Serratia marcescens B4A chitinase thermostability enhancement by S390I QuikChange site directed mutagenesis

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Abstract
Thermostable chitinases are useful for industrial and biotechnological applications. This paper reports the stabilization of chitinase from Serratia marcescens B4A through rational mutagenesis. Changing of Ser 390 to Ile in S. marcescens. The stabilization was enhanced through entropic stabilization by reduction of the loop length and also by increasing of the beta chain length. With this replacement, polar uncharged residue changed to non-polar one and increased the hydrophobic interactions. Furthermore Isoleucine has branched β-carbon that restricts the backbone conformation more than non-branched residues. Finally all of these factors lead to entropic stabilization and thermal stabilization. The results exhibited that the optimal temperature and pH for enzyme activity of native chitinase were not changed by mutagenesis which showed that mutation didn’t affect the original characteristics of the enzyme, the K_m values of native and mutant chitinase were different very little, showing that the affinity of enzyme towards the substrate and also the natural flexibility of chitinase did not change by mutation. Besides the V_max value of the mutant chitinase was decreased, while its pH stability was increased briefly, but its thermal stability was increased remarkably. Mutation made chitinase to tolerate high temperatures up to 90°C. In addition its activity was increased at 50°C, 60°C for 120 min and up to 2 hours of incubation period and the mutant chitinase demonstrated a high level of activity at 60°C. These results show that entropic stabilization works well for chitinase and this approach may be generally applicable for stabilization of other proteins.

Keywords: Chitinase, Thermal stability, Serratia marcescens B4A, Loop, β chain, unfolding

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Introduction

Chitin, a polysaccharide made of β-(1,4) linked N-acetyl D-glucosamine units, is the second most plentiful biopolymer and a source of renewable raw materials (Patil et al., 2000; Annamalai et al., 2011; Wang et al., 2015). Chitin has applications in industrial, biotechnology, agriculture, material and medical, food and nutrition sciences (Xia et al., 2011). Chitinases (EC 3.2.1.14) are widespread enzymes in nature with ability to degrade chitin into smaller chito-oligosaccharide segments (Bhattacharya et al., 2007; Tsuji et al., 2010). These enzymes are efficient biomarkers in cancer disease (Wang et al., 2015) and are used for treatment of CF-associated fungal disease (Hector et al., 2016). Some chitinases such as chitinase 3-like 2 and chitotriosidase are used in early MS diagnosis (Elmonem et al., 2016; Møllgaard et al., 2016).

Thermostable chitinases are in demand in various biotechnology sectors (Bjørk et al., 2003; Aliabadi et al., 2016). The need, similar to the other interesting enzymes, has been addressed through protein engineering. However, the use of a rational design for protein stability improvement seems to be protein specific and any of such attempts would require to be directed by the detailed structural information (Giver et al., 1998; Gåseidnes et al., 2003; Khurana et al., 2011; Matroodi et al., 2013) and understanding the correlations between the sequence, function and the structure (Eijsink et al., 2004; Khurana et al., 2011).

Several reasons have been ascribed to the superior stability of the thermophilic proteins including greater hydrophobicity (Mozhaev and Martinek, 1984; Haney et al., 1997; Kumar et al., 2000) better packing, shorter loops (Russell et al., 1997), smaller and less numerous cavities (Salminen et al., 1996; Kumar et al., 2000), less uncharged polar residues (Haney et al., 1999; Fields, 2001), increased occurrence of proline residues (Bogin et al., 1998; Kumar et al., 2000), and increased hydrogen bonding (Vogt and Argos, 1997).

*Serratia marcescens* is one of the pioneer bacterium in chitin degradation (Watanabe et al., 1997; Bjørk et al., 2003). *S. marcescens* B4A chitinase (Babashpour et al., 2012) plays an important role in its plant antipathogenic properties. 3D-Structure analysis of this enzyme suggests that substitution of Serine 390 with Isoleucine could improve its thermostability by shortening the loop size and increasing the β-sheet length. To examine the assumption, Quik Change Site Directed Mutagenesis (QCSDM) was employed to modify *S. marcescens* B4A chitinase structure and its thermostability. The outcome of the research has been presented in this paper.
Materials and methods

Materials

The plasmid pQE60 chitinase (previous work) (Babashpour et al., 2012; Zarei et al., 2012) was used for amplification of chitinase gene. *Escherichia coli* strain DH5α (Invitrogen. Carlsbad, CA, USA) was used as the host for molecular cloning of recombinant pTZ57R/T (Fermentas. Waltham, Massachusetts, USA). pET26b (Fermentas. Waltham, Massachusetts, USA) plasmids harboring native and mutant chitinase gene, respectively. BL21 (DE3) *Escherichia coli* strain (Invitrogen. Carlsbad, CA, USA) was also used for protein expression. Flake crab shell chitin and 3, 5-dinitrosalicylic acid (DNS) were purchased from Sigma (St. Louis, MO, USA). Colloidal chitin was prepared according to the modified method of Roberts and Selitrennikoff and use of commercial chitin (Takiguchi, 1991).

In silico studies

Homology modeling and 3D-structure analysis

The amino acid sequence of chitinase retrieved from NCBI database was used as a template in BLASTP search against PDB to find appropriate templates for homology modeling. Amino acid sequence of *Serratia marcescens* B4A chitinase (accession number HM473183) was retrieved from the GenBank database (http://www.ncbi.nlm.nih.gov). BLASTP search (Altschul et al., 1997) of a query (chitinase sequence) was performed against Protein Data Bank (PDB). Homology modeling for the native and the mutant chitinase of *Serratia marcescens* B4A was performed by MODELLER v9.10 (http://www.salilab.org/modeller/) (Webb and Sali, 2014) using opened form of chitinase (1edq_A) downloaded from PDB server as the template. The MODELLER generated structure of the mutant chitinase was further analyzed by Ramachandran analysis generated by Procheck (Laskowski et al., 1993), Qmean server (Benkert et al., 2008), ProSA-web (Wiederstein and Sippl, 2007), and RMSD value calculation (Humphrey et al., 1996). Qmean is a parameter (between 0 and 1) for assessment of model credibility and ProSA shows quality score of the predicted model in the context of all known protein structures. Furthermore, the structural similarity between the native and mutant chitinase was checked by the RMSD value calculation using VMD software (Humphrey et al., 1996).

Experimental studies

Mutagenesis

The Quik Change Site Directed Mutagenesis strategy was used for mutagenesis. The plasmid pQE60 harboring the chitinase gene (previous work) (Babashpour et al., 2012; Zarei et al., 2012) was used as the template for producing the mutant chitinase. The forward (5’-CACATCTTCTGATGATCTACGACTTCTACG-3’) and reverse (5’
CGTAGAAGTCGTA
GAT
CATCAGG
AAGATGTG
-3')

ATAATCATA
tG
G
CGCAAATT

AATAACCC-3', NdeI site with ATG as
the start codon) and reverse primer
(Emr-R-chit: 5'-ATATACTCGAGTC
t
TTTAACGC
CGGCCG-3', XhoI site). Following, the
PCR product (1,701 bp) was cloned
into pTZ57R/T then recombinant
plasmid was digested with NdeI and
XhoI and the restricted segments were
isolated and ligated into the pET26b
eexpression vector which has been
digested with the same enzymes. A
single colony of recombinant plasmid
carrying the mutant chitinase which had
been transformed into BL21 (DE3)
Escherichia coli strain was cultured in
LB medium. Therefore protein
expression was induced by addition of 1
mM isopropyl-β-d-thiogalactoside
(IPTG) and was verified by SDS-PAGE
analysis. The protein content of the
extracts was determined by the

Enzyme activity
Chitinase activity was assayed by the
modified method of Roberts and
Selitrennikoff (Takiguchi, 1991) using
colloidal chitin (1% w/v chitin in 20
mM phosphate buffer, pH 7.5) as the
substrate. The enzymatic reaction
was run for 60 min at 50°C. To stop
the reaction, the mixture was boiled. Then,
DNS (1% w/v) was added and the
resulting mixture was maintained at
100°C for 10 min. The extent of the
enzymatic hydrolysis was evaluated
spectrophotometrically at 540 nm
(Miller, 1959).

Stability and kinetics of the native and
mutant recombinant chitinases
To determine the optimum pH, the
chitinases activity was measured at
various pH (s) ranging from 3–12 using
50 mM glycine–acetate–phosphate
buffer. The enzyme stability at various
pH(s), was determined by preincubation
of the enzyme at the desired pH in the
absence of the substrate for 90 min,
followed by the enzyme assay at
optimum pH.

To determine the optimum
temperature, the enzymatic reaction
was run in the presence of the substrate
at different temperatures between 10–
90°C in 20 mM phosphate buffer (pH 6)
for 45 min, followed by the enzyme
assay at pH and °C.

Thermal stability was measured by
two methods: (1) measurement of the
thermal stability at different
temperatures. To do so, the enzyme was
incubated at different temperatures

Cloning and expression
For reinforcement of the mutant
chitinase gene and insertion of
restriction sites in two sides of gene
were designed two oligonucleotide
primers, a forward primer (Emr-F-chit:
5'-

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ranging from 50 to 90 °C for 90 min, then colloidal chitin was added, and the enzyme assay was done at 50 °C (optimum temperature). (2) Measurement of the thermal stability at different times at two selected temperatures of 50 and 60 °C. Every 20 min (up to 120 min), the reaction mixture was sampled and the samples were placed on ice for 30 min, then colloidal chitin was added to assay the enzyme at 50 °C (optimum temperature).

Kinetics parameters of the chitinases were calculated by double-reciprocal analysis of the corresponding kinetics curves obtained from the corresponding enzymatic reactions in the presence of various concentrations of the substrate ranging from 0 to 50 mg mL⁻¹ at pH and °C (Lineweaver and Burk, 1934).

**Results**

**Homology modeling and structure validation**

Using *Serratia marcescens* B4A chitinases (PDB: HM473183) as the reference structure in the computational studies, it was found that Ser390 could be more important to the structural stability of HM473183 as compared with the other suggestions. Therefore, substitution of Ser390 with Ile was further studied. This substitution not only decreases the number of amino acids with polar residues, but it also increases the chance of the beta sheet extension in the protein skeleton since Ile, as a branched amino acid, has conformational preference for the beta sheet structure. As a result, the suggested substitution was anticipated to shorten the loop section (Fig. XX), where ser390 was placed, in order to enhance the structural stability through the reduction of the entropy of the whole molecule. To appraise the change in protein stability after having the substitution, the thermodynamic stability (ΔΔG) using flexible and fixed backbone method was measured by Eris server (Yin et al., 2007). This resulted in ΔΔG of 2.18 kcal mol⁻¹ for the mutant chitinase. Moreover, the appropriateness of the mutation site was verified by an automated web site: http://bioinformatics.org/primerx (Zheng et al., 2004).

Homology modeling was performed to predict the 3D structure (opened form) of the mutant. Based on the sequence identity between the query and the template protein sequence, the PDB number of 1edq_A with 99% identity was recognized for prognostication of the opened form of both chitinases. Then, the 3D structures of native and mutant chitinase were optimized using MODELLER v9.10 (Fig. 1). ProSA and QMEAN scores were found to be -9.75 and 0.837 (Z-score=0.88) for the mutant chitinase and -9.56 and 0.84 (Z-score=0.91) for native chitinase, respectively.
Ramachandran plot demonstrated that 93.7% of the amino acid residues of the mutant chitinase were in the most favored region and 6.3% in the additional allowed region. Superimposition of native and the mutant resulted in RMSD value of 2.9 Å for the Cα atoms. These results indicated that the structures of the mutant and template are similar and the homology model is reliable.

Mutagenesis and expression
After mutagenesis with the QCSDM strategy, the accuracy of mutation was verified by DNA sequencing. Then, the mutant chitinase gene was sub cloned in pET26b expression vector which resulted in successful expression of the enzyme in BL21 (DE3) Escherichia coli strain. The protein produced a 62 kDa band on SDS-PAGE (Fig. 2).

Kinetics parameters
Measurement of the Michaelis constant was showed that the $K_m$ values of native and mutant chitinases were 4/5 mg mL$^{-1}$ (Fig. 3a) and 4/7 mg mL$^{-1}$ (Fig. 3b), respectively. Very little different of the $K_m$ values of native and mutant
chitinase showing that the affinity of enzyme towards the substrate has not been changed. The $V_{\text{max}}$ value of native and mutant chitinase were observed to be $588/23 \text{ Unit (µM \text{min}^{-1})}$ of protein (Fig. 3a) and $153/84 \text{ Unit (µM/min)}$ of protein (Fig. 3b), respectively.

![Figure 3: Michaelis–Menten curve of native and mutant chitinase activity. Inset Lineweaver–Burk plot. a) The Km values and the Vmax value of native chitinase were found to be 4/5 mg mL$^{-1}$ and 588/23 Unit (µmol min$^{-1}$), respectively. b) The Km values and the Vmax value of mutant chitinase were found to be 4/7 mg/ml and 153/84 Unit (µmol min$^{-1}$) of protein, respectively.](image)

Effect of pH and Temperature
The optimal pH(s) for native and the mutant chitinases were found to be 6 and 5, (Fig. 4a), respectively. However, the enzymes were stable between pH range of 6-10 for native and 5-10 for the mutant chitinase (Fig. 4b). The optimal temperatures of 50°C and 45°C were obtained for native and the mutant, (Fig. 4c), respectively.

Thermal stability
Thermal stability of chitinases was measured at different temperatures. Although the activities of both chitinases were reduced with the rise of temperature, native chitinase activity was reduced faster and reached zero at 70°C, while the mutant chitinase could maintain about 20 percent of its activity (Fig. 5a). Measurement of temperature stability during the time showed that the native enzyme activity at 50 and 60°C was decreased, but the activity of mutant chitinase increased over the time (Fig. 5b).
Figure 4: Effects of pH and temperature on the activity and stability of the native and mutant chitinase. a) Effect of pH on enzymes activity, enzymes assays performed over a range of pH values (3–10) using colloidal chitin as substrate under standard assay conditions. The optimal pH for the chitinase activity of native and mutant chitinases was found to be 6 and 5 respectively. b) Effect of pH on enzyme stability, enzymes preincubated at various levels of pH (3–12) without the substrate for 90 min. Following adjustment of the enzymes to pH 6, colloidal chitin was added and then enzymes assays performed under standard assay conditions. The enzymes were stable between pH range of 6-10 for native and 5-10 for mutant chitinase. c) Effect of temperature on enzymes activity, enzymes assays performed in the range of 10–90 °C in pH 6 using colloidal chitin as substrate under standard assay conditions. The optimal temperature for enzyme activity of native and mutant was found to be 50 °C and 45 °C, respectively.

Figure 5: Thermal stability of chitinase. a) Effect of different temperatures on the activity and stability of native and mutant chitinase after incubation for 90 min. b) Effect of temperature on the stability of native and mutant chitinase after incubation at 50, 60°C in various time intervals.
**Discussion**

Research on the thermostable chitinases is attractive for finding efficient enzymes with more usage in industrial process (Roodi et al., 2017). Recent studies clearly exhibit that the kinetic stability of proteins might be better by rational design, but that it may be difficult to exactly determine which region and amino acid of the protein should be considered. There are a commonplace strategy for increasing protein thermal stability through shortening of exposed loop regions which leads to decreasing the entropy and elevating the free energy of protein unfolding (Thompson and Eisenberg, 1999). Furthermore other researchers showed that the proportion of beta sheet amino acids is meaningfully high in thermophiles. This result might be obtained through enhancing hydrogen bonding which leads to increased thermal stability (Chakravarty and Varadarajan, 2000). In this study we wanted to stabilize the chitinase by reduction of the loop length and increasing of beta sheet length altogether. For this purpose we determined amino acids that locate in the border of the loop and beta sheet, and considered that candidate sites did not have unfavorable steric interactions.

Also researchers showed that for increasing thermal stability, declining uncharged polar amino acids is very significant. The elimination of these residues may also reduce deamination of asparagine and glutamine by serine and threonine at high temperatures (Vogt and Argos, 1997; Haney et al., 1999). Serine due to its short side chain mostly involve in local interactions, but large side chains amino acids generally contribute in short and long range interactions (Hornby, 1993; Kumar et al., 2000). Thus we selected Serine 390 resides in the end of the loop and replaced it with Isoleucine residue because of several reasons: Isoleucine residue tends to place in beta sheet and this replacement causes the reduction of the loop length and enhancement of the beta sheet length. Furthermore, increase of protein hydrophobicity stabilizes thermophile proteins (Mozhaev and Martinek, 1984; Haney et al., 1999). Also entropy of unfolding dependent on the kind of the side chain, for example in this case flexibility of the selected loop is decreased by the presence of long or bulky side chains (Némethy et al., 1966). Also studies showed that polar uncharged residue in comparison of one non-polar is less favorable in thermostable protein (Mozhaev and Martinek, 1984; Chakravarty and Varadarajan, 2000). Our studies demonstrated that mutant chitinase has been compacted and required time and temperature for its activity. Native and mutant chitinase kinetic studies showed that mutant chitinase had more activity than the native type because the native chitinase could not tolerate the assay condition, but this condition is favorable for the mutant chitinase to increase its flexibility and exposed catalytic site for substrate. Because the enzyme was partially purified, in
chitinase activity assay two negative controls were used. The first negative control was cocktail without chitinase; the second negative control was cocktail with BL21 containing pET26b bacterial cell extract. Any enzymatic activity was observed in two negative controls. Alteration in enzyme binding site may lead to increase in $K_m$ (Fields and Somero, 1998; Fields, 2001). In our study the brief change of mutant chitinase $K_m$ showed that the enzyme substrate affinity was similar to the native enzyme. The $V_{\text{max}}$ of mutant chitinase is smaller than native chitinase because the calculation of the $V_{\text{max}}$ value was performed in routine condition while mutant chitinase was rigided and at this condition showed low activity. Reduction of mutant chitinase $V_{\text{max}}$ without change of $K_m$ was similar to the noncompetitive inhibition. Studies have shown that there is correlation between thermal stability and the optimum temperature (Danson et al., 1996; Daniel et al., 2001). Optimum temperature is increased in several engineered thermostabilized enzymes and doesn’t change in the others (Thomas and Scopes, 1998; Björk et al., 2003). In our research the optimum temperature and pH didn’t change remarkably for both the mutant and native chitinases, indicating that mutation did not affect the main characteristic of the enzyme. pH stability of the mutant and native chitinases were the same, but thermal stability was increased remarkably. Mutation caused the chitinase to tolerate high temperatures up to $90^\circ$C and it remained stable at $50^\circ$C, $60^\circ$C for up to 120 minutes. This characteristic is very important for industrial and biotechnological applications.

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