Growth and feed utilization of goldfish (*Carassius auratus*) fed graded levels of brewers yeast (*Saccharomyces cerevisiae*)

Gumus E.¹; Aydin B.²; Kanyilmaz M.²

Received: October 2014                      Accepted: March 2016

Abstract

In this study, a feeding trial was conducted to examine the potential of replacing fish meal with brewers yeast in practical diet of goldfish (*Carassius auratus*). Five isoproteic (37% CP) and isocaloric (3350 kcal/kg) diets were formulated to contain graded levels of brewers yeast. Fish meal protein was replaced by 0%, 15%, 25%, 35%, and 45% of yeast. Each diet was randomly allocated to triplicate groups of 20 fish (initial average weight of 0.56 g fish⁻¹) in glass aquarium (65L). Fish were fed three times per day to apparent satiation for 84 days. At the end of the experiment, weight gain, specific growth rate (SGR), feed conversion ratio (FCR), protein efficiency ratio (PER), condition factor (CF), survival rate (SR), hepatosomatic indices (HSI) and body composition of goldfish fry were determined. According to the results, weight gain, SGR, FCR and PER of fish fed the diet including yeast replaced 35% of the fish meal were better than those of fish fed the other diets. There were no significant differences in SR and HSI values among fish fed diets (*p* > 0.05). However, CF among fish fed the experimental diets was significantly differ (*p* > 0.05). Whole body composition was similar among fish fed different diets. The optimal replacement level of fishmeal protein by brewers yeast was determined by second-order polynomial regression to be (*y* = 2, 2237 - 0.0004*x*² + 0.0279*x*; *R*² = 0.9977) 34.875%, on the basis of SGR.

Keywords: *Saccharomyces cerevisiae*, *Carassius auratus*, Fish meal replacement, Growth, Feed utilization.

¹-Akdeniz University, Faculty of Fisheries, 07058 Antalya, Turkey
²-Mediterranean Fisheries Res Prod & Training Inst, Kepez Unit, 07192 Antalya, Turkey
*Corresponding author’s Email: egumus@akdeniz.edu.tr
Introduction
Ornamental fish has become increasingly important today all around the world. The ornamental fish trade in the world is to have a volume of US $1 billion annually (FAO, 2010). Ornamental fish sector has consisted of marine and freshwater ornamental fishery. One of the most important species of freshwater ornamental fishery is goldfish, *Carassius auratus*. It is a kind of fish that widely preferred by hobbyists because of species diversity, attractive and variety of colors, and high tolerance to environmental demands. The healthy and fast growing individuals for goldfish are very important to be met of market demand. In this reason, goldfish rearing is very important to feed diet with balanced energy/protein ratio and low price.

Fish meal as a major source of protein, being high nutritional value and taste are preferred in fish feed preparation. However, various researches are made as to the use of cheaper alternative protein sources to replace partially or completely of fish meal due to the expensive raw material and difficulties experienced in the production of fish meal (Tacon and Metian, 2009). One of alternative protein sources in aquafeeds is single cell protein micro algae, bacteria and yeast (Tacon, 1994). Among them, the yeasts are important species including *Candida* sp, *Hansenula* sp, *Pichia* sp. and *Saccharomyces* sp. Brewers yeast (*S. cerevisiae*) is a natural product from the brewing industry that contains rich in protein, various immunostimulating compounds such as B-glucans, nucleic acids as well as mannan oligosaccharides (Li and Gatlin, 2003; Ghosh *et al*., 2005; Abdel-Tawwab *et al*., 2008; 2010), and has been used as a diet additive for various animals. It has been observed to be capable of enhancing growth (Lara-Flores *et al*., 2003; Li and Gatlin, 2003; Ebrahim and Abou-Seif, 2008; Zerai *et al*., 2008; Korkmaz and Çakiroğulları, 2011; Guroy *et al*., 2012; Omar *et al*., 2012) and immune responses (Ortuno *et al*., 2002) of various fish species.

The studies on using brewers yeast for growth performance of goldfish have not been adequate. Therefore, the present study was to evaluate the effect of partial replacement of fish meal in the diet with brewers yeast (*Saccharomyces cerevisiae*) on growth, feed utilization and carcass composition of goldfish fingerlings.

Materials and methods
Diet preparation
Five experimental diets were formulated to 37% crude protein, 8% crude lipid and 3350 kcal/kg digestible energy with 0%, 15%, 25%, 35%, and 45% of brewers yeast (Table 1). Fishmeal and soybean meal were provided by the Abalioglu Feed-Soybean and Textile Industries, Inc., Denizli, Turkey. Brewer’s yeast was obtained from private beer producing Ltd. All ingredients were thoroughly mixed, and 400 mL of water was added per kg diet.
Table 1: Formulation and proximate analysis of the experimental diets (% dry matter basis) fed to goldfish (*Carassius auratus*) fry

<table>
<thead>
<tr>
<th>Ingredients (%)</th>
<th>0</th>
<th>15</th>
<th>25</th>
<th>35</th>
<th>45</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fish meal</td>
<td>40</td>
<td>34</td>
<td>30</td>
<td>26</td>
<td>22</td>
</tr>
<tr>
<td>Brewers yeast</td>
<td>0</td>
<td>8.87</td>
<td>14.78</td>
<td>20.71</td>
<td>26.63</td>
</tr>
<tr>
<td>Soybean meal</td>
<td>20</td>
<td>20</td>
<td>20</td>
<td>20</td>
<td>20</td>
</tr>
<tr>
<td>Corn starch</td>
<td>25.15</td>
<td>21.93</td>
<td>20.12</td>
<td>19.19</td>
<td>17.17</td>
</tr>
<tr>
<td>Fish oil</td>
<td>2.75</td>
<td>3.6</td>
<td>4</td>
<td>4.5</td>
<td>5</td>
</tr>
<tr>
<td>Vitamin premix</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>Mineral premix</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>Methionine</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>NaCl</td>
<td>0.1</td>
<td>0.1</td>
<td>0.1</td>
<td>0.1</td>
<td>0.1</td>
</tr>
<tr>
<td>CaHPO$_4$.2H$_2$O$_5$</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>CMC</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Cellulose</td>
<td>2.5</td>
<td>2</td>
<td>0.5</td>
<td>0.1</td>
<td>1</td>
</tr>
<tr>
<td>Total</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
</tr>
</tbody>
</table>

Proximate analysis (% dry weight basis)

<table>
<thead>
<tr>
<th>Components</th>
<th>0</th>
<th>15</th>
<th>25</th>
<th>35</th>
<th>45</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dry matter</td>
<td>94.99±0.04</td>
<td>95.09±0.01</td>
<td>95.01±0.01</td>
<td>95.05±0.06</td>
<td>95.00±0.01</td>
</tr>
<tr>
<td>Crude protein</td>
<td>37.31±0.05</td>
<td>37.44±0.03</td>
<td>37.39±0.05</td>
<td>37.37±0.03</td>
<td>37.49±0.05</td>
</tr>
<tr>
<td>Crude lipid</td>
<td>8.01±0.01</td>
<td>7.96±0.04</td>
<td>7.95±0.02</td>
<td>7.92±0.04</td>
<td>7.99±0.04</td>
</tr>
<tr>
<td>Crude ash</td>
<td>4.76±0.01</td>
<td>2.93±0.40</td>
<td>3.44±0.50</td>
<td>3.43±0.42</td>
<td>3.06±0.28</td>
</tr>
<tr>
<td>Energy (DE kcal/kg)$^8$</td>
<td>3443</td>
<td>3381</td>
<td>3336</td>
<td>3328</td>
<td>3285</td>
</tr>
</tbody>
</table>

$^1$Brewers yeast (*Saccharomyces cerevisiae*)

$^2$Vitamin premix (per mg/kg feed, except where indicated); Vitamin A Retinol, 4 000 000 IU; Vitamin D$_3$, 600 000 IU; Vitamin E Tocopherol, 40 000; Vitamin K$_1$, Menadione, 2 400, Vitamin B1, 5 000; Vitamin B$_2$, 8 000; Vitamin B$_6$, 4 000; Vitamin B$_12$, 12, Vitamin C, 40 000; Niacin, 50 000; Folic acid, 1400; D-calcium Pantothenate, 8 000; D-Biotin, 50; and Inositol, 40 000.

$^3$Mineral premix (per mg/kg feed); MnSO$_4$, 60 000; FeSO$_4$, 10 000; ZnSO$_4$, 75 000; CuSO$_4$, 5 000; CoSO$_4$, 1 000; KIO$_3$, 2 500; NaSeO$_3$, 100; and MgSO$_4$, 65 000.

$^4$Natrium chloride, $^5$Calcium hydrogen phosphate, $^6$Carboxi-metil cellulose. $^7$Values are mean (±SE) of three replicate. $^8$Digestible energy calculated as protein 4.9 kcal g$^{-1}$, lipid 9.01 kcal g$^{-1}$ and carbohydrate 3.49 kcal g$^{-1}$ (NRC, 1993).

Then the mixture of each diet was separately passed through a meat grinder, and pelleted through 2-mm-diameter diet. The pellets were dried in a drying oven for 24 hours at 50°C, crushed into desirable particle sizes, then sealed in bags and stored in frozen at -20°C prior to use in the feeding experiment. The dietary composition and chemical analyses of the experimental diets are shown in Table 1.

**Fish and Experimental conditions**

This study was conducted from March 2 through May 25, 2012 at Aquaculture Experiment Unit, Akdeniz University, Antalya. Goldfish fry were obtained from local ornamental fish farm in Antalya Province. After two weeks acclimation, fish were randomly distributed at 20 fish with initial mean weight 0.56±0.01g to each 65L glass aquarium. Each treatment was run in triplicate. A 100g sample of fish was frozen at -20°C for initial proximate analysis. Each aquarium was supplied with compressed air by air stone using...
central air pump. Half of the aquarium water was siphoned daily for removing fish wastes, and water volume was replaced by aerated water from the storage tank. A 12h light: 12h dark photoperiod was maintained with fluorescent lights controlled by timers, and light intensity at the water surface of the aquaria was kept constant as 150 lux. Fish in all treatments were hand-fed to visual satiation three times a day at 9:00, 13:00 and 16:00 and feed intake was recorded daily for 12 weeks. Dead fish were recorded daily and removed from each aquarium. Water quality parameters were monitored and maintained during the experiment. Dissolved oxygen and temperature were measured on site using a WTW multimeter. pH was measured using a pH-meter (WTW, Wissenschaftlich-Weilheim, Germany).

**Growth parameters**

At the end of the experiment, before weighing, fish were anesthetized using clove oil (v:v, 1/20) to reduce their stress. Fish were individually weighed and measured. Five fish from each aquarium were dissected, and the liver and viscera weighed. Fish growth parameters and feed utilization were calculated as follows (Gümüş et al., 2013):

- Weight gain (WG, g)=Wf-Wi; Specific growth rate (SGR, %/day)=100×(LnWf-LnWi)/days,
- where Wi and Wf are the initial and final weights (g), respectively.
- Feed conversion ratio (FCR)=Feed intake (g)/WG;
- Protein efficiency ratio (PER)=WG/Protein intake (g);
- Condition Factor (CF)=100×[(final fish weight (g)/final fish length (cm)³];
- Hepatosomatic indices (HSI, %)=100×(liver weight (g)/Wf (g));
- Survival rate (SR, %)= 100 × (Final number of fish/Initial number of fish).

**Chemical analysis**

Proximate composition of experimental diets and fish samples were analyzed according to standard methods of Association of Official Analytical Chemists (AOAC, 1995). Dry matter was estimated by drying samples to constant weight at 105°C in a drying oven. Crude protein determined by Kjeldahl method (N×6.25) after acid digestion, crude lipid by ether extraction using Soxtec system, and ash by incineration at 550°C for 24 h.

**Statistical analysis**

Data were subjected to one-way analysis of variance (ANOVA). Differences between means were tested at the 5% probability level using Duncan’s multiple-range test. Results were presented as mean±SE. All statistical analyses were performed using SPSS software 15.0 (SPSS Inc., Chicago, IL, USA). A second-order polynomial regression analysis method was conducted to analyze the relationship between SGR and the replacement levels of fish meal protein by brewers yeast in the diets of goldfish.
Results
The parameters for water quality throughout feeding experiment were recorded for temperature were in the range of 24-28°C, dissolved oxygen from 5.00 to 6.23 mg/L, and pH from 7.71 to 7.89. Water quality parameters were maintained within the acceptable ranges for goldfish (Lochmann and Phillips, 2002).

The growth performance, feed efficiency and survival rate of goldfish fed the experimental diets are shown in Table 2. Goldfish fed the diets supplemented with brewers yeast tended to have better growth performance during the 12-week feeding experiment. Final fish weight, weight gain and specific growth rate for fish fed 35% brewers yeast were higher than those of the other groups (p<0.05). The relationship between dietary brewers yeast levels and the SGR was the best described by the second-order polynomial regression equation as follows: \( Y = 2.2237 - 0.0004x^2 + 0.0279x \). According to differential equation, maximum SGR occurred at brewer yeast level of approximately 34.875% (Fig. 1).

Feed conversion ratio and protein efficiency ratio were improved by increasing dietary levels up to 35% brewers yeast. However, condition factor was tended to decrease in fish fed diets inclusion graded levels of brewers yeast (p<0.05).

Hepatosomatic index and survival rates were not significantly different among the different treatments. No significant differences also were found in whole-body dry matter, protein, lipid or ash contents of fish fed the different experimental diets (p>0.05) (Table 3).

Table 2: Growth and feed utilization of goldfish (*Carassius auratus*) fed with experimental diets.

<table>
<thead>
<tr>
<th>Parameters (% )</th>
<th>0</th>
<th>15</th>
<th>25</th>
<th>35</th>
<th>45</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial body weight (g)</td>
<td>0.56±0.00</td>
<td>0.56±0.00</td>
<td>0.56±0.00</td>
<td>0.56±0.00</td>
<td>0.56±0.00</td>
</tr>
<tr>
<td>Final body weight (g)</td>
<td>3.64±0.10a</td>
<td>4.79±0.12b</td>
<td>5.25±0.34ab</td>
<td>5.51±0.16a</td>
<td>5.26±0.16ab</td>
</tr>
<tr>
<td>Weight gain (g/fish)</td>
<td>3.08±0.10a</td>
<td>4.24±0.12ab</td>
<td>4.69±0.34ab</td>
<td>4.96±0.16a</td>
<td>4.70±0.16ab</td>
</tr>
<tr>
<td>SGR (% day(^{-1}))</td>
<td>2.22±0.20a</td>
<td>2.55±0.03b</td>
<td>2.65±0.07ab</td>
<td>2.72±0.03a</td>
<td>2.66±0.03ab</td>
</tr>
<tr>
<td>FCR</td>
<td>3.32±0.18a</td>
<td>2.68±0.03b</td>
<td>2.42±0.18ab</td>
<td>2.26±0.03a</td>
<td>2.34±0.04ab</td>
</tr>
<tr>
<td>PER</td>
<td>0.68±0.04a</td>
<td>0.86±0.01b</td>
<td>0.97±0.08ab</td>
<td>1.04±0.01a</td>
<td>1.01±0.01ab</td>
</tr>
<tr>
<td>CF</td>
<td>1.79±0.00a</td>
<td>1.83±0.01a</td>
<td>1.65±0.10ab</td>
<td>1.67±0.03ab</td>
<td>1.54±0.05b</td>
</tr>
<tr>
<td>HSI (%)</td>
<td>2.98±0.01</td>
<td>3.18±0.42</td>
<td>2.85±0.13</td>
<td>2.44±0.30</td>
<td>2.96±0.41</td>
</tr>
<tr>
<td>Survival (%)</td>
<td>95.00</td>
<td>96.67</td>
<td>96.67</td>
<td>96.67</td>
<td>98.33</td>
</tr>
</tbody>
</table>

Values are mean (±SE) of three replicate. Values with the same superscripts within the same row are not significantly different (p>0.05).
Table 3: Chemical composition of goldfish (Carassius auratus) fed with experimental diets.

<table>
<thead>
<tr>
<th>Parameters (%</th>
<th>% yeast replaced by FM</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0</td>
</tr>
<tr>
<td>Crude protein</td>
<td>15.32±0.04</td>
</tr>
<tr>
<td>Crude lipid</td>
<td>5.48±0.05</td>
</tr>
<tr>
<td>Crude ash</td>
<td>3.34±0.31</td>
</tr>
<tr>
<td>Dry matter</td>
<td>24.72±0.44</td>
</tr>
</tbody>
</table>

Values are mean (±SE) of three replicate.

Discussion
The present study indicated that brewers yeast was replaced up to 35% of fish meal protein in the diets and improving growth and feed utilization. However, inclusion of brewers yeast above 35% fish meal protein replacement resulted in a slightly decrease in growth performance of goldfish.

The nutritional value of brewers yeast has been tested to affect on growth performance and feed utilization in many fish species such as Nile tilapia (Lara-Flores et al., 2003; Abdel-Tawwab et al., 2008; Korkmaz and Cakirogullari, 2011; Asadi Rad et al., 2012), rohu (Tewary and Patra, 2011), rainbow trout (Rumsey et al., 1991; Guroy et al., 2012), carp (Omar et al., 2012), red drum (Li and Gatlin, 2005), beluga (Hoseinifar et al., 2011), and sea bass (Oliva-Teles and Goncalves, 2001). The results were obtained that supplementation with 25% brewers yeast for rainbow trout (Rumsey et al., 1991), 50% brewers yeast for sea bass, Dicentrarchus labrax (Oliva-Teles and Goncalves, 2001), 30% yeast for koi carp (Korkmaz and Cakirogullari, 2011), 27-41% bio-ethanol yeast for sunshine, M. chrysops × M. saxatalis,
(Gause and Trushneski, 2011), 50% 
yeast for carp (Omar et al., 2012), 30% 
organically certifiable yeast for rainbow 
trout (Guroy et al., 2012), and 40% 
brewers yeast for Atlantic salmon 
(Overland et al., 2013). Similar pattern 
that improved growth performance in 
tilapia fed yeast diets have been 
reported by Olvera-Novoa et al. (2002), 
Lara-Flores et al. (2003), Li and Gatlin 
(2005), Abdel-Tawwab et al. (2008), 
Zerai et al. (2008), Osman et al. (2010), 
and Ozorio et al. (2012).

Polynomial regression has been 
recommended for describing and 
approximating the relationship between 
SGR and dietary brewers yeast levels. 
In the present study, the second-order 
polynomial regression analysis has been 
suggested that the optimal replac 
ment level of fish meal protein was about 
34.875% (Fig. 1). Based on this 
analysis, no significant improvements 
of weight gain or SGR were observed in 
goldfish fed higher concentration of the 
brewers yeast supplemented in the 
diets.

The better FCR and the higher PER 
values were observed in fish that fed up 
to 35% brewers yeast supplemented 
diets which improved feed utilization of 
goldfish. Oliva-Teles and Gonçalves 
(2001) found better feed efficiency 
when 30% of the fish meal protein was 
replaced with brewers yeast for sea 
bass. At the end of the experiment, fish 
survival was high and ranged from 93% 
to 98% without significant difference 
among treatments (p>0.05). Also no 
significant differences were observed in 
condition factor and HSI values of 
goldfish fed the experimental diets.

Whole-body dry matter, crude 
protein, crude lipid, and ash content 
were not affected by the different 
dietary treatments (Table 3). These 
results took the same trend of those 
results obtained by Olvera-Novoa et al. 
(2002), that they observed no 
differences in carcass composition 
when substituting fish meal protein with 
torula yeast in diet for tilapia fry. 
Ebrahim and Abou-Seif (2008) reported 
that carcass composition was not 
affected by dietary brewers yeast 
(p>0.05). Moreover, Guroy et al. (2012) 
reported that use of organically certified 
yeast did not change the whole-body 
contents of rainbow trout. According to 
the Gaese and Trushenski (2011) there 
were no significant differences in 
proximate composition, but lipid 
composition demonstrated decrease fish 
fed with all yeast supplemented diets. 
Oliva-Teles and Gonçalves (2001) and 
Hunt et al. (2013) reported that protein 
content which was significantly higher 
in fish fed diets containing yeast than 
the control diet. In contrast with our 
findings, rainbow trout fed the diets 
containing yeast showed decreased 
whole body protein lipid when 
compared with control diet (Hunt et al., 2013).

In conclusion, this study suggested 
that brewers yeast could replace 35% of 
fish meal protein in the diet of goldfish 
without any adverse effect on growth 
performance and feed utilization, but an 
optimal replacement level of 34.875% 
is suggested.
Acknowledgements
The authors would like to thanks M. Emin Akdoğan and Gizem Ülker for their helps throughout the period of the study.

References


Gümüş, E., Erdoğan, F. and Aydin, B., 2013. Evaluation of skate meal as a replacement of fishmeal in diets for nile tilapia fry (Oreochromis niloticus). The Israeli Journal of
Aquaculture - Bamidgeh, IJA_65.2013.922, 9P.


Olvera-Novoa, M.A., Martinez-Palacios, A.A. and Olvera-Castillo, L., 2002. Utilization of torula yeast (Candida utilis) as a protein source in diets for tilapia...
Gumus et al., Growth and feed utilization of goldfish (*Carassius auratus*) fed graded levels of *Oreochromis niloticus* peters fry. *Aquaculture Nutrition*, 8, 257-264.


