Research Article

Expression differences of interferon regulatory factor 3 and non-specific cytotoxic cell receptor protein-1 in grass carp (Ctenopharyngodon idella) after challenges with two genotypes of grass carp reovirus, and analysis of antiviral signaling pathways

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Abstract
Grass carp reovirus (GCRV), with high genetic diversity, seriously affects grass carp farming. Meanwhile, interferon regulatory factor 3 (IRF3) and non-specific cytotoxic cell receptor protein-1 (NCCRP-1) play crucial roles in immunity system of fish. In this paper, we took IRF3 and NCCRP-1 as molecular markers in order to explore the differences in immune responses of grass carp infected with different GCRV strains, which is helpful in better understanding of the GCRV pathogenesis. Accordingly, we analyzed the expression differences of IRF3 and NCCRP-1 in grass carp after challenges with GCRV GD108 (genotype III) and GCRV 096 (genotype I) by qRT-PCR. We found IRF3 and NCCRP-1 was up-regulated in all tested tissues after challenges of GCRV GD108 and GCRV 096. However, the expression levels of IRF3 and NCCRP-1 depended on the GCRV strain, and the up-regulation changes were qualitatively similar but quantitatively different after challenges of GCRV GD108 and GCRV 096, that is, NCCRP-1 was up-regulated more, earlier, and more broadly than IRF3 after GCRV challenge. Besides, the NCCRP-1 was more rapidly up-regulated to a greater extent after challenges with GCRV 096 than GCRV GD108. In order to clarify whether the antiviral immune response and development of grass carp hemorrhage in grass carp differed depending on GCRV genotypes needs more investigation. Moreover, we speculated that the GCRV-induced signaling pathways in grass carp were enriched with measles- and influenza A-induced signaling pathways, and found 13 types of new genes in the GCRV-induced signaling pathway.

Keywords: Interferon regulatory factor 3 (IRF3), Non-specific cytotoxic cell receptor protein-1 (NCCRP-1), Grass carp (Ctenopharyngodon idella), Grass carp reovirus (GCRV)

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Introduction

GCRV causes grass carp hemorrhage and has seriously affected the grass carp farming industry (Zhang et al., 2010; Wang et al., 2012; Yan et al., 2014). Diseased grass carp had different hemorrhagic symptoms, “red fins and opercula, red muscles, or red intestines” (Chen et al., 2010). However, recent epidemiology (from 2012.04 to 2015.06) of grass carp hemorrhage indicated that all the detected sick grass carp showed the symptom of “red intestines”, much less symptoms of “red muscles” or “red fin and opercula” (Huang et al., 2016). In addition, most of the detected GCRV isolates were attributed to GCRV genotype III (Huang et al., 2016).

GCRVs and antiviral immune responses of grass carp have been studied extensively (Chen et al., 2010; Zhang et al., 2010; Wang et al., 2012; Yan et al., 2014; Xu et al., 2015; Ou et al., 2017). However, the pathogenesis of GCRVs is not completely understood, and the prevention and treatment of grass carp hemorrhage are still difficult, in part owing to the high genetic diversity of GCRVs. Three of four GCRV genotypes are found in China, where GCRV strains of genotype I and genotype III are common. Substantial differences among GCRV genotypes have been reported (Wang et al., 2012; Yan et al., 2014). Thus, we hypothesized whether different GCRV genotypes are related to the antiviral immune responses of grass carp. Moreover, studies on the differences in the immune response of grass carp after challenges with GCRVs are necessary for development of prevention and treatment strategies for grass carp hemorrhage, and pathogenesis of GCRV.

The innate immune response of the host is the first line of defense against viral infection by involving in the subsequent activation of the adaptive immune response (Feng et al., 2011). Among the major immune mechanisms against viral infections in fish, the interferon (IFN) pathway and nonspecific cytotoxic cell (NCC) activity are critical. IRF3 and NCCRP-1 respectively play crucial roles in the IFN (interferon) pathway and NCC activity (Jaso-Friedmann et al., 2001; Kallio et al., 2011; Tremblay et al., 2016).

In the induction of type I IFNs following virus infection, IRF3-mediated IFN-β production and the antiviral innate immune response are regulated by the IRF3 nuclear translocation, activation, polyubiquitination, phosphorylation, and so on (Wang et al., 2015; Song et al., 2016; Tremblay et al., 2016; Wang et al., 2016; Bakshi et al., 2017). Viruses have evolved multiple strategies to abrogate or inhibit the activation of IRF3 to counteract IFN-mediated responses and successfully infect the host (Kuo et al., 2016; Zhang et al., 2016). Fish possess an IFN system similar to that of mammals (Sun et al., 2010; Avunje et al., 2011; Yao et al., 2012). For example, IRF3 also plays essential roles in controlling the IFN response during viral infection in fish. IRF3 is expressed broadly in various tissues in fish and is transcriptionally regulated by viral stimulation (Sun et al., 2010; Yao et al.,...
NCCRP-1 also functions in the NCC immune response in teleosts (Evans and Jaso-Friedmann, 1992; Jaso-Friedmann et al., 2001). NCCs play a number of important roles in immune responses by lysing tumor target cells, protozoan parasites, and virus-infected cells (Jaso-Friedmann et al., 2000). Moreover, NCCRP-1 is a type III protein receptor on NCCs, the only known receptor on NCCs, with an antigen-binding domain, signaling domain, a transcriptional activation domain, and F-box-associated (FBA) domain. NCCRP-1 may play dual roles in the activation of NCC, i.e., the recognition and binding to target cells and the activation of the JAK/STAT signaling pathway (Jaso-Friedmann et al., 2004; Ye et al., 2012).

Furthermore, GCRV GD108 belongs to GCRV genotype III and causes 60–80% mortality in grass carp fingerlings (about 10 cm in length) with “red intestines and without obvious cytopathic effects in their kidney cells (Tamura et al., 2007). GCRV 096 also belongs to GCRV genotype I, and causes 20–30% mortality in grass carp fingerlings with “red muscle”, and with obvious cytopathic effects in their kidney cells (Yan et al., 2014). Therefore, in the present study, we examined the expression differences in IRF3 and NCCRP-1 of grass carp after challenges with GCRV GD108 and GCRV 096 in order to verify our hypothesis. Moreover, we predicted the signaling pathways involved in grass carp to GCRV infection.

**Materials and methods**

**Virus**

GCRV 096 was isolated from grass carp with hemorrhagic disease in Hubei Province of China and GCRV GD108 was isolated from grass carp with hemorrhagic disease in Guangdong province of China. Both viruses were stored in our laboratory (Ye et al., 2012; Yan et al., 2014).

**Fish and viral infection**

All grass carp (200±15 g) were obtained from the Hedi reservoir in Guangdong Province. We examined these fishes for pathogens by PCR, RT-PCR, microscope, etc. and did not find any pathogens or disease symptoms in them. So, we assumed the grass carp were healthy. The fish were acclimated in an aerated fresh water tank at 26±0.5 °C under natural photoperiod and fed for 2 weeks to adjust to the laboratory conditions before GCRV infection. The water quality parameters DO, NH3, NO2 and pH were measured as 8.1 mg L⁻¹, 0.1 mg L⁻¹, 0.03 mg L⁻¹ and 7.5, respectively. The fishes in the infection group were intraperitoneally injected with 200 μl of GCRV GD108 or GCRV 096 suspension (approximately 2000 μg ml⁻¹) (Hiscott, 2007; Yan et al., 2014), and the same amount of 0.7% saline solution was administered to the fish of the control group.

**Sample collection**

After the experiment was set up, three grass carp from each of the infection groups and the control group were captured at days 0.5, 1, 2, 3, 4, 5, 6, and
7 after the infection induction, and tissues samples were collected from the kidney, spleen, liver, heart, brain, skin, gill, intestine, and muscle (3 specimens at each time in order to calculate the mean value of the measured parameters for each tissue). These samples were immediately stored at -80 °C until total RNA was extracted for dynamic expression analyses.

**Total RNA extraction**
All collected samples were homogenized in TRIzol reagent (Invitrogen, Carlsbad, CA, USA), and total RNAs were isolated according to the manufacturer’s instructions. The quality and concentration of total RNA were analyzed by agarose gel electrophoresis and ultraviolet spectrophotometry based on the absorbance at 260 and 280 nm. Total RNA was incubated with RNase-free DNase I (Fermentas), followed by reverse transcription into cDNA, using oligo (dT)20 primers with MMLV reverse transcriptase (Toyobo, Osaka, Japan) following the instructions provided.

**Cloning and sequencing of IRF3 and NCCRP-1 in grass carp**
Degenerate primers were designed based on IRF3 or NCCRP-1 sequences in fish to amplify the 3’ and 5’ flanking sequences using the SMART RACE cDNA Amplification Kit (Takara, Shiga, Japan) according to the manufacturer’s instructions. These degenerate primers were used to perform polymerase chain reaction (PCR), and the PCR products were purified on 0.8% agarose gels using the DNA Gel Extraction Kit (Axygen, Union City, CA, USA) following the manufacturer’s instructions. Purified DNA fragments were ligated into pMD-18T vectors (Takara), transformed into competent *Escherichia coli* DH5α cells, and incubated at 37 °C on LB agar plates. Positive colonies were screened by colony PCR using M13 universal primers and sent to Sangon Biotech (Shanghai, China) for sequencing. To acquire the full-length cDNA sequences of IRF3 and NCCRP-1 in grass carp, rapid amplification of cDNA ends (RACE) was performed using the BD SMART RACE cDNA Amplification Kit (Clontech, Mountain View, CA, USA) according to the manufacturer’s instruction.

**Sequence analysis of IRF3 and NCCRP-1 and phylogenetic analyses**
The nucleotide and deduced amino acid sequences of IRF3 and NCCRP-1 from grass carps were analyzed using BioEdit 7.0.1 and Expasy (www.expasy.org/tools). The sequences of IRF3 and NCCRP-1 from different species were aligned using DNAstar with ClustalV, and final adjustments were performed manually. The transmembrane domains were predicted with the DAS-TMfilter (http://mendel.imp.ac.at/sat/DAS/DAS.html), PRED-TMR (http://athina.biol.uoa.gr/PRED-TMR/ inp ut.html), MINNOU (http://minnou.cchmc.org/), and TMMOD (http://liao.cis.udel.edu/website/servers/TM OD/scripts/frame.php?p=submit). Signal peptides were predicted using SignalP (http://www.cbs.dtu.dk/
services/SignalP/). Domains were predicted using the SMART server (http://smart.embl-heidelberg.de/).

Neighbor-joining (NJ), maximum parsimony (MP), and minimum evolution trees were constructed using MEGA 4.1, with *Homo sapiens* as the outgroup and bootstrap analysis (Livak and Schmittgen, 2001).

**Determination of the tissue-specific expression of IRF3 and NCCRP-1 by qRT-PCR**

Real-time quantitative PCR was performed using the Rotor Gene 3000 with SYBR Green Master Mix (Toyobo) to quantify the expression levels of *IRF3* and *NCCRP-1*. Each pair of gene-specific primers (Table 1) for RT-qPCR was designed to amplify 200 bp. *GAPDH* and β-actin was used as reference genes. The final volume reactions were 20 μl, including 1 μl of 2× SYBR Green Master Mix, and 250 nM each primer. Cycling parameters were as follows: 95 °C for 5 min, followed by 40 cycles of 95 °C for 20 s, 55 °C for 20 s, and 72 °C for 30 s. Three parallel assays were performed for each sample to assure the specificity of the amplification products.

### Table 1: Oligonucleotide primers used for PCR.

<table>
<thead>
<tr>
<th>Primer name</th>
<th>Sequence (5′-3′)</th>
<th>Usage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Oligo(dT)₃₀-anchor</td>
<td>AAGCAGTGTGATCAACGCAGAGTACT(30)VN</td>
<td>3′ RACE-PCR</td>
</tr>
<tr>
<td>NCC F317</td>
<td>TGCTCTAGTACAGGGTGTGTTCTCT</td>
<td>5′ RACE-PCR</td>
</tr>
<tr>
<td>NCC R1</td>
<td>GACGGTGATCATCGACAGTCC</td>
<td>PCR</td>
</tr>
<tr>
<td>NCC R2</td>
<td>CCAGGTGTGTGAGTAGTTCTCC</td>
<td>5′ RACE-PCR</td>
</tr>
<tr>
<td>NCC R3</td>
<td>CACACTGGATTTGCTGGTCTTG</td>
<td></td>
</tr>
<tr>
<td>Oligo(dT)-anchor</td>
<td>GACCGACGCGTA TCGATGTCGACT(16) V</td>
<td>Identification</td>
</tr>
<tr>
<td>Anchor primer</td>
<td>GACCGACGCGTA TCGATGTCGACT</td>
<td></td>
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<tr>
<td>RV</td>
<td>GACCGGATAAACAAATTCACACAGGA</td>
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<tr>
<td>M13</td>
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<tr>
<td>IRF3-31</td>
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<tr>
<td>IRF3-32</td>
<td>CGAGTCTTTCCACTTGGTCAC</td>
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<tr>
<td>R1 IRF3a</td>
<td>CCAGTGAAGATGTCCTCACA</td>
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<tr>
<td>IRF3-R1b</td>
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<td>IRF3-R1c</td>
<td>TACAGACATCGTCTTGGTTC</td>
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<td>5′PCR anchor</td>
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<td>β-actinF</td>
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<td>GAPDH</td>
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<td>NCCRP-F</td>
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<td>NCCRP-R</td>
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<tr>
<td>IRF3-R</td>
<td>TTAGCTAGACTTTGTAAGGACT</td>
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**Statistical analysis**

Relative *IRF3* and *NCCRP-1* expression levels were normalized against *GAPDH* and β-actin expression using the 2⁻ΔΔCt method (Kuleshov et al., 2016). Statistical analyses of *NCCRP-1* and *IRF3* expression levels were performed using MS Excel 2003 (Microsoft Corporation, Redmond, WA, USA). Groups were compared using unpaired t-tests.
Antiviral signaling pathways in grass carp

Based on the 160 antiviral immune-related genes of grass carp identified in GenBank (Supporting information), using the KEGG (Kyto Encyclopedia of Gene and Genomes) database, Wiki (Wikipedia) Pathways database, Reactome database, BioCarta database, HumanCyc database, NCI-Nature database, Panther database, and the Enrichr tool (Evans et al., 1999), signaling pathways involved in the immune response of grass carp against GCRV were predicted. The 160 antiviral immune-related genes were listed in the supplementary information.

Results

Characteristics of IRF3

The cDNA of IRF3 was 1831 bp (GenBank accession number: KC898261), with a 192-bp 5' UTR (Untranslated regions), a 1377-bp open reading frame (ORF), and a 262-bp 3' UTR. The predicted gene product was 459 amino acids. The sequence we acquired was identical to the sequence reported by Xu (Xu et al., 2015). A phylogenetic tree based on IRF3 (Fig. 1) showed that grass carp IRF3 is highly similar to Carassius auratus IRF3.

Characteristics of NCCR-P-1

The full-length cDNA of NCCR-P-1 was 900 bp (GenBank accession number: HQ388293), with a 3-bp 5' UTR, a 714-bp ORF, and a 183-bp 3' UTR. The predicted gene product was 237 amino acids, with a molecular weight of 27.3 kDa. Based on the analyses using SignalP and the TMHMM server, NCCR-P-1 in grass carp had no signal peptide or transmembrane region. NCCR-P-1 contained Box-1 (PxPxxP), Box-2 (PxP), an FBA domain, and a putative antigen-recognition site (LPxxxWFxxEQxVxLKA) (Jaso-Friedmann et al., 2001; Huang et al., 2014). A phylogenetic tree based on NCCR-P-1 (Fig. 2) showed that grass carp is closely related to Cyprinus carpio.
Dynamic expression of IRF3 and NCCR-P-1 in grass carp after GCRV challenge

Dynamic expression of IRF3 and NCCR-P-1 in the kidney, spleen, liver, heart, brain, skin, gill, intestine, and muscle of grass carp during a 7-day period after infection with GCRV GD108 and GCRV 096 was detected by qRT-PCR. IRF3 was clearly expressed in all tissue types and its expression level differed among tissues (Fig. 3). For IRF3 in grass carp after challenges with GD108 and GCRV 096, the most pronounced up-regulation was detected in the kidney and spleen (~10–15-fold greater expression than that of the control group), intermediate up-regulation was observed in the liver, skin, gills, and intestine (~5–8-fold increase compared with the control group), and low up-regulation was observed in the heart, brain, and muscle. Dynamic expression of IRF3 was similar after the challenge with GCRV GD108 and GCRV 096. For NCCR-P-1 in grass carp after challenges with GD108 and GCRV 096, the most pronounced up-regulation was detected in the kidney, skin, gills, intestine, and muscle (~10–80-fold greater expression than that of the control group), intermediate up-regulation was observed in the liver, spleen, and heart (~5–9-fold increase compared with the control group), and low up-regulation was observed in the brain. Especially, expression of NCCR-P-1 in the liver, heart, and intestine was higher and earlier after the challenge of GCRV 096 than GCRV GD108. The results suggested antiviral immune responses of grass carp to GCRV 096 and GCRV GD108 were different to an extent.
Yan et al., Expression differences of interferon regulatory factor 3 and NCCRP-1 mRNA expression

A Kidney

B Spleen

C Liver

Days post-infection

Relative folds of IRF3 and NCCRP-1 mRNA expression

Days post-infection

Relative folds of IRF3 and NCCRP-1 mRNA expression

Days post-infection

Relative folds of IRF3 and NCCRP-1 mRNA expression
Relative folds of IRF3 and NCCRP-1 mRNA expression

D. Heart

E. Brain

F. Skin
Figure 3: Expression of *IRF3* and *NCCR-P-1* in grass carp with the challenge of GCRV GD108 and GCRV 096.
Pathway-enrichment analysis

Based on 160 identified antiviral immune-related genes in grass carp, using the KEGG database, Wiki Pathways database, Reactome database, BioCarta database, HumanCyc database, NCI-Nature database, Panther database, and the Enrichr tool, we predicted that the most enriched pathways in virus-infected grass carp were the toll-like receptor signaling pathway, RIG-I (retinoic acid-inducible gene-I)-like receptor signaling pathway, NOD (nucleotide-binding oligomerization domain)-like receptor signaling pathway, measles-induced pathway, and influenza A-induced pathway (Fig. 4).

Using the KEGG database, we determined the potential virus-induced signaling pathways in grass carp (Fig. 4). In addition, homologs of TRAF (tumor necrosis factor receptor-associated factor)3, TANK (TRAF family member associated nuclear factor κB activator), NAP1 (nucleosome assembly protein 1), ERK (extracellular signal-regulated kinase), MKK (Mitogen-activated protein kinase kinase), IP-10 (IFN-γ-inducible protein 10), MIG (monokine induced by IFN-γ), RANTES (chemokine ligand 5), MIP-1 (macrophage infectivity protein 1), FADD, MEKK1 (mitogen-activated protein kinase kinase kinase 1), IP-10 (IFN-γ-inducible protein 10), MIG (monokine induced by IFN-γ), RANTES (chemokine ligand 5), MIP-1 (macrophage infectivity protein 1), FADD, MEKK1 (mitogen-activated protein kinase kinase kinase 1), IRAK (interleukin-1 receptor-associated kinase) 4, and RIP (receptor-interacting protein) 2 should be involved in the immune response in grass carp (Fig. 4), according to KEGG database pathways and the reported immune response relative genes in other fishes (GenBank accession numbers of them in other fishes are respectively KJ789921, KP861886, NM_201318, NM_001124424, NC_031739, AJ417078, NC_024318, NC_019876, BT074182, NC_030425, NC_007121, FN598578 and NC_007113).
Discussion
Despite many studies of GCRV and immune responses in grass carp, the mechanisms of grass carp against GCRV are still unclear, and prevention and treatment of grass carp hemorrhage is still a difficult problem. In the present study, we sequenced IRF3 and NCCRP-1 of grass carp, sharing conserved motifs with the IRF3 subfamily and the FBA subfamily (Jaso-Friedmann et al., 2001; Xu et al., 2015). Additionally, NCCRP-1 in grass carp was a soluble protein, similar to NCCRP-1 in the orange-spotted grouper (Panne et al., 2007), rather than a type III membrane protein (Jaso-Friedmann et al., 2001).

In this study, the dynamic expression of IRF3 and NCCRP-1 were qualitatively similar, but quantitatively different after challenges with GCRV GD108 and GCRV 096. Both genes were up-regulated in all tested tissue types, especially the kidney. The kidney might be an important organ for the grass carp defense against GCRV infection.

Our results suggested that IRF3 in grass carp participates in the immune response against GCRV infection and might play an important role in defense against viral infection. After viral infection, fish IRF3 is activated and regulated by phosphorylation, ubiquitination, dimerization, nuclear translocation, etc. Once activated, fish
IRF3 mediates the induction of type I interferons (Kumar et al., 2000; Bergstroem et al., 2010; Sun et al., 2010; Briolat et al., 2014; Gu et al., 2016; Zhao et al., 2016). Moreover, the overexpression of fish IRF3 affects components of the antiviral response in fish (Hiscott, 2007; Bergstroem et al., 2010; Sun et al., 2010; Zhu et al., 2015). IRF3 is expressed in all tested tissues and is up-regulated in response to poly I:C stimulation in various fishes, such as the large yellow croaker, grass carp, miuuy croaker, and tilapia (Yao et al., 2012; Xu et al., 2015; Gu et al., 2016).

In this study, IRF3 in most of the tested tissues in grass carp was up-regulated after challenges with GCRVs, similar to the results obtained in fishes after stimulation with viruses and poly I:C (Yao et al., 2012; Xu et al., 2015; Gu et al., 2016). NCCR-1 expression was also up-regulated in various tissues, indicating that NCCs in grass carp are involved in the immune response to GCRV challenge. Expression of NCCR-1 is up-regulated following the activation of NCCs by stimuli, resulting in higher killing activity (Chaves-Pozo et al., 2012; Huang et al., 2014). The increased NCCR-1 expression has been linked to the augmentation of killing mechanisms by NCCs (Huang et al., 2014). NCCs kill virally transformed cells without prior activation. They collaborate with other effector cells of the immune system, acting as an important component of innate immunity in fish, and they provide innate resistance against virus challenges. In addition, NCCR-1 is widely expressed in various organs and is responsible for the recognition of target cells to initiate the lytic cycle. NCCR-1 may have at least two functions, i.e., target cell binding and the activation of cytokine release (Jaso-Friedmann et al., 2001). However, few studies of fish NCC activity have focused on the roles of NCC and NCCR-1 in defense against viral infections. Our results were similar to those of Chaves-Pozo, who also showed that NCCR-1 is up-regulated in sea bream and sea bass infected with nodavirus (Vaziri et al., 2012).

Further, our results suggest that the responses of IRF3 and NCCR-1 expression in grass carp differed depending on the GCRV strain. NCCR-1 expression increased to a greater extent and more rapidly after challenges with GCRV 096 than GCRV GD108. Immune responses of hosts differ after challenges with different virus strains. For example, differences in immune response are influenced by the virulence and pathogenicity of virus strains (Eldaghayes et al., 2006; Adams et al., 2009; Ecco et al., 2011; Camp et al., 2013; Badouzi et al., 2014; Kumar et al., 2017; Shevtsova et al., 2017). The virus-induced host immune response plays an important role in the differences in viral pathogenesis among pathotypes of influenza virus (Cui et al., 2014). Little is known about the response of grass carp infected with GCRV and the relationship between the innate immune response and the severity of the clinical disease. Based on the findings of the present study, differences in the host immune response
might explain, in part, differences in pathogenicity between high and low pathogenic GCRV strains in grass carp.

The immune response to GCRV in the early stages of infection may be a critical determinant of disease course (Kumar et al., 2017). In addition, the expression of NCCR1 was up-regulated more, earlier, and more broadly than that of IRF3 after GCRV challenge. For IRF3, the earliest and highest expression was observed at 3 days (for GCRV 096, Fig. 3 (H)) or 4 d (for GCRV GD108, Fig. 3 (A, B, C, F)) after infection. IRF3 was highly up-regulated in the kidney, gill, and intestine (4–15-fold higher expression than that in the control group). For NCCR1, the earliest and highest expression in different tissues was detected 1 day (for GCRV 096, Fig. 3 (D)) or 0.5 days (for GCRV GD108, Fig. 3 (H)) after infection. NCCR1 was highly up-regulated in the kidney and intestine (40–80-fold higher expression than that in the control group) (Fig. 3). The dynamic expression pattern of IRF3 was obviously different from that of NCCR1 after the GCRV challenge (Fig. 3). However, the mechanism underlying this difference is not clear.

To explore the antiviral immune responses of grass carp and the development of grass carp hemorrhage depending on GCRV genotypes needs more researches.

We also found 13 types of new genes in grass carp that may play roles in the antiviral immune response (Fig. 4). Based on 160 known antiviral immune-related genes in grass carp and the KEGG database, we predicted that the virus-induced signaling pathways in grass carp was highly enriched with toll-like receptor signaling pathway, RIG-I-like receptor signaling pathway, and NOD-like receptor signaling pathway (Fig 4). According to the three enriched pathways in KEGG database and reported immune response related genes in other fishes, homologs of TRAF3, TANK, NAP1, ERK, MKK, IP-10, FADD (fas associated via death domain), MEKK1, MIG, RANTES, MIP, IRAK4, and RIP2 in grass carp were expected to be involved in the antiviral immune signaling pathway of grass carp (Fig 4).

Furthermore, the virus-induced signaling pathways in grass carp was highly enriched with the measles- and influenza A-induced signal pathways (Fig. S1). According to the 173 antiviral immune-related genes in grass carp and the KEGG database, we speculated that the toll-like receptor signaling pathway (Fig. 4), T-cell receptor signaling pathway, NF-κB and MAPK signaling pathway, B-cell receptor signaling pathway, antigen-processing and presentation, complement cascades (e.g. C2, C3, C5, C6), and chemokine signaling pathway should be involved in the immune response of grass carp against virus infection, similar to previous results (Li et al., 2017). And that, Measles-induced pathways include T-cell receptor signaling pathway, Toll-like receptor, B cell receptor and RIG-I-like receptor signaling pathway, which were involved in the virus-induced signaling pathways in grass carp. Influenza A-induced pathways include
Toll-like receptor, NOD-like receptor and RIG-I-like receptor signaling pathway, which were also involved in the virus-induced signaling pathways in grass carp. Therefore, this study may explain the enrichment of the virus-induced pathways in grass carp with measles- and influenza A-induced pathways. Maybe measles- and influenza A-induced pathways provide new thoughts for exploring GCRV’s pathogenic mechanism.

In conclusion, we found that the expression levels of IRF3 and NCCRP-1 were depended on the GCRV strain. More investigations on the expression differences of IRF3 and NCCRP-1 in grass carp infected with different GCRV strains may help to explain the mechanisms underlying grass carp hemorrhage and host immune responses. Bioinformatics speculation of signaling pathway suggested that GCRV-induced signaling pathway was enriched with measles- and influenza A-induced signaling pathways.

Acknowledgments
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References


Briolat, V., Jouneau, L., Carvalho, R., Palha, N., Langevin, C. and Herbomel, P., 2014. Contrasted innate responses to two viruses in
zebrafish: insights into the ancestral repertoire of vertebrate IFN-stimulated genes. *Journal of Immunology*, 192(9), 4328-4341.


**Gu, Y.F., Wei, Q., Tang, S.J., Chen, X.W. and Zhao, J.L., 2016.**
Molecular characterization and functional analysis of IRF3 in tilapia (Orochromis niloticus). Developmental and Comparative Immunology, 55, 130-137.


pattern underlying differential host response to high or low pathogenic H5N1 avian influenza virus in ducks. *Acta Virologica*, 61(1), 66-76.


