1984). With regard to our study, high concentration of nitrate approximately showed no increase of phycobiliprotein content in Spirulina.

Briefly, the use of ammonium sulfate as an alternative source instead of the classic-nitrate source of Zarrouk’s medium was not an appropriate strategy for increasing the biomass growth and pigment content of
S. maxima. Although the addition of sodium nitrate introduced into the culture medium in high concentration had no positive effect on the biomass growth and pigments production, the absence of a nitrogen source could accelerate the biomass growth and promote pigments production under the conditions tested in this study. Moreover, the biomass growth would be limited by pH drop toward acidity in the medium. Besides, using ammonium sources was also found to be ‘toxic’ to algae growth. Another result is that sodium nitrate concentration was not sensitive to biomass growth changes of S. maxima. According to the experimental results of this study, either sodium nitrate or ammonium sulfate does not affect the final cell dry weight and phycobiliproteins content. This study highlights the advantages of nitrogen starvation both in terms of lower costs and higher biomass growth, which can be used for large-scale production in the real environment.

Acknowledgments
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